

Insights into Bacteriotherapy: Harnessing Bacteria for the Greater Good

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ABSTRACT

Bacteriotherapy, the process of using bacteria or bacterial products for therapeutic purposes, is gaining more popularity over the years. It has been used for the prevention and treatment of a wide variety of ailments and has been used as either the primary mode of therapy or in conjunction with other therapies. Bacteriotherapy has employed live bacteria, bacterial products, probiotics, prebiotics, synbiotics, and engineered organisms. There have been significant advancements over the years but there are still challenges that need to be addressed. In this article, we review bacteriotherapy in the context of infectious diseases, inflammatory diseases, and cancers.

Keywords: Bacteriotherapy; Probiotics; Prebiotics; Synbiotics; Fecal Microbiota Transplantation

Abbreviations: FMT: Fecal Microbiota Transplantation; IBD: Inflammatory Bowel Disease; CDIs: *C. difficile* Infections; UTIs: Urinary Tract Infections; TURBT: Transurethral Resection of Bladder Tumor; BCG: Bacillus Calmette-Guerin

Introduction

The concept of using bacteria to treat ailments is ancient. There are records that fecal slurries in the form of 'yellow soup' were consumed in the 4th century for treating food poisoning and diarrhea (Stripling C, et al. [1]). Since then, many advancements have been made to use bacteria and bacterial products as therapeutics, referred to as bacteriotherapy, for treating a variety of ailments (Huovinen,

et al. [2]). Bacteriotherapy has been successfully employed for the prevention and treatment of different conditions (Otto [3]). From using live naturally occurring bacteria, bacterial compounds and metabolites, to synthetic compounds and engineered bacteria, bacteriotherapy has come a long way (Figure 1). In the current minireview, we will summarize different bacteriotherapy-based strategies that have been used for the prevention or treatment of infectious diseases, inflammatory diseases, and cancer.

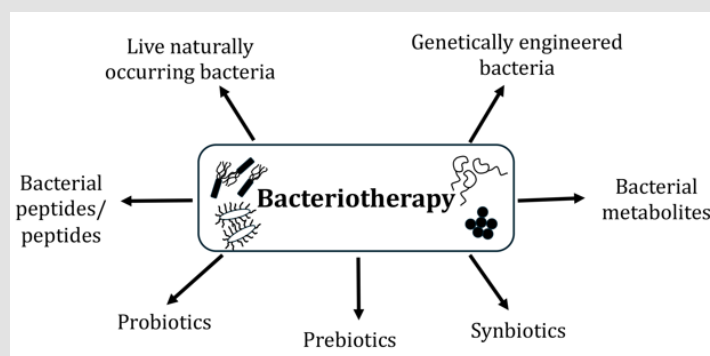


Figure 1: Bacteriotherapy involves the use of bacteria and their products for different therapeutic purposes. The image created by J. K. Iyer is under the CC0 license.

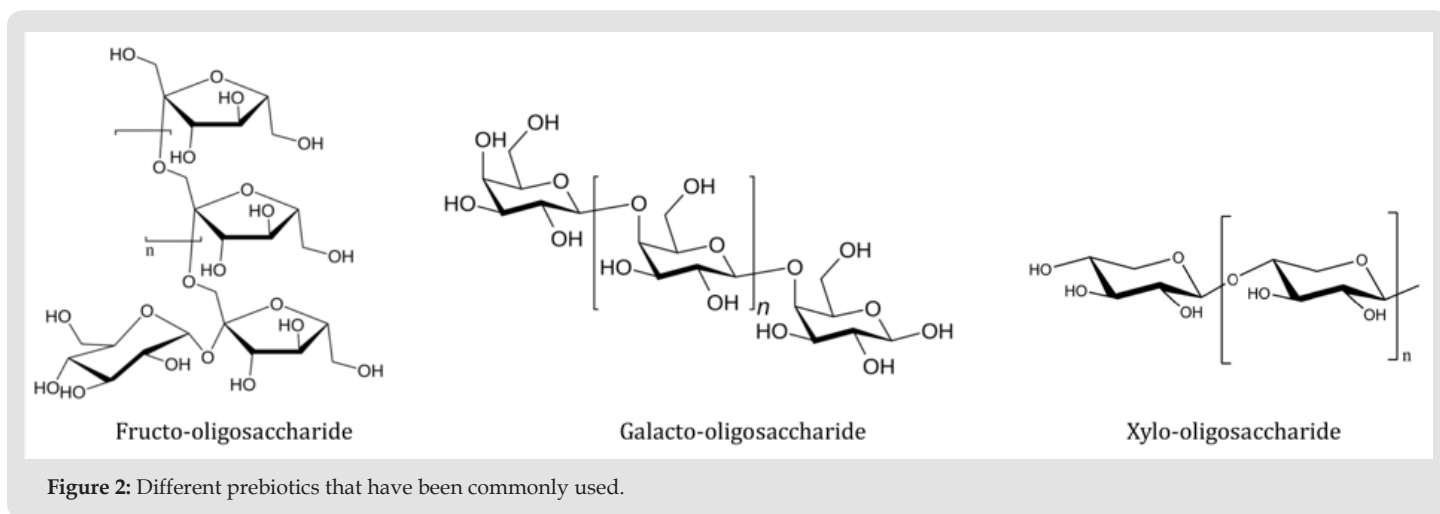
Bacteriotherapy and Infectious Diseases

Bacteriotherapy has been used to treat different types of diseases caused by pathogens. It is most popularly known for its therapeutic potential in treating gastrointestinal infections caused by *Clostridioides difficile* (*C. difficile*). *C. difficile* infections (CDIs) are treated with antibiotics and immunotherapy but bacteriotherapy has also been proven to be an effective strategy, especially for recurrent infections caused by *C. difficile* [Al Jashaami, et al. [4,5]]. CDIs are associated with dysbiosis of the gut microbiome due to factors such as antibiotic use, genetic predisposition, weakened immune systems, and others. Fecal microbiota transplantation (FMT), also known as fecal bacteriotherapy, involves addressing this dysbiosis by introducing normal fecal microflora associated with healthy individuals into a patient [Bakken, et al. [6,7]]. Fecal transplants are effective against primary and recurrent CDIs but are used more commonly to treat recurrent CDIs [Kaakoush, et al. [8]]. Fecal microflora can be introduced in the form of feces from healthy donors or mixtures of specific bacterial strains that are found in the gut and can be administered through enemas, colonoscopy, or oral capsules [Rode, et al. [9-11]]. Due to the efficacy of FMT and relatively low risks associated with the therapy, the FDA (Food and Drug Administration) has approved microbiome-based therapies for the treatment of CDIs [Cold, et al. [11,12]].

