

# A Case Report of Atypical Mycobacterial Cutaneous Infection in a Brazilian Cat

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## ARTICLE INFO

**Received:** 📅 April 07, 2025

**Published:** 📅 April 25, 2025

**Citation:** Paolo Ruggero Errante. A Case Report of Atypical Mycobacterial Cutaneous Infection in a Brazilian Cat. Biomed J Sci & Tech Res 61(4)-2025. BJSTR. MS.ID.009615.

## ABSTRACT

Infections caused by cutaneous mycobacteriosis are distributed worldwide and manifest in three forms: atypical mycobacteriosis, feline leprosy and cutaneous tuberculosis. Atypical cutaneous mycobacteriosis in cats is a rare disease commonly caused by rapidly growing mycobacteria belonging to group IV of Runyon classification. This case report describes a case of dermatitis resulting from infection by an atypical mycobacterium of *Mycobacterium fortuitum* complex in a five-year-old female, mixed breed feline. The cat presented ulcerated lesions, accompanied by exudative fistulas, with caseous-looking material. The presence of mycobacteria from the *Mycobacterium fortuitum* complex was identified after histopathology, bacterial culture and biochemical tests.

**Keywords:** Dermatitis; Dermatopathies; Atypical Bacteria; Feline Cutaneous Mycobacteriosis; *Mycobacterium fortuitum*

## Introduction

Mycobacterial diseases have an infectious, granulomatous, chronic and progressive nature, and are caused by bacteria of the order Actinomycetale, family Mycobacteriaceae, genus *Mycobacterium*, which have a cell wall rich in complex lipids and mycolic acid, which promotes the characteristic of alcohol-acid resistance when subjected to Ziehl-Neelsen staining [1]. Mycobacteria isolated from domestic animals are divided into two large groups according to the culture growth time, being classified as slow or fast (group IV of the Runyon classification), and according to the type of infection triggered, such as leprosy, tuberculous or non-tuberculous [1,2]. All are non-chromogenic, fast-growing, Gram-positive, acid-alcohol resistant, aerobic and non-spore forming. They are ubiquitous in nature, being found in water and moist soil, and in the enteric tract of pigs and ruminants, and are not pathogenic for healthy and immunocompetent animals [3]. Cutaneous mycobacteriosis is uncommon in dogs and cats, and dermatopathies related to mycobacterial skin diseases include atypical cutaneous mycobacteriosis, canine leproid granuloma syndrome, feline leprosy, and cutaneous tuberculosis [4,5]. Cutaneous mycobac-

teriosis is uncommon in dogs and cats, and dermatopathies related to mycobacterial skin diseases include atypical cutaneous mycobacteriosis, canine leproid granuloma syndrome, feline leprosy, and cutaneous tuberculosis [6]. Feline mycobacteriosis associated with infections by atypical mycobacteria are uncommon and, in felines, include *Mycobacterium fortuitum*, found in soil and aquatic biofilms in different geographic regions. It is a fast-growing mycobacterium belonging to Runyon group IV, which has three biovariants (biovar peregrinum, biovar fortuitum and biovar inominata), in addition to the species *M. phlei*, *M. smegmatis* and *M. chelonae* [7-10].

Felines infected with mycobacteria may present a clinical picture characterized by weight loss, coughing and dyspnea, associated or not with lymphadenopathy [11,12]. The cutaneous form of disease is characterized by multifocal nodules with or without ulcerations, and regional lymphadenopathy may be the only clinical finding [13]. Cats are more susceptible to mycobacterial infections, since the entry point is through the presence of lacerations and traumatic skin breaks [14]. These lesions are normally observed in the ventral abdominal region (mesogastric and hypogastric), with the presence of

macules, nodules and draining fistulas presenting a chronic and recurrent character [15]. On palpation, the panniculus is thickened, firm and nodular. These lesions must be differentiated from other diseases such as nodular panniculitis, pansteatitis, foreign body granulomas, nocardiosis, protothecosis, actinomycosis, cryptococcosis and sporotrichosis, pseudomycetoma, neoplasms, feline leprosy, tuberculosis and abscesses secondary to immunosuppressive infections (feline leukemia virus/FeLV and feline viral immunodeficiency/FIV) [16,17]. Diagnosis is based on direct examination of exudate or decals of injured tissue stained by the Ziehl-Neelsen technique, bacterial culture (Löwestein-Jensen, Petragnani or Stonebrink culture media), biochemical studies of isolated strains and histopathology [14,18]. Therapy involves the use of antibiotics associated with aggressive surgical debridement [19]. The objective of this study was to describe a case of atypical cutaneous mycobacteriosis in a feline resident in the city of São Paulo, state of São Paulo, Brazil.

