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GC-MS Analysis of Terpenoids Extracted from *Buchholzia Coriacea* Seed Displayed Inhibitory Potentials Against Sars Spike Protein Receptor Binding Pocket

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ARTICLE INFO	ABSTRACT
Received: iiii May 16, 2024 Published: iiii June 04, 2024	Corona Viruses (CoVs), family of viruses that cause intestinal and respiratory- mild colds illnesses in humans and animals. The emergence of the severe acute respiratory syndrome (SARS) epidemic in China in 2002- 2003 and the Middle Fast respiratory syndrome (MERS) was a treat to the world. HCoVs have been linked
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Olusola A O. GC-MS Analysis of Terpenoids Extracted from *Buchholzia Coriacea* Seed Displayed Inhibitory Potentials Against Sars Spike Protein Receptor Binding Pocket. Biomed J Sci & Tech Res 56(5)-2024. BJSTR. MS.ID.008915. and animals. The emergence of the severe acute respiratory syndrome (SARS) epidemic in China in 2002-2003 and the Middle East respiratory syndrome (MERS) was a treat to the world. HCoVs have been linked with major outbreaks of human fatal pneumonia since the beginning of the 21st century. *Buchholzia coriacea* (BC) plant is useful in folk medicine for the treatment of a wide range of diseases due to its secondary metabolites. In this research work, the *in-silico* study of BC terpenoid compounds was evaluated for their inhibitory potential against SARS spike protein receptor binding pocket. Terpenoid compounds were extracted from BC seed and the Gas Chromatography-Mass Spectrophotometer (GC-MS) analysis showed that it contained; rutin, phytic acid, caffeic acid, flavone, ferulic acid, gallic acid and phenol. These compounds were then docked with SARS spike protein receptor binding domain. Molecular docking revealed that they had high binding affinities compared to the standard drug remdesivir having the binding affinity (-4.740 kcal/mol). The ADME properties of the terpenoids from BC were evaluated to explain their pharmacokinetic and the results revealed that caffeic acid, gallic acid and ferulic acid could therefore serve as lead compounds in the treatment of SARS-COV.

Keywords: Terpenoid; Inhibitory; Corona; Affinities; Spike Protein

Abbreviations: CoVs: Corona Viruses; SARS: Severe Acute Respiratory Syndrome; MERS: Middle East Respiratory Syndrome

Literature Review

Human Corona Viruses

Corona viruses (CoVs), a family of viruses that cause respiratory and intestinal illnesses in humans and animals (Cui et al. [1]). They usually cause mild colds in people but the emergence of the severe acute respiratory syndrome (SARS) epidemic in China in 2002-2003 and the Middle East respiratory syndrome (MERS) on the Arabian Peninsula in 2012 show they can also cause severe disease (Cui et al. [1]). Today, HCoVs are well known for rapid evolution due to high nucleotide substitution and recombination rate (Vijgen et al. [2]). HCoVs have been appeared periodically in deferent places around the world and linked with major outbreaks of human fatal pneumonia since the beginning of the 21st century (Wu et al. [3]). First CoV outbreak as severe acute respiratory syndrome corona virus (SARS-CoV) started in November, 2002 at Foshan, China (Ge et al. [4]) and later turned into global infection in 2003 with a lethal rate of 10% worldwide (Lee et al. [5]). Following one decade, second HCoV pandemic was caused by Middle East respiratory syndrome corona virus (MERS-CoV), originated in June, 2012 at Jeddah, Saudi Arabia (Ge et al. [4]), with a global fatality rate of 35% (de Groot et al. [6]). Recent third major HCoV explosion, occurred in December, 2019 at Wuhan province of China, caused by highly homologous novel strain of SARS-CoV, classified as severe acute respiratory syndrome corona virus-2 (SARS-CoV-2); designated the infection as COVID-19 (Corona Virus Disease 2019) pandemic (Zhu et al. [7]). These HCoVs outbreaks are classified as continuous threat to humans and world economy because of their unpredicted emergence, rapid and easy proliferation that lead to catastrophic consequences (Zhu et al. [7]). The virus and its host shared a complex relationship, including numerous viral and host factors, for initiation of viral infection; subsequently in potential pathogenesis (Lim et al. [8]). As intracellular obligate parasites, viruses have also advanced various strategies to hi- jack host-cell machineries (Lim et al. [8]). For HCoVs, there is no potential therapeutic agent such as drug or vaccine; thus, strict implementation of high vigilance such as for SARS-CoV-2 has been recommended for prevention and to check the infection (Cheng et al. [9]). Researchers and Authorities (WHO, CDC Atlanta and others) across the globe are working to combat the current on-going SARS-CoV-2 outbreaks, and identifying the possible origin of this novel virus to develop effective therapeutics (Cohen, et al. [10-12]). Therefore, identifying the route of origin and pathogenesis of major pathogenic HCoVs may provide cognizance in the development of potential therapeutics.