In addition to FMT, probiotics and prebiotics also show promise in the treatment of infectious diseases, especially those associated with the gastrointestinal and urogenital tract [Li, et al. [13,14]]. Probiotics are live microorganisms that provide health benefits when administered in adequate amounts. The most commonly used probiotics to treat gastrointestinal ailments belong to the genera *Lactobacillus* and *Bifidobacterium* [Patel, et al. [14]]. When *Lactobacillus rhamnosus* was administered to children diagnosed with rotaviral infections, there was a significant decrease in the duration of diarrhea [Szyman'ski, et al. [15]]. Probiotics can also be used to prevent and treat diarrhea caused by other agents in children and adults [Newberry, et al. [16,17]]. Different systematic reviews have demonstrated that when probiotics are included in standard therapies, there is higher eradication of *Helicobacter pylori* along with lower adverse events [Mestre, et al. [18-20]]. In addition to gastrointestinal infections, probiotics have also shown promise in the treatment and prevention of respiratory tract infections [Bellussi, et al. [21]]. In clinical trials as well as in vivo animal models, probiotics that include different species of *Lactobacillus* and *Bifidobacterium* have aided in the reduction of incidence and duration of respiratory tract infections caused by dif-

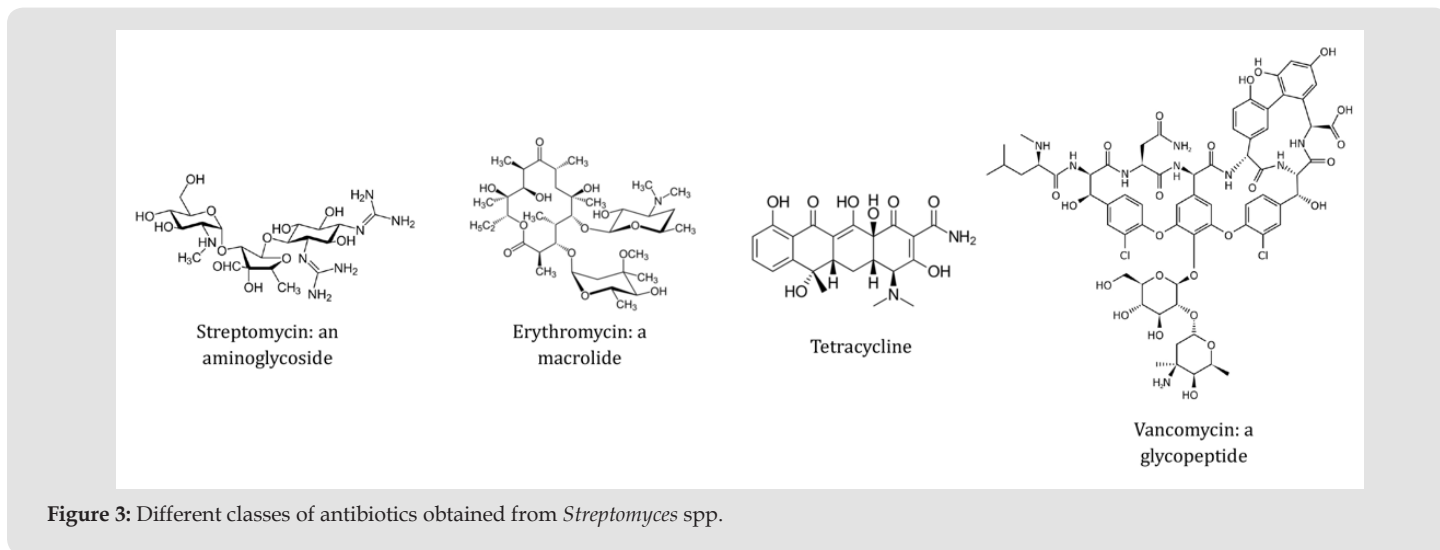
ferent viruses and bacteria [Darbandi, et al. [22,23]]. Probiotics have also been used in the prevention and treatment of urogenital diseases, including urinary tract infections (UTIs) [Kenneally, et al. [24]]. UTIs are commonly caused by bacteria and routinely treated with antibiotics [Flores-Mireles, et al. [25,26]]. This has resulted in a dramatic increase of antimicrobial resistance among uropathogens and the need for different treatment strategies [Gupta, et al. [27,28]]. Probiotics can directly influence the vaginal flora when administered orally or in the form of vaginal suppositories [Patel, et al. [14]]. A non-inferiority study comparing orally administered probiotic treatment with a trimethoprim-sulfamethoxazole treatment regimen in women with recurrent UTIs demonstrated that, while probiotic treatment did not meet the non-inferiority criteria in the prevention of UTIs, the probiotic group did not show an increase in antibiotic resistance [Beerepoot, et al. [29]]. A recent study compared the effect of orally and vaginally administered probiotics on the prevention of recurrent UTIs and found that prophylactic supplementation of vaginal probiotics in the presence or absence of oral probiotics resulted in significantly lesser incidences of UTIs [Gupta, et al. [30]].

In addition to probiotics, prebiotics and synbiotics can also aid in the prevention or treatment of some infectious diseases [Markowiak, et al. [31]]. Prebiotics are non-digestible oligosaccharides (Figure 2) that can stimulate the growth of selective and beneficial gut bacteria, thus promoting gut microbiota homeostasis [You, et al. [32]]. Most prebiotics are oligosaccharides like fructo-oligosaccharides, galacto-oligosaccharides, polydextrose, etc., and are fermented to form short chain fatty acids [Davani-Davari, et al. [33]]. These short chain fatty acids influence different physiological processes in the body and improve immune responses [Fusco, et al. [34]]. Prebiotic administration of fructo-oligosaccharides in patients with CDIs resulted in lesser incidence of relapse in diarrhea (34% in placebo vs 8.3% in treatment group) demonstrating the therapeutic effect of the prebiotic [Lewis, et al. [35]]. Prebiotics are not effective in the treatment of other infections by themselves [van Stuijvenberg, et al. [36]] but in combination with probiotics, the resulting synbiotics are usually more effective. When infants were provided with a mixture of *Lactobacillus rhamnosus* GG and LC705, *Bifidobacterium breve* Bb99, and *Propionibacterium freudenreichii* ssp *shermanii* along with galacto-oligosaccharide for 6 months, they had fewer respiratory tract infections than the placebo group [Kukkonen, et al. [37]]. As more clinical trials are performed, the effects of probiotics, prebiotics, and synbiotics on the prevention and treatment of infectious diseases will become clearer.