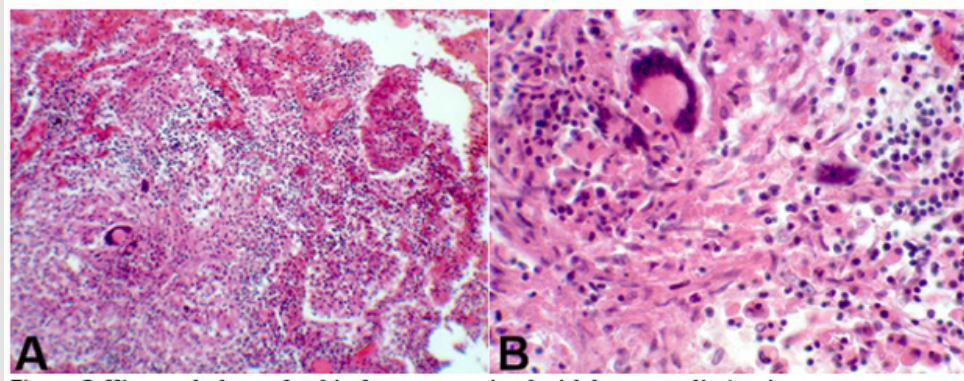
### Case Report

In 2025, a nulliparous, mixed breed, spayed, five-year-old female cat with yellow fur, who frequently goes outside, was seen by owners living in the outskirts of the city of São Paulo, São Paulo, Brazil. When consulted, the guardians reported the appearance a few months ago of multiple exuding wounds in the cat's abdomen region. According to the owners, the cat did not present any changes in appetite, water intake, bowel or urination habits. The physical examination revealed normal temperature, normal sphygmomanometry, normal opnea, normal colored mucous membranes, no lymphadenopathy or chang-

es in cardiopulmonary auscultation or abdominal palpation. On clinical examination, the skin lesions were characterized as ulcerated areas, with fistulous tracts containing caseous-looking material inside (Figure 1). Based on the appearance of the lesions, a presumptive diagnosis of bacterial panniculitis caused by atypical mycobacteria was established. The animal underwent hematological and biochemical tests, electrocardiogram, abdominal ultrasound and chest X-ray. From the ulcerative lesions, material was collected for histopathological examination, bacterioscopy using decals of injured tissues subjected to Ziehl-Neelsen staining, and bacteriological culture. No hematological, biochemical, radiographic or ultrasound alterations were evidenced by complementary examinations. Biopsy specimens were fixed in 10% formalin and routinely processed for histopathology, and histological sections stained with hematoxylin and eosin (H/E) were evaluated. Histopathological examination of a skin fragment demonstrated the presence of granulomas (Figure 2A) containing macrophages with the appearance of epithelioid cells and multinucleated giant cells (Figure 2B). For the investigation of acid-alcohol resistant bacteria, decals of damaged tissues subjected to Ziehl-Neelsen staining were used, where the presence of bacilli stained in red was verified (Figure 3). In the bacterial culture examination, there was growth of mycobacteria in Löwestein-Jensen medium over a period of five days at room temperature. Through biochemical tests (iron uptake, catalase activity, growth in sodium citrate, fructose and mannitol, and nitrate and urease tests) the presence of atypical mycobacteria, in this case *M. fortuitum*, was confirmed.