These viruses have been confronted on several occasions with prerequisite to classify a newly emerged virus associated to a severe or even fatal disease in humans under existing genera or a new species (Gorbalenya et al. [13]). In this context, current classification distributed 39 species of CoVs in 27 subgenera, 5 genera, and 2 subfamilies that categorized under family Coronaviridae, suborder Cornidovirineae, order Nidovirales, and realm Riboviria (Gorbalenya et al. [13,14]). Herein, HCoVs are categorized under subfamily Coronavirinae of family Coronaviridae, are genotypically and serologically alienated into four major genera; AlphaCoV, BetaCoV, GammaCoV, and DeltaCoV, by International Committee for Taxonomy of Viruses (Wu et al. [3]). Remdesivir, a prodrug of an adenosine nucleotide analogue, is an antiviral agent with broad-spectrum activity against viruses from several families (Brown, et al. [15]). Remdesivir was previously under development for the treatment of Ebola virus disease in the wake of the 2014-2016 Ebola outbreaks in West Africa. While it was a promising therapeutic agent for Ebola virus disease in preclinical studies (Warren, et al. [16]), monoclonal antibodies out performed remdesivir in a phase III clinical trial (Mulangu, et al. [17]) and remdesivir is no longer being developed in this indication. The antiviral activity of remdesivir against corona viruses, however, has rendered the drug of great interest during the current global pandemic.

The novel corona virus disease 2019 (COVID-19), caused by severe acute respiratory syndrome corona virus 2 (SARS-CoV-2) infection, was first reported in Wuhan, China, in December, 2019. The World Health Organization declared COVID-19 a Public Health Emergency of International Concern on 30 January, 2020 and a pandemic on 11 March, 2020 spurring an international effort to rapidly identify treatments that might ease the burden on health care systems. Phase III trials of remdesivir in COVID-19 were initiated as early as February, 2020 (WHO, 2020). Based on data from the multinational phase III ACTT-1 and SIMPLE-severe trials, remdesivir received an emergency use authorization in the USA on 1 May, 2020, and a special approval for emergency use in Japan on 7 May, 2020. Remdesivir received its first conditional approval for use in patients with severe COVID-19 in Taiwan in late May, 2020 with this conditional approval

requiring the pharmaceutical company to implement a risk management plan to ensure safety.

Buchholzia. Coriacea Plant

Buchholzia, coriacea was named after R. W. Buchholz who collected plants in Cameroon in the late 19th century (Anie, et al. [18]). It belongs to the Capparaceae family. The seed of B. coriacea has medicinal values. These seeds gave the plants its common name of "wonderful kola" because of its usage in traditional medicine. The seeds are covered in a purple aril which is chewed in Ivory Coast and has a sharp pungent taste. (Burkill [19]) carried out a research on the medical uses of this plant in different parts of Africa. It has been used for years to meet a variety of illnesses; since it has been used continually over many generations it is likely that wonderful kola actually has an effect against illnesses. As a result of its supposed broad-spectrum activity, there is need to conduct studies on potential utilization of wonderful kola in foods. This research is aimed at studying the molecular interaction between secondary metabolites analyzed from the terpenoids of B. coriacea, and SARS spike protein receptor binding domain using Schrödinger Suite and also to evaluate the ADME analysis of the terpenoids of B. coriacea using Qikprop.

Materials and Methods

Extraction of Terpenoid from Buchholzia Coriacea

The method described by (Jiang et al., 2016) was employed for the extraction of terpenoid from the seed of *Buchholzia coriacea*. The powdered seed were soaked in n-hexane for 72 hours at room temperature and filtered. The filtrate was concentrated in a rotary evaporator at 50°C. Terpenoids were extracted from the crude by 85:15 v/v n-hexane, ethyl acetate, and silica gel (pore size 230–400 mesh, Sigma-Aldrich catalog number: 227196). Agilent 6890a GC equipped with an HP5-MS column (30 m × 0.250 mm × 0.25 µm) coupled to an Agilent 5975C Mass Spectral Detector or Flame Ionization Detector (FID) was employed for the characterization of the terpenoids.