The use of antimicrobial agents, such as antibiotics, is also a form of bacteriotherapy because many antibiotics are metabolites obtained from bacteria. Since Sir Alexander Fleming discovered penicillin from the fungus *Penicillium* in 1928, there have been many classes of antibiotics that have been identified and made available as therapeutics [Hutchings, et al. [38]]. Many classes of antibiotics were isolated from actinomycetes, especially from the genus *Streptomyces* [de Lima Procopio, et al. [39]]. In fact, antibiotics obtained from *Streptomyces* are some of the most widely prescribed therapeutics to treat bacterial infections [Quinn, et al. [40]]. Some common classes of antibiotics obtained from *Streptomyces* (Figure 3) include aminoglycosides, macrolides, glycopeptides, and tetracyclines [Alam, et al. [41]]. Due to an increase in antimicrobial resistance among pathogens, there

is an urgent need for finding novel antimicrobial agents and other therapeutics [De Kraker, et al. [42]]. The novel antibiotics can be in the form of newly discovered molecules or chemical modifications of existing molecules to increase their potency and efficacy. By using innovative culturing methods like iChip technology, researchers can identify antibiotics produced by bacteria that are not easy to culture using traditional methods [Chabib, et al. [43,44]]. iChip technology has led to the discovery of novel antibiotics such as teixobactin and clovibactin among others [Chabib, et al. [43-46]]. Chemical synthesis approaches that aid in changing the structure of existing antibiotics has led to the creation of antibiotics like iboxamycin and cresomycin that display broad spectrum antimicrobial properties [Brodiazhenko, et al. [47-49]].



In addition to antibiotics, bacteria can make antimicrobial compounds that negatively affect growth of other bacteria (Danquah, et al. [50]). These compounds include bacteriocins, rhamnolipids, siderophores, and polyketides, which can be used for treatment of infections themselves or in conjunction with other therapeutics (Figure 4). Bacteriocins are ribosomally synthesized antimicrobial peptides produced by Gram-positive and Gram-negative bacteria (Darbandi, et al. [51]). Bacteriocins produced by lactic acid bacteria like *Lactococcus lactis* and *Bacillus* species can inhibit the growth of different types of bacteria including those that are multidrug resistant (Aunpad, et al. [52,53]). Rhamnolipids are another class of antimicrobial molecules that are produced by different bacteria and the rhamnolipids synthesized by *Pseudomonas* and *Burkholderia* species have been well characterized (Kumar, et al. [54,55]). These molecules are glycosurfactants that can disrupt bacterial cell membranes (Herzog, et al. [56]). Rhamnolipids isolated from *Pseudomonas aeruginosa* have demonstrated antimicrobial activity towards Gram-positive organisms like *Staphylococcus aureus* and *Enterococcus faecium*, Gram-negative organisms like *Acinetobacter baumannii*, and viruses like respi-

ratory syncytial virus and coronavirus (Cerqueira dos Santos, et al. [57,58]). Siderophores are antimicrobial molecules that aid with iron acquisition and are produced by Gram-positive and Gram-negative bacteria in iron deficient conditions (Kramer, et al. [59]). Some siderophores inhibit the growth of pathogenic protists like *Plasmodium falciparum* and *Trypanosoma cruzi* while some are being tested to be used as 'Trojan horses' in the effort to make drug delivery more efficient (Khasheii, et al. [60]). By linking antimicrobial compounds with siderophores, these compounds can be introduced into the pathogenic bacteria when it transports the siderophore and thereby interfere with cellular processes (Peukert, et al. [61]).

In addition to the antimicrobial molecules mentioned above, there are other naturally occurring molecules that are used as therapeutics. There is also a lot of interest in designing novel molecules to treat infectious diseases. For this purpose, researchers are using more and more in silico methods such as quantitative Structure-Activity Relationship analysis, molecular docking, and structure-based virtual screening, to design novel molecules to treat infectious diseases (Filipić, et al. [62]).

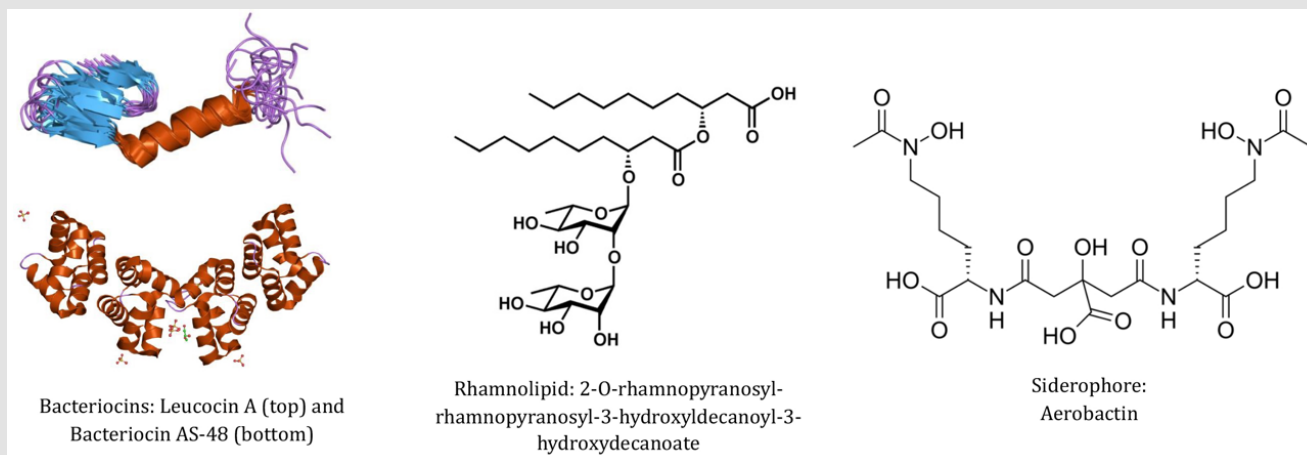


Figure 4: Examples of bacteriocins, rhamnolipids, and siderophores that exhibit antibacterial properties.