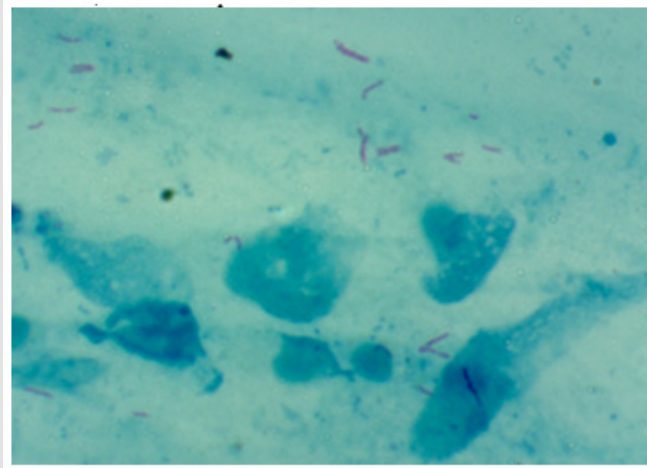
**Figure 1:** Ulcerated areas, with exudative fistulous tracts in the ventral abdominal region.



**Figure 2:** Histopathology of a skin fragment stained with hematoxylin/eosin.

A. Presence of granuloma. Magnification 10X.

B. Inflammatory infiltrate containing epithelioid cells and multinucleated giant cells. Magnification 40X.



**Figure 3:** Investigation of acid-fast bacilli from a smear of damaged tissue. Presence of acid-fast bacteria in tissue smear using Ziehl-Neelsen staining. Magnification using a 100x objective and immersion oil.

## Discussion

In Brazil, the occurrence of atypical mycobacteriosis is uncommon. Infections by atypical mycobacteria, also called opportunistic, are more commonly described in felines than dogs, and are associated with trauma, inoculation during fights, use of surgical material or contaminated needles. In felines, lesions are more common in the ventro-caudal (inguinal, umbilical) or lumbar abdominal region [15], our clinical findings are in agreement with the literature. These lesions are usually painless and unaccompanied by lymph node enlargement, pyrexia or dysorexia, as observed in the cat in this case report. In some immunosuppressed animals, lesions may spread or develop into sepsis [4,11]. In humans, deficits in T-lymphocyte-mediated immunity, compromising the functioning of interleukin-12/interferon-gamma

axis, or secondarily the use of immunosuppressive drugs, neoplasms, malnutrition, advanced age and endocrinopathies can favor the emergence of infectious conditions caused by mycobacteria [20]. In the cat in this case report, hematologic, biochemical, hormonal, and serologic (FIV/FeLV) findings did not indicate the presence of immunosuppression. According to the literature, hematological and biochemical changes are not specific, usually suggesting a chronic inflammatory condition [19]. Atypical mycobacteriosis in cats mimics other dermatopathies that present nodules and tissue loss in the form of erosions, ulcers and fistulas [21], it is essential that they are differentiated from degenerative panniculitis (pansteatitis) or infectious panniculitis (nocardiosis, sporotrichosis, dermatophytosis, cryptococcosis, actinomycosis, tuberculosis and leprosy) [9,13,22-26]. The infection usually remains localized in the cutaneous and subcutaneous tissues in a

healthy, immunocompetent host. Although adjacent structures such as the abdominal wall may be affected, spread of infection to internal organs and locally draining lymph nodes is uncommon [5].

In cases of cutaneous infection by atypical mycobacteria, pyogranulomatous inflammation of the subcutaneous adipose tissue, overlying dermis, abdominal fascia and underlying musculature is histologically observed [5,8,9]. Mycobacteria are usually difficult to observe in Ziehl-Neelsen-stained tissue sections, although acid-fast bacilli can be detected within macrophages or extracellular lipid vacuoles [5,19]. In this case report, the histopathological examination provided elements for establishing the diagnosis, prior to microbiological identification, through the presence of granulomas, although mycobacteria can be seen in 50% of cases, especially those in the fast-growing group [19]. Although it is not always possible to directly observe mycobacteria from exudate or fragments of damaged tissue stained by the Ziehl-Neelsen method, examination of damaged tissue decals allowed the observation of acid-resistant microorganisms. Fine needle aspirates or smears from skin lesions (ulcers, drainage tracts) or granulomatous lymph nodes should be stained for acid-fast bacteria using the Ziehl-Neelsen method [13,19]. The sensitivity of the test is variable, since the number of bacteria within the macrophages varies depending on the mycobacterial species and host's immune response [1,5,8]. Even when mycobacteria are visualized in material stained by the Ziehl-Neelsen method, it is important to perform a bacterial culture to differentiate atypical mycobacterial infections from those originating from feline leprosy, nocardiosis or tuberculosis [4,19]. Culture of a fresh tissue sample is useful for confirming mycobacterial infection and identifying the species involved, and has implications for treatment, prognosis and assessment of zoonotic risk, and is usually performed in a specialized laboratory. As many mycobacterial species grow slowly or do not grow in culture media, it is advisable to send tissue samples for examination using polymerase chain reaction (PCR) [27].