Gas Chromatography-Mass Spectrophotometer (GC/MS)

The sample was injected into Gas Chromatography (GC) in a port which was heated up to 3000C where the material then volatilized. The column was wound within a special oven which controls temperatures from -200 to 3200. The column surface is coated with a material which will separate the various chemical compounds in the sample based on size and/or polarity. Sample components that are more volatile and smaller in size will travel through the column faster than others; hence, gaseous components are separated as they flow through the column into the mass spectrophotometer. The mass spectrum is used to identify the components by comparing each to reference libraries of over 275,000 unique spectra. To quantify compounds within the analyzed sample, analysts establish a standard curve of known concentrations of each material.

Ligand Library Generation and Preparation

Structures of Terpenoid compounds (two dimensional) Caffeic acid, Ferulic acid, Flavone, Gallic acid, Phenol, and Rutin were mined from PubChem online database (Kim et al., 2016) in sdf format and was prepared using ligprep tool (Schrödinger, 2021), (Using Epik at pH 7.0 with OPLS3 force field). Maestro, Schrödinger Suite 2021 (Schrödinger, 2021). A maximum of 32 possible stereoisomers per ligand were obtained.

Target Retrieval and Protein Preparation

X-ray crystallographic structure of SARS spike protein receptor binding domain (2GHV) complexed with an inhibitor as co-crystallized ligand at the active site (RNA 960) (PDB ID: 2GHV) (Asthana et al., 2014) was retrieved from Protein Data Bank (Berman et al., 2000). It was selected as a result of the presence of an inhibitor as ligand at the active site of the protein (David et al., 2018). The retrieved protein was viewed and prepared using Maestro 12.5 (Schrödinger, 2021), Protein preparation wizard, optimized at pH 7.0 and minimized using OPLS3 as force field. Charges and bond orders were assigned and water molecules were deleted. Hydrogen was added to the heavy atoms.

Receptor Grid Generation

Receptor grid shows the area between the ligand and the protein where interaction takes place. For Glide docking, grids were generated by using OPLS-3 force field by keeping the van-der Waals scaling factor of 1.0 and charge cut-off value of 0.25. A box was generated to each direction with 14 Å × 14 Å × 14 Å for docking experiments.

Extra Precision (XP) Ligand Docking

XP ligand docking was performed rather than SP docking because XP is better than SP in scoring function and it also predicts the false positive results (Friesner et al., 2006). This docking was performed in Glide of Schrödinger Maestro v9.4 (Friesner et al., 2004). Final result of docking can be found as glide score by energy minimization. For docking, van-der Waals scaling factor was set to 0.85 and 0.15 for ligand compounds and partial charges cut-off value was fixed at -10.0 kcal/mole. The lowest glide score containing compounds were then subjected to MM-GBSA analysis for binding free energy calculation and best poses were recorded for every ligand compounds.

Prime MM-GBSA

Binding free energy calculation was also carried out for the protein ligand complexes. MM-GBSA is a combined method for binding free energy calculation which was used in this experiment that accumulates OPLSAA molecular mechanics energies (EMM), an SGB solvation model for polar solvation (GSGB), and a non-polar solvation term (GNP) composed of the non-polar solvent accessible surface area and van-der Waals interactions (Rastelli et al., 2010). The best poses from the Glide score were used for binding free energy calculation. The total free energy of binding: Δ Gbind = Gcomplex – (Gprotein + Gligand), where G = EMM + GSGB + GNP

Ligand based ADME Analysis

Analysis of physiological descriptors of a compound such as adsorption, distribution, metabolism and excretion behaviour of the ligand compounds ADME analysis was done in QikProp module of Schrodinger (Natarajan et al., 2015). It also predicts the physico-chemical nature of the compounds as well as their pharmacokinetics properties. In this study, we used the Qikprop 3.2 module of Schrodinger (Sharma et al., 2011). There are also several other descriptors also analyzed such as Predicted IC50 for blocking HERG K+channel *in-vitro*, predicted octanol or water partition coefficient [log P(o/w)], number of hydrogen bond acceptors (HBA), number of hydrogen bond donors (HBD), predicted aqueous solubility (log s), solvent-accessible surface area (SASA), skin permeability (logKp), MDCK cell permeability (MDCK), binding to human serum albumin (logKhsa), blood-brain partition coefficient (logBB), percentage human oral absorption rate (Table 1).