Bacteriotherapy and Inflammatory Diseases

Bacteriotherapy has been successfully used in the treatment of different diseases that arise due to inflammatory responses including inflammatory bowel disease, cystic fibrosis, and dermatitis. Inflammatory bowel disease (IBD), which comprises Crohn's disease and ulcerative colitis, is caused by different factors such as genetic predisposition, aberrant immune responses, and gut microbiome dysbiosis. It can be treated by different bacteriotherapeutics like antibiotics, probiotics, synbiotics, and fecal transplants (Hu, et al. [63]). The efficacy of the antibiotics, ciprofloxacin, metronidazole, rifaximin, and clarithromycin, have been tested in different randomized

clinical trials and rifaximin was found to be the most effective among them (Nitzan, et al. [64]). In one study, patients with moderately active Crohn's disease who received 800 mg of an extended intestinal release rifaximin twice a day over a span of 12 weeks, showed 62% remission compared to 43% who received the placebo (Prantera, et al. [65]).

Probiotic therapy has also shown promise in the treatment of IBD. Patients with mild to moderate ulcerative colitis showed a significant decrease in their ulcerative colitis disease activity index and Rachmilewitz endoscopic index when they were treated with *Bifidobacterium longum* 536 (Tamaki, et al. [66]). Another study demonstrated

that 42.7% of patients that were provided with VSL#3, a probiotic preparation comprising of four *Lactobacillus* species, three *Bifidobacterium* species, and *Streptococcus thermophilus*, achieved remission compared to 15.7% of patients who were given the placebo (Sood, et al. [67]). There is not much evidence to support that prebiotics can be used for the treatment of IBD but synbiotics-based therapy has shown to alleviate symptoms in ulcerative colitis patients and improvement in clinical activity (Dhaneshwar, et al. [68]). In addition to antibiotics, probiotics and synbiotics, FMT has also been used in the treatment of IBD (Hu, et al. [63]). A study that involved patients with refractory Crohn's disease found that 86.7% of the patients showed clinical improvement and 76.7% of the patients showed remission upon FMT treatment (Cui, et al. [69]). Similarly, 87.5% patients with ulcerative colitis achieved a clinical response to FMT and had reduced levels of different cytokines in the serum, indicating that immune responses can be modulated by FMT (Wang, et al. [70]).

In addition to IBD, bacteriotherapy has been utilized to treat skin conditions like atopic dermatitis. A phase 1 randomized clinical trial involving patients with atopic dermatitis caused by *Staphylococcus aureus* showed that topical application of *Staphylococcus hominis* A9 resulted in fewer adverse events associated with atopic dermatitis (Nakatsuji, et al. [71]). In addition to *Staphylococcus hominis*, other bacteria such as *Roseomonas mucosa* have shown therapeutic activity against atopic dermatitis (Myles, et al. [72]). A meta-analysis of studies involving the use of probiotics for the treatment of atopic dermatitis found that probiotics decreased the Scoring Atopic Dermatitis (SCORAD) and improved quality of life (Umborowati, et al. [73]) in patients. More clinical studies need to be performed to gain more insight into the molecular mechanisms of action of bacteriotherapy in inflammatory diseases.

Bacteriotherapy and Cancer

Cancers are normally treated using multi-pronged approaches like chemotherapy, surgery, and radiotherapy. Each one of these has significant risks and adverse side effects associated with it; hence, more alternative and targeted therapies are needed. The concept of using bacteria to treat cancer was used by William B. Covey, a bone sarcoma surgeon, who used a mixture of heat-killed *Streptococcus pyogenes* and *Serratia marcescens* to treat malignant tumors (Nguyen, et al. [74,75]). Since then, different bacteria and bacterial products have been tested for their anti-cancer potential. Studies have demonstrated that colonization of instilled bacteria in the tumor environment interferes with tumor metabolic and proliferation characteristics and promotes host immune responses towards the cancerous cells in the tumor (Soleimanpour, et al. [76]). There have been reports that some bacterial strains belonging to the genera of *Escherichia*, *Salmonella*, *Bifidobacterium*, *Clostridioides* etc., can kill tumor cells by colonizing in hypoxic regions of the tumor (Kang, et al. [77,78]). They show anti-tumor activities in animal models but more clinical trials are needed to evaluate their efficacy in humans. A study involving

administering a *Pseudomonas aeruginosa* preparation (PAP) in conjunction with chemotherapy in patients with advanced non-small-cell lung cancer found that these patients had a higher objective response rate of 46.88% compared to 23.53% of patients who received only the chemotherapeutic (Chang, et al. [79]).

A bacteriotherapy strategy approved by the FDA involves the use of Bacillus Calmette-Guerin (BCG) for the treatment of non-muscle invasive bladder cancer. BCG utilizes a strain of *Mycobacterium bovis*, which was isolated from a cow infected with bovine tuberculosis. The strain was continuously cultured from 1908 to 1921 before it was weakened enough to be used in humans (Mukherjee, et al. [80]). BCG is delivered intravesically and relies on the host's own immune responses to eliminate remaining bladder cancer cells after a surgical procedure involving transurethral resection of the bladder tumor, also known as TURBT (Jiang, et al. [81]). BCG is equally or more effective than some chemotherapeutics. According to a study by Lamm DL, the probability of being disease-free at five years' survival among patients with carcinoma in situ (CIS) was 45% after treatment with BCG compared to 18% percent following treatment with doxorubicin (Lamm, et al. [82]). Delaying BCG treatment for more than 15 weeks post TURBT as compared to 6 weeks, increases the risk of recurrence by 50% and disease progression by 200% for every week of delay (Krajewski, et al. [83]). Another study found that patients with less than 6 instillations of BCG showed a recurrence rate of 59% compared to 13% shown by patients who had ~18 instillations of BCG (Alhogbani, et al. [84]). BCG therapy aids in activating immune responses that result in killing cancer cells. A study using a rat model of bladder cancer found that BCG therapy resulted in a transient increase of helper T-cells in the urothelial lining of the bladder (Kates, et al. [85]). A recent study demonstrated that BCG therapy resulted in the secretion of Fms-related receptor tyrosine kinase 3 ligand (FLT3LG) that can activate cytotoxic T-cells (Zhang, et al. [86]). Thus, BCG therapy is an effective therapeutic for the treatment of bladder cancer. While BCG has its benefits, it can also cause adverse side effects in some patients and hence patients should be monitored closely.