In bacterial culture, strains identified as *M. fortuitum* are acid-resistant, grow in less than 7 days at 28 °C and 37 °C, produce non-pigmented colonies, have a positive three-day arylsulfatase reaction, are positive for iron uptake, and reduce nitrate [28], as observed in the culture obtained from material from the cat in this case report. Also, the characterization in the cultivation in Lowenstein-Jensen medium, associated with rapid aerobic growth, presenting itself as an immobile, non-spore-forming, acid-alcohol resistant bacillus, provided the necessary elements for the identification of *M. fortuitum*, even without resorting to molecular biology techniques [27,28]. Despite the diagnosis being made, it was not possible to carry out treatment due to the loss of contact with tutor.

## Conclusion

Cutaneous infection by atypical mycobacteria in felines is rarely diagnosed and little described in the veterinary literature in small

animals in Brazil. This is due to its low frequency, lack of knowledge of the disease by clinical veterinarians and its form of diagnosis and treatment. However, the combination of histopathology, biochemical and culture tests allows for a highly sensitive diagnostic approach.

## References

- Middlemiss C, Clark J (2018) Mycobacterium in pets. *J Vet Rec* 183(18): 571.
- Wallace E, Hendrickson D, Tolli N, Mehaffy C, Peña M, et al. (2021) Culturing Mycobacteria. *Methods Mol Biol* 2314: 1-58.
- Tortoli E (2014) Microbiological features and clinical relevance of new species of the genus Mycobacterium. *Clin Microbiol Rev* 27(4): 727-752.
- Gunn Moore DA (2014) Feline mycobacterial infections. *Vet J* 201(2): 230-238.
- Malik R, Wigney DI, Dawson D, Martin P, Hunt GB, et al. (2000) Infection of the subcutis and skin of cats with rapidly growing mycobacteria: a review of microbiological and clinical findings. *J Feline Med Surg* 2(1): 35-48.
- Howard ST, Byrd TF (2000) The rapidly growing mycobacteria: saprophytes and parasites. *Microbes Infect* 2(15): 1845-1853.
- Horne KS, Kunkle GA (2009) Clinical outcome of cutaneous rapidly growing mycobacterial infections in cats in the South-Eastern United States: a review of 10 cases (1996-2006). *J Feline Med Surg* 11(8): 627-632.
- Youssef S, Archambault M, Parker W, Yager J (2002) Pyogranulomatous panniculitis in a cat associated with infection by the Mycobacterium fortuitum/peregrinum group. *Can Vet J* 43(4): 285-287.
- Alander Damsten YK, Brander EE, Paulin LG (2003) Panniculitis, due to Mycobacterium smegmatis in two Finnish cats. *J Feline Med Surg* 5(1): 19-26.
- Jang SS, Hirsh DC (2002) Rapidly growing members of the genus Mycobacterium affecting dogs and cats. *Am Anim Hosp Assoc* 38(3): 217-220.
- Pekkarinen H, Airas N, Savolainen LE, Rantala M, Kilpinen S, et al. (2018) Non-tuberculous Mycobacteria can cause disseminated Mycobacteriosis in cats. *J Comp Pathol* 160: 1-9.
- Barandiaran S, Martínez Vivot M, Falzoni E, Marfil MJ, Pérez Tort G, et al. (2017) Mycobacterioses in dogs and cats from Buenos Aires, Argentina. *J Vet Diagn Invest* 29(5): 729-732.
- Malik R, Smits B, Reppas G, Laprie C, O'Brien C, et al. (2013) Ulcerated and nonulcerated nontuberculous cutaneous mycobacterial granulomas in cats and dogs. *J Vet Dermatol* 24(1): 146-153.
- Krajewska Wedzina M, Dabrowska A, Augustynowicz Kopec E, Weiner M, Szulowski K (2019) Nontuberculous mycobacterial skin disease in cats: diagnosis and treatment-Case report. *Ann Agric Environ Med* 26(3): 511-513.
- Gunn Moore DA, McFarland SE, Brewer JI, Crawshaw TR, Clifton Hadley RS, et al. (2011) Mycobacterial disease in cats in Great Britain: I. Culture results, geographical distribution and clinical presentation of 339 cases. *J Feline Med Surg* 13(12): 934-944.
- Baral RM, Metcalfe SS, Krockenberger MB, Catt MJ, Barrs VR, et al. (2006) Disseminated Mycobacterium avium infection in young cats: overrepresentation of Abyssinian cats. *J Feline Med Surg* 8(1): 23-44.
- Paharsingh I, Suepaul R, Gyan L, Hosein A, Pargass I, et al. (2020) Disseminated Mycobacterium avium subsp. hominissuis infection and ascites in a FIV-positive cat. *Vet Clin Pathol* 49(3): 465-469.