Table 1.	2D atoms atoms	of town on oide from	hu chholmio comi	a a a a suture at a latest	ad fram NCDI	DUDCHEM and EM	DI EDI databasa
Table 1:	2D structure	of terpenolas from	buchnoizia cori	acea extract obtai	ned from NCBI F	PUBCHEM and EM	BL-EBI database.

S/N	COMPOUND NAME	DRUG BANK ID	2D STRUCTURE			
1	Caffeic acid	689043	O- OH OH			
2	Ferulic acid	445858	0- O- OH			



Results

(Tables 2-4) and (Figures 1-7).

Table 2: Docking score of SARS spike protein receptor binding domain (2GHV) with the secondary metabolites analysed from B. coriacea and remdesivir as the standard drug.

SARS SPIKE PROTEIN RECEPTOR BINDING DOMAIN (2GHV)						
S/N	COMPOUNDS	DOCKING SCORE (kcal/mol)	XPGSCORE (kcal/mol)			
1	Rutin	-7.545	-7.574			
2	Caffeic acid	-7.297	-7.297			
3	Flavone	-7.276	-7.276			
4	Ferulic acid	-6.696	-6.696			
5	Gallic acid	-6.071	-6.078			
6	Phenol	-4.732	-4.733			
7	Remdesivir(Standard drug)	-4.74	-6.329			

Table 3: ADME properties of some secondary metabolites analysed from B. coriacea using QIKPROP.

S/N	COMPOUNDS	SASA	DonorHB	AcceptHB	QPlogPo/w
1	Rutin	767.872	9	20.55	-2.567
2	Caffeic acid	389.661	3	3.5	0.539
3	Flavone	457.132	0	2.5	3.553
4	Ferulic acid	408.488	2	3.5	1.389
5	Gallic acid	341.213	4	4.25	-0.577
6	Phenol	276.612	1	0.75	1.459

Table 4: ADME properties of some secondary metabolites analysed from B. coriacea using QIKPROP.

S/N	COMPOUNDS	QPlogHERG ⁺	QPlogBB	QPlogKhsa	%HOA
1	Rutin	-5.127	-4.278	-1.293	0
2	Caffeic acid	-2.176	-1.559	-0.803	54.072
3	Flavone	-5.451	0.078	0.137	100
4	Ferulic acid	-2.058	-1.066	-0.626	68.81
5	Gallic acid	-1.401	-1.661	-0.985	41.457
6	Phenol	-3.41	0.101	-0.531	100

Note: WHERE;

Total solvent accessible surface area, SASA = 300.0-1000.0

Hydrogen bonds donor, HB donor = 0.0-6.0

Hydrogen bonds acceptor, HB acceptor = 2.0-20.0

Predicted IC_{50} value for blockage of HERG K+ channels, QPlogHERG = Concern below -5

Predicted qualitative human or al absorption, HOA (%) = >80% is high, <25% is poor

Predicted blood/brain partition coefficient, QPlogBB = -3.0-1.2

Predicted value for serum protein binding QPlogKHsa

Predicted octanol/water partition coefficient, QPlog Po/w = -2.0-6.5

Note: Results of Gas Chromatography-Mass Spectrophotometer (GC-MS) showing terpenoid compounds extracted from *Buchholzia coriacea*. The compounds were quantified and characterized to give rutin, gallic acid, caffeic acid, ferulic acid, flavone, and phenol. **Figure 1:** Diagramatic representation of terpenoid compounds of *Buchholzia coriacea*.

Figure 2: Result of molecular interaction of the standard drug (remdesivir) and the secondary metabolites analysed from *B. coriacea* with sars spike protein binding domain receptor.

Figure 3: Molecular interaction of remdesivir with sars spike protein binding domain receptor.

Figure 4: Molecular interaction of rutin with sars spike protein binding domain receptor.

Figure 5: Molecular interaction of caffeic acid with sars spike protein binding domain receptor.

Figure 6: Molecular interaction of flavone with sars spike protein binding domain receptor.

Figure 7: Molecular interaction of feruleic acid with sars spike protein binding domain receptor.