In addition to bacteria, bacterial toxins have been investigated as therapeutic agents for treatment of cancers in different cell lines and animal models (Trivanovic', et al. [87,88]). A study from Michl et al., showed that the *Clostridium perfringens* enterotoxin promoted necrosis in tumor cells along with significant reduction of tumor growth in a mouse xenograft model (Michl, et al. [89]). Studies with botulinum toxin demonstrated that it improved radio- and chemotherapy outcomes in a mouse model by increasing tumor oxygenation and perfusion and is being considered for other cancer-related therapies (Ansiaux, et al. [90,91]). There is also interest in conjugating bacterial toxins to cell proteins that allow the toxins to be targeted to cancers. Many different conjugates have been made with diphtheria toxin, and the conjugate of IL-2 and diphtheria toxin is used for treatment of patients with T-cell lymphoma (Shafiee, et al. [92]).

Currently, genetically engineered bacteria that can express different types of genes that aid in the identification and/or destruction of cancer cells have been created (Nguyen, et al. [74]). These genes can code for a variety of molecules including cytotoxic proteins, tumor specific antigens, immunomodulators, and small interfering RNA (Nguyen, et al. [74,75]). A study that used engineered attenuated *Salmonella typhimurium* expressing the flagellin of *Vibrio vulnificus* demonstrated increased infiltration of immune cells and activation of intratumoral macrophages in tumors generated in mouse models (Zheng, et al. [93]). Similarly, engineered attenuated *Salmonella typhimurium* VNP20009 expressing the eukaryotic apoptosis-inducing factor triggered an anti-tumor effect in in vitro and in vivo models (Wang, et al. [94]). In another study, *Escherichia coli* that was engineered to express the toxin cytolysin A, significantly reduced tumor growth in primary and metastatic tumor mouse models when used in combination with radiotherapy. It should be noted that radiotherapy or treatment with the engineered *E. coli* by itself was not sufficient to rid the mice of tumors (Jiang, et al. [95]). There are more examples of genetically engineered bacteria that can deliver therapeutics to a variety of cancers in vivo models, thereby allowing them to function as potential therapeutics in the treatment of cancers (Liu, et al. [96]). The clinical efficacy of these agents in treating cancers remains to be observed in patients.

Conclusion

Bacteriotherapy has proven to be an effective therapeutic and in many cases shows little to no adverse effects. It can be used not only to treat ailments but also improve the general health of patients. There are challenges associated with bacteriotherapy that are being addressed but it is clear that ongoing research will lead to improvements that will create innovative and effective treatment strategies that harness the power of bacteria and bacterial products.

Conflict of Interest

The authors declare that they have no conflicts of interest.

References

- Stripling J, Rodriguez M (2018) Current evidence in delivery and therapeutic uses of fecal microbiota transplantation in human diseases—*Clostridium difficile* disease and beyond. *The American Journal of the Medical Sciences* 356(5): 424-432.
- Huovinen P (2001) Bacteriotherapy: the time has come. *BMJ* 323(7309): 353-354.
- Otto G (2021) Commensal bacteriotherapy. *Nature Reviews Microbiology* 19(5): 283.
- Al Jashaami LS, Dupont HL (2016) Management of *Clostridium difficile* infection. *Gastroenterology & Hepatology* 12(10): 609-616.
- Keller JJ, Kuijper EJ (2015) Treatment of recurrent and severe *Clostridium difficile* infection. *Annual Review of Medicine* 66: 373-386.
- Bakken JS, Borody T, Brandt LJ, Brill JV, Demarco DC, et al. (2011) Treating *Clostridium difficile* infection with fecal microbiota transplantation. *Clinical Gastroenterology and Hepatology* 9(12): 1044-1049.
- Floch MH (2010) Fecal bacteriotherapy, fecal transplant, and the microbiome. *Journal of Clinical Gastroenterology* 44(8): 529-530.
- Kaakoush NO (2020) Fecal transplants as a microbiome-based therapeutic. *Current Opinion in Microbiology* 56: 16-23.
- Rode A A, Chehri M, Krogsgaard L R, Heno K K, Svendsen A T, et al. (2021) Randomised clinical trial: a 12-strain bacterial mixture versus faecal microbiota transplantation versus vancomycin for recurrent *Clostridioides difficile* infections. *Alimentary Pharmacology & Therapeutics* 53(9): 999-1009.
- Verdier C, Denis S, Gasc C, Boucinha L, Uriot O, et al. (2021) An oral fnt capsule as efficient as an enema for microbiota reconstruction following disruption by antibiotics, as assessed in an *in vitro* human gut model. *Microorganisms* 9(2): 1-24.
- Yakout A, Bi Y, Harris D M (2024) *Clostridioides difficile*: A concise review of best practices and updates. *Journal of Primary Care and Community Health* 15: 21501319241249644.
- Cold F, Svensson CK, Petersen AM, Hansen LH, Helms M, et al. (2022) Long-term safety following faecal microbiota transplantation as a treatment for recurrent *Clostridioides difficile* infection compared with patients treated with a fixed bacterial mixture: Results from a Retrospective Cohort Study. *Cells* 11(3): 435.
- Li X, Wang Q, Hu X, Liu W (2022) Current status of probiotics as supplements in the prevention and treatment of infectious diseases. *Frontiers in Cellular and Infection Microbiology* 12: 789063.
- Patel R, Dupont H L (2015) New approaches for bacteriotherapy: Prebiotics, new-generation probiotics, and synbiotics. *Clinical Infectious Diseases* 60: S108-S121.
- Szyman'ski H, Pejcz J, Jawien' M, Chmielarczyk A, Strus M, et al. (2006) Treatment of acute infectious diarrhoea in infants and children with a mixture of three *Lactobacillus rhamnosus* strains—a randomized, double-blind, placebo-controlled trial. *Alimentary Pharmacology & Therapeutics* 23(2): 247-253.
- Newberry S J (2012) Probiotics for the prevention and treatment of antibiotic-associated diarrhea. *JAMA* 307(18): 1959.
- Zhang L, Zeng X, Guo D, Zou Y, Gan H, et al. (2022) Early use of probiotics might prevent antibiotic-associated diarrhea in elderly (>65 years): a systematic review and meta-analysis. *BMC Geriatrics* 22(1): 562.
- Mestre A, Sathiyarayanan R, Rivas D, John J, Abdulqader MA, et al. (2022) Role of probiotics in the management of *Helicobacter pylori*. *Cureus* 14(6): e26463.
- Yang Z, Zhou Y, Han Z, He K, Zhang Y, et al. (2024) The effects of probiotics supplementation on *Helicobacter pylori* standard treatment: an umbrella review of systematic reviews with meta-analyses. *Scientific Reports* 14(1): 10069.
- Zhang M M, Qian W, Qin Y Y, He J, Zhou Y H (2015) Probiotics in *Helicobacter pylori* eradication therapy: a systematic review and meta-analysis. *World Journal of Gastroenterology* 21(14): 4345-4357.
- Bellussi L, Passali F, Ralli M, De Vincentiis M, Greco A, Passali D (2019) An overview on upperrespiratory tract infections and bacteriotherapy as innovative therapeutic strategy. *European Review for Medical and Pharmacological Sciences* 23(1): 27-38.