18. Davies JL, Sibley JÁ, Myers S, Clark EG, Appleyard GD, et al. (2006) Histological and genotypical characterization of feline cutaneous mycobacteriosis: a retrospective study of formalin-fixed paraffin-embedded tissues. *Vet Dermatol* 17(3): 155-162.
19. Lloret A, Hartmann K, Pennisi MG, Gruffydd Jones T, Addie D, et al. (2013) Mycobacterioses in cats: ABC guidelines on prevention and management. *J Feline Med Surg* 15(7): 591-597.
20. Noma K, Mizoguchi Y, Tsumura M, Okada S (2022) Mendelian susceptibility to mycobacterial diseases: state of the art. *Clin Microbiol Infect* 28(11): 1429-1434.
21. Lilly ML, Siracusa C (2024) Skin disease and behavior changes in the cat. *Vet Clin North Am Small Anim Pract* 54(1): 135-151.
22. Nakanishi A, Mashita T, Akiyama K, Nakanisk W, Mori T, et al. (2015) Suppurative granulomatous sinorhinitis associated with *Nocardia* spp. infection in a cat. *J Vet Med Sci* 77(5): 597-599.
23. Malik R, Ktockenberger MB, O Brien CR, White JD, Foster D, et al. (2006) *Nocardia* infections in cats: a retrospective multi-institutional study of 17 cases. *Aust Vet J* 84(7): 235-245.
24. Lloret A, Hartmann K, Pennisi MG, Ferrer L, Addie D, et al. (2013) Sporotrichosis in cats: ABC guidelines on prevention and management. *J Feline Med Surg* 15(7): 619-623.
25. Pennisi MG, Hartmann K, Lloret A, Ferrer L, Addie D, et al. (2013) Cryptococcosis in cats: ABC guidelines on prevention and management. *J Feline Med Surg* 15(7): 611-618.
26. Moriello KA, Coyner K, Paterson S, Mignon B (2017) Diagnosis and treatment of dermatophytosis in dogs and cats: Clinical Consensus Guidelines of the World Association for Veterinary Dermatology. *Vet Dermatol* 28(3): 266-268.
27. Reppas G, Fyfe J, Foster S, Smits B, Martin P, et al. (2013) Detection and identification of mycobacteria in fixed stained smears and formalin-fixed paraffin-embedded tissues using PCR. *J Small Anim Pract* 54(12): 638-646.
28. Silcox VA, Good RC, Floyd MM (1981) Identification of clinically significant *Mycobacterium fortuitum* complex isolates. *J Clin Microbiol* 14(6): 686-691.

ISSN: 2574-1241

DOI: 10.26717/BJSTR.2025.61.009615

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