Discussion and Conclusion

Discussion

The corona viral surface spike protein S is a type-I transmembrane glycoprotein that mediates initial host binding via the cell surface receptor angiotensin-converting enzyme 2 (ACE2), as well as the subsequent membrane fusion events required for cell entry. The SARS-CoV surface spike protein S mediates viral entry into the host cell and includes two functional domains as follows: S1 (Gly13-Arg667) and S2 (Ser668-Thr1255). S1contains the host-specific receptor binding domain (RBD), whereas S2 mediates fusion between viral and host cell membranes (Xu, et al. [20]). Angiotensin-converting enzyme2 (ACE2) was identified as a functional receptor for the SARS-CoV (Li, et al. [21]). The recently determined structure of the S1-RBD in com-

plex with the extracellular domain of ACE2 illustrates the structural basis for the initial step of virus-host recognition. As the mediator of host-specific SARS infection and a major viral surface antigen, the S protein is an attractive candidate for both vaccine development and immunotherapy. Marasco and co-workers in 2005 previously identified a potent neutralizing human monoclonal antibody against the S1 RBD, designated "80R," from two non-immune (i.e not restricted by B cell re-combination) human antibody libraries. 80R binds S1 with nano-molar affinity, blocks the binding of S1 to ACE2, prevents the formation of syncytia in-vitro (Sui et al. [22]), and inhibits viral replication in-vivo.

Deletion studies have shown that the 80R epitope on S1 is located in the minimal ACE2 binding domain, between residues 324 and 503 (Sui, et al. [23]). Terpenoid compounds used in this study were extracted from Buchholzia coriacea. Using Gas Chromatography-Mass Spectrophotometer (GC-MS), the compounds were quantified and characterized to give rutin, gallic acid, caffeic acid, ferulic acid, flavone, and phenol which have strong antioxidant properties. This study tends to predict the best inhibitor for SARS spike protein receptor binding domain (2GHV) from terpenoid compounds of Buchholzia coriacea using molecular docking approach. The binding affinity of the standard drug remdesivir was compared with other ligands of the compounds derived from B. coriacea. After docking of the phytochemicals with SARS spike protein receptor binding domain, the docking scores were as follows; rutin -7.545 kcal/mol, caffeic acid -7.297 kcal/ mol, flavone -7.276 kcal/mol, ferulic acid -6.696 kcal/mol, gallic acid -6.071 kcal/mol, phenol -4.732 kcal/mol and remdesivir -4.740 kcal/ mol [24-38]. From the result of ADME analysis, the blood brain barrier permeability of the tested compounds was nearly between the acceptable ranges which is very important for a drug to pass through those barriers rutin, gallic acid, caffeic acid, ferulic acid, flavone, and phenol showed QPlogBB value of -4.278, -1.661 -1.559, -1.066, 0.078, and 0.101 respectively.

Where the acceptable range is -3.0 to 1.2, rutin could not adhere to the QPlogBB result therefore, would not be able to penetrate blood brain barrier while the remaining compounds sailed through. The secondary metabolites: rutin, gallic acid, caffeic acid, ferulic acid, phenol and flavones showed the number of hydrogen bonds donor value of 9, 4, 3, 2, 1 and 0 respectively. Where acceptable range is 0.0-6.0 and also showed the number of hydrogen bonds acceptor value of 20.55, 4.25, 3.5, 3.5, 0.75 and 2.5 respectively. These are in the value of acceptable range (2.0-20.0) [39-59]. Rutin could not sail through the hydrogen bond donor and hydrogen bond acceptor because it has exceeded the acceptable range while phenol having the least number could also not sail through because it could not meet up the acceptable range. The other compounds were able to adhere to the range. Rutin, caffeic acid, flavone, ferulic acid, gallic acid and phenol showed solvent accessible surface area value of 767.872, 389.661, 457.137, 408.488, 341.213 and 276.612 respectively where the acceptable range value is 300.0-1000. All compounds sailed through the solvent accessible surface area value except phenol which could not meet up the acceptable range. Predicted IC50 value for blocking Human ethera-go-go related gene (HERG) K+ channel for caffeic acid, ferulic acid, gallic acid and phenol were in the acceptable range (below -5), while rutin (-5.127) and flavone (-5.451) were above the acceptable range.