22. Darbandi A, Asadi A, Ghanavati R, Afifirad R, Darb Emamie A, et al. (2021) The effect of probiotics on respiratory tract infection with special emphasis on COVID-19: Systemic review 2010–20. *International Journal of Infectious Diseases* 105: 91-104.
23. Du T, Lei A, Zhang N, Zhu C (2022) The beneficial role of probiotic *Lactobacillus* in respiratory diseases. *Frontiers in Immunology* 13: 908010.
24. Kenneally C, Murphy CP, Sleator RD, Culligan EP (2022) The urinary microbiome and biological therapeutics: Novel therapies for urinary tract infections. *Microbiological Research* 259: 127010.
25. Flores-Mireles AL, Walker JN, Caparon M, Hultgren SJ (2015) Urinary tract infections: Epidemiology, mechanisms of infection and treatment options. *Nature Reviews Microbiology* 13(5): 269-284.
26. Foxman B (2014) Urinary tract infection syndromes. Occurrence, recurrence, bacteriology, risk factors, and disease burden. *Infectious Disease Clinics of North America* 28(1): 1-13.
27. Gupta K, Hooton TM, Stamm WE (2001) Increasing antimicrobial resistance and the management of uncomplicated community-acquired urinary tract infections. *Annals of Internal Medicine* 135(1): 41-50.
28. Shah C, Baral R, Bartaula B, Shrestha L B (2019) Virulence factors of uropathogenic *Escherichia coli* (UPEC) and correlation with antimicrobial resistance. *BMC Microbiology* 19(1).
29. Beerepoot MAJ (2012) *Lactobacilli* vs antibiotics to prevent urinary tract infections. *Archives of Internal Medicine* 172(9): 704.
30. Gupta V, Mastromarino P, Garg R (2024) Effectiveness of prophylactic oral and/or vaginal probiotic supplementation in the prevention of recurrent urinary tract infections: A randomized, double-blind, placebo-controlled trial. *Clinical Infectious Diseases* 78(5): 1154-1161.
31. Markowiak P, S'lizewska K (2017) Effects of probiotics, prebiotics, and synbiotics on human health. *Nutrients* 9(9): 1021.
32. You S, Ma Y, Yan B, Pei W, Wu Q, et al. (2022) The promotion mechanism of prebiotics for probiotics: A review. *Frontiers in Nutrition* 9: 1000517.
33. Davani-Davari D, Negahdaripour M, Karimzadeh I, Seifan M, Mohkam M, et al. (2019) Prebiotics: Definition, types, sources, mechanisms, and clinical applications. *Foods* 8(3): 92.
34. Fusco W, Lorenzo MB, Cintoni M, Porcari S, Rinninella, et al. (2023) Short-chain fatty-acid-producing bacteria: key components of the human gut microbiota. *Nutrients* 15(9): 2211.
35. Lewis S, Burmeister S, Brazier J (2005) Effect of the prebiotic oligofructose on relapse of *Clostridium difficile*-associated diarrhea: a randomized, Controlled Study. *Clinical Gastroenterology and Hepatology* 3(5): 442-448.
36. van Stuijvenberg M, Eisses A M, Grüber C, Mosca F, Arslanoglu S, et al. (2011) Do prebiotics reduce the number of fever episodes in healthy children in their first year of life: a randomised controlled trial. *British Journal of Nutrition* 106(11): 1740-1748.
37. Kukkonen K, Savilahti E, Haahtela T, Juntunen-Backman K, Korpela R, et al. (2008) Long-term safety and impact on infection rates of postnatal probiotic and prebiotic (synbiotic) treatment: randomized, double-blind, placebo-controlled trial. *Pediatrics* 122(1): 8-12.
38. Hutchings MI, Truman AW, Wilkinson B (2019) Antibiotics: past, present and future. *Current Opinion in Microbiology* 51: 72-80.
39. de Lima Procopio RE, da Silva IR, Martins M K, de Azevedo JL, de Araujo JM (2012) Antibiotics produced by *Streptomyces*. *The Brazilian Journal of Infectious Diseases* 16(5): 466-471.
40. Quinn G A, Banat A M, Abdelhameed A M, Banat I M (2020) *Streptomyces* from traditional medicine: sources of new innovations in antibiotic discovery. *Journal of Medical Microbiology* 69(8): 1040-1048.
41. Alam K, Mazumder A, Sikdar S, Zhao YM, Hao J, et al. (2022) *Streptomyces*: The biofactory of secondary metabolites. *Frontiers in Microbiology*, p. 13.
42. De Kraker MEA, Lipsitch M (2022) Burden of antimicrobial resistance: compared to what? *Epidemiologic Reviews* 43(1): 53-64.
43. Chabib L, Rustandi T, Fawwazi MHAF, Kumalasari E, Lestari DA, et al. (2025) Harnessing iChip technology for novel antibiotic discovery from peat soil microbiomes to combat antimicrobial resistance. *Frontiers in Microbiology* 16: 1530273.
44. Nichols D, Cahoon N, Trakhtenberg EM, Pham L, Mehta A, et al. (2010) Use of iChip for high-throughput in situ cultivation of 'uncultivable' microbial species. *Applied and Environmental Microbiology* 76(8): 2445-2450.
45. Ling LL, Schneider T, Peoples AJ, Spoering AL, Engels I, et al. (2015) A new antibiotic kills pathogens without detectable resistance. *Nature* 517(7535): 455-459.
46. Shukla R, Peoples A J, Ludwig K C, Maity S, Derks M G N, et al. (2023) An antibiotic from an uncultured bacterium binds to an immutable target. *Cell* 186(19): 4059-4073.
47. Brodiazenko T, Turnbull KJ, Wu KJY, Takada H, Tresco BIC, et al. (2022) Synthetic oxepanoprolinamide iboxamycin is active against *Listeria monocytogenes* despite the intrinsic resistance mediated by VgaL/Lmo0919 ABCF ATPase. *JAC Antimicrobial Resistance* 4(3).
48. Mitcheltree MJ, Pisipati A, Syroegin EA, Silvestre KJ, Klepacki D, et al. (2021) A synthetic antibiotic class overcoming bacterial multidrug resistance. *Nature* 599(7885): 507-512.
49. Wu K J Y, Tresco B I C, Ramkissoon A, Aleksandrova E V, Syroegin E A, et al. (2024) An antibiotic preorganized for ribosomal binding overcomes antimicrobial resistance. *Science* 383(6684): 721-726.
50. Danquah CA, Minkah PAB, Junior IOD, Amankwah KB, Somuah SO, et al. (2022) Antimicrobial compounds from microorganisms. *Antibiotics* 11(3): 285.
51. Darbandi A, Asadi A, Mahdizade Ari M, Ohadi E, Talebi M, et al. (2022) Bacteriocins: Properties and potential use as antimicrobials. *Journal of Clinical Laboratory Analysis* 36(1): e24093.
52. Aunpad R, Na Bangchang K (2007) Pumilicin 4, a novel bacteriocin with anti-MRSA and anti-VRE activity produced by newly isolated bacteria *Bacillus pumilus* strain WAPB4. *Current Microbiology* 55(4): 308-313.
53. Shin J M, Gwak J W, Kamarajan P, Fenno J C, Rickard A H, et al. (2016) Bio-medical applications of nisin. *Journal of Applied Microbiology* 120(6): 1449-1465.
54. Kumar R, Barbhuiya RI, Bohra V, Wong JWC, Singh A, et al. (2023) Sustainable rhamnolipids production in the next decade – Advancing with *Burkholderia thailandensis* as a potent biocatalytic strain. *Microbiological Research* 272: 127386.
55. Thakur P, Saini N K, Thakur V K, Gupta V K, Saini R V, et al. (2021) Rhamnolipid the glycolipid biosurfactant: Emerging trends and promising strategies in the field of biotechnology and biomedicine. *Microbial Cell Factories* 20(1): 1.
56. Herzog M, Tiso T, Blank LM, Winter R (2020) Interaction of rhamnolipids with model biomembranes of varying complexity. *Biochimica et Biophysica Acta - Biomembranes* 1862(11): 183431.

57. Cerqueira dos Santos S, Araujo Torquato C, De Alexandria Santos D, Orsato A, Leite K, et al. (2024). Production and characterization of rhamnolipids by *Pseudomonas aeruginosa* isolated in the Amazon region, and potential antiviral, antitumor, and antimicrobial activity. *Scientific Reports* 14(1).
58. Firdose A, Maeda T, Sukri MAM, Yasin NHM, Sabturan N, et al. (2024). Antibacterial mechanism of *Pseudomonas aeruginosa* UKMP14T rhamnolipids against multidrug resistant *Acinetobacter baumannii*. *Microbial Pathogenesis* 193: 106743.
59. Kramer J, Özkaya Ö, Kümmerli R (2020) Bacterial siderophores in community and host interactions. *Nature Reviews Microbiology* 18(3): 152-163.
60. Khasheii B, Mahmoodi P, Mohammadzadeh A (2021) Siderophores: Importance in bacterial pathogenesis and applications in medicine and industry. *Microbiological Research* 250: 126790.
61. Peukert C, Gasser V, Orth T, Fritsch S, Normant V, et al. (2023) Trojan horse siderophore conjugates induce *Pseudomonas aeruginosa* suicide and qualify the TonB protein as a novel antibiotic target. *Journal of Medicinal Chemistry* 66(1): 553-576.
62. Filipić B, Usjak D, Rambaher MH, Oljatic S, Milenković MT, et al. (2024) Evaluation of novel compounds as anti-bacterial or anti-virulence agents. *Frontiers in Cellular and Infection Microbiology* 14: 1370062.
63. Hu KA, Gubatan J (2023) Gut microbiome-based therapeutics in inflammatory bowel disease. *Clinical and Translational Discovery* 3(2).
64. Nitzan O, Mazen Elias, Avi Peretz, Walid Saliba (2016) Role of antibiotics for treatment of inflammatory bowel disease. *World Journal of Gastroenterology* 22(3): 1078-1087.
65. Prantera C, Lochs H, Grimaldi M, Danese S, Scribano M L, et al. (2012) Rifaximin-extended intestinal release induces remission in patients with moderately active Crohn's disease. *Gastroenterology* 142(3): 473-481.
66. Tamaki H, Nakase H, Inoue S, Kawanami C, Itani T, Ohana M, et al. (2016) Efficacy of probiotic treatment with *Bifidobacterium longum* 536 for induction of remission in active ulcerative colitis: A randomized, double-blinded, placebo-controlled multicenter trial. *Digestive Endoscopy* 28(1): 67-74.
67. Sood A, Midha V, Makharia G K, Ahuja V, Singal D, et al. (2009) The probiotic preparation, VSL#3 induces remission in patients with mild-to-moderately active ulcerative colitis. *Clinical Gastroenterology and Hepatology* 7(11): 1202-1209.
68. Roy S, Dhaneshwar S (2023) Role of prebiotics, probiotics, and synbiotics in management of inflammatory bowel disease: Current perspectives. *World Journal of Gastroenterology* 29(14): 2078-2100.
69. Cui B, Feng Q, Wang H, Wang M, Peng Z, et al. (2015) Fecal microbiota transplantation through mid-gut for refractory Crohn's disease: Safety, feasibility, and efficacy trial results. *Journal of Gastroenterology and Hepatology* 30(1): 51-58.
70. Wang Y, Ren R, Sun G, Peng L, Tian Y, et al. (2020) Pilot study of cytokine changes evaluation after fecal microbiota transplantation in patients with ulcerative colitis. *International Immunopharmacology* 85: 106661.
71. Nakatsuji T, Hata TR, Tong Y, Cheng JY, Shafiq F, et al. (2021) Development of a human skin commensal microbe for bacteriotherapy of atopic dermatitis and use in a phase 1 randomized clinical trial. *Nature Medicine* 27(4): 700-709.
72. Myles IA, Earland NJ, Anderson ED, Moore IN, Kieh MD, et al. (2018) First-in-human topical microbiome transplantation with *Roseomonas mucosa* for atopic dermatitis. *JCI Insight* 3(9): e120608.
73. Umborowati M A, Damayanti D, Anggraeni S, Endaryanto A, Surono I S, et al. (2022) The role of probiotics in the treatment of adult atopic dermatitis: a meta-analysis of randomized controlled trials. *Journal of Health, Population and Nutrition* 41(1): 37.
74. Nguyen DH, Chong A, Hong Y, Min JJ (2023) Bioengineering of bacteria for cancer immunotherapy. *Nature Communications* 14(1): 3553.
75. Sedighi M, Bialvaei A Z, Hamblin M R, Ohadi E, Asadi A, et al. (2019) Therapeutic bacteria to combat cancer; current advances, challenges, and opportunities. *Cancer Medicine* 8(6): 3167-3181.
76. Soleimanpour S, Hasanian S M, Avan A, Yaghoubi A, Khazaei M (2020) Bacteriotherapy in gastrointestinal cancer. *Life Sciences* 254: 117754.
77. Kang SR, Nguyen DH, Yoo SW, Min JJ (2022) Bacteria and bacterial derivatives as delivery carriers for immunotherapy. *Advanced Drug Delivery Reviews* 181: 114085.
78. Wei M Q, Ellem K A O, Dunn P, West M J, Bai C X, et al. (2007) Facultative or obligate anaerobic bacteria have the potential for multimodality therapy of solid tumours. *European Journal of Cancer* 43(3): 490-496.
79. Chang J, Liu Y, Han B, Zhou C, Bai C, et al. (2015) *Pseudomonas aeruginosa* preparation plus chemotherapy for advanced non-small-cell lung cancer: a randomized, multicenter, double-blind phase III study. *Medical Oncology* 32(5): 139.
80. Mukherjee N, Wheeler KM, Svatek RS (2019) Bacillus Calmette-Guérin treatment of bladder cancer: A systematic review and commentary on recent publications. *Current Opinion in Urology* 29(3): 181-188.
81. Jiang S, Redelman-Sidi G (2022) BCG in bladder cancer immunotherapy. *Cancers* 14(13): 3073.
82. Lamm DL, Blumenstein BA, Crawford ED, Montie JE, Scardino P, et al. (1991) A randomized trial of intravesical doxorubicin and immunotherapy with Bacille Calmette-Guérin for transitional-cell carcinoma of the bladder. *The New England Journal of Medicine* 325(17): 1205-1209.
83. Krajewski W, Moschini M, Chorbin'ska J, Nowak Ł, Poletajew S, et al. (2021) Delaying BCG immunotherapy onset after transurethral resection of non-muscle-invasive bladder cancer is associated with adverse survival outcomes. *World Journal of Urology* 39(7): 2545-2552.
84. Alhogbani MM, Picard JA, Fassi Fehri MH, Badet JL, Colombel CM, et al. (2017) Prognostic impact of Bacillus Calmette-Guérin interruption at the time of induction and consolidation. *Urology Annals* 9(4): 315-320.
85. Kates M, Nirschl T, Sopko NA, Matsui H, Kochel CM, et al. (2017) Intravesical BCG induces CD4+ T-cell expansion in an immune competent model of bladder cancer. *Cancer Immunology Research* 5(7): 594-603.
86. Zhang W, Yu L, Chang Z, Xiong H (2024) BCG immunotherapy promotes tumor-derived T-cell activation through the FLT3/FLT3LG pathway in bladder cancer. *Journal of Cancer* 15(3): 623-631.
87. Trivanović D, Pavelić K, Peršurić Z (2021) Fighting cancer with bacteria and their toxins. *International Journal of Molecular Sciences* 22(23): 12980.
88. Patyar S, Joshi R, Prasad Byrav D, Prakash A, Medhi B, et al. (2010) Bacteria in cancer therapy: a novel experimental strategy. *Journal of Biomedical Science* 17(1): 21.
89. Michl P, Buchholz M, Rolke M, Kunsch S, Löhr M, et al. (2001) Claudin-4: A new target for pancreatic cancer treatment using *Clostridium perfringens* enterotoxin. *Gastroenterology* 121(3): 678-684.
90. Ansiaux R, Gallez B (2007) Use of botulinum toxins in cancer therapy. *Expert Opinion on Investigational Drugs* 16(2): 209-218.

91. Grenda T, Grenda A, Krawczyk P, Kwiatek K (2022) Botulinum toxin in cancer therapy—current perspectives and limitations. *Applied Microbiology and Biotechnology* 106(2): 485-495.
92. Shafiee F, Aucoin M G, Jahanian Najafabadi A (2019) Targeted diphtheria toxin-based therapy: A Review Article. *Frontiers in Microbiology* 10: 2340.
93. Zheng J H, Nguyen V H, Jiang S N, Park S H, Tan W, et al. (2017) Two-step enhanced cancer immunotherapy with engineered *Salmonella typhimurium* secreting heterologous flagellin. *Science Translational Medicine* 9(376): eaak9537.
94. Wang H, Chen T, Wan L, Lu J, Wei H, et al. (2020) Attenuated *Salmonella* engineered with an apoptosis-inducing factor (AIF) eukaryotic expressing system enhances its anti- tumor effect in melanoma in vitro and in vivo. *Applied Microbiology and Biotechnology* 104(8): 3517-3528.
95. Jiang SN, Phan TX, Nam TK, Nguyen VH, Kim HS, et al. (2010) Inhibition of tumor growth and metastasis by a combination of *Escherichia coli*-mediated cytolytic therapy and radiotherapy. *Molecular Therapy: The Journal of the American Society of Gene Therapy* 18(3): 635-642.
96. Liu J, He C, Tan W, Zheng JH (2024) Path to bacteriotherapy: From bacterial engineering to therapeutic perspectives. *Life Sciences* 352: 122897.

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