The predicted octanol or water partition coefficient for the compounds were also analysed rutin, caffeic acid, flavone, ferulic acid, gallic acid and phenol showed -2.567, 0.539, 3.553, 1.389, -0.577 and 1.459 respectively where the acceptable range is -2.0-6.5. Human oral absorption rate was greater for flavone (100%), phenol (100%), ferulic acid (68.81%), caffeic acid (54.072%), gallic acid (41.457%) and rutin (0%) [60-70]. where the acceptable percentage < 25% is poor and > 80% is good. Caffeic acid, gallic acid and ferulic acid with the binding affinities -7.297 kcal/mol, -6.071 kcal/mol and -6.696 kcal/ mol greater than the binding affinity of the standard drug (remdesivir) -4.740 kcal/mol sailed through the 8 ADME analysis. Therefore, they could be lead compounds in the treatment of SARS-COV [71-74].

Conclusion

Conclusively, further wet research should be carried out on caffeic acid, gallic acid and ferulic acid using *in-vivo* analysis to validate the result. Clinical verification is needed to ascertain them as better drugs with little or no side effects in the management of SARS-COV.

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Statement of Disclosure

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References

- 1. Cu J, Li F, Shi Z L (2019) Origin and Evolution of Pathogenic Corona Viruses. Nature Reviews Microbiology 17(3): 181-192.
- Vijgen L, Keyaerts E, Moes E, Maes P, Duson G, et al. (2005) Development of one-step, real-time, quantitative reverse transcriptase PCR assays for absolute quantitation of human coronaviruses OC43 and 229E. J Clin Microbiol 43: 5452-5456.
- 3. Wu C, Liu Y, Yang Y, Zhang P, Zhong W, et al. (2020) Analysis of therapeutic targets for SARS-CoV-2 and discovery of potential drugs by computational methods. Acta Pharm Sin B 10(5): 766-788.
- Ge X Y, Hu B, Shi Z L (2015) Bat corona viruses. Bats and Viruses, pp. 127-155.
- Lee N, Hui D, Wu A, Chan P, Cameron P, et al. (2003): A major outbreak of severe acute respiratory syndrome in Hong Kong. N Engl. J Med 348(20): 1986-1994.
- de Groot R J, Baker S C, Baric R S, Brown C S, Drosten C, et al. (2013) Middle East respiratory syndrome corona virus (MERS-CoV): announcement of the corona virus study group. J Virol 87(14): 7790-7792.
- Zhu N, Zhang D, Wang W, Li X, Yang B, et al. (2020) A novel coronavirus from patients with pneumonia in China, 2019. N Engl J Med 382(8): 727-733.
- Lim Y X, Ng Y L, Tam J P, Liu D X (2016) Human coronaviruses: a review of virus- host interactions. Diseases 4(3): 26.
- Cheng V C, Wong S C, To K W, Ho P L, Yuen K Y, et al. (2020) Preparedness and proactive infection control measures against the emerging novel corona virus in China. The Journal of Hospital Infection 104(3): 254-255.
- 10. Cohen J (2020) New corona virus threat galvanizes scientists. Science 367: 492-493.
- 11. Cyranoski D (2020) This scientist hopes to test corona virus drugs on animals in locked- down Wuhan. Nature 577(7792): 607.

- 12. Lu H (2020) Drug treatment options for the 2019-new coronavirus (2019-nCoV). Bioscience Trends 14(1): 69-71.
- 13. Gorbalenya A E, Baker S C, Baric R S, de Groot R J, Drosten C, et al. (2020) Corona viridae Study Group of the International Committee on Taxonomy of V, The species Severe acute respiratory syndrome-related corona virus: classifying 2019-nCoV and naming it SARS-CoV-2. Nat Microbiol 5: 536-544.
- 14. Siddell S G, Walker P J, Lefkowitz E J, Mushegian A R, Adams M J, et al. (2019) Additional changes to taxonomy ratified in a special vote by the International Committee on Taxonomy of Viruses (October 2018). Arch Virol 164: 943-946.
- Brown A J, Won J J, Graham R L, Kenneth H Dinnon, Amy C Sims, et al. (2019) Broad spectrum antiviral remdesivir inhibits human endemic and zoonotic deltacorona viruses with a highly divergent RNA dependent RNA polymerase. Antiviral Res 169: 104541.
- Travis K Warren, Robert Jordan, Michael K Lo, Adrian S Ray, Richard L Mackman, et al. (2016) Therapeutic efficacy of the small molecule GS-5734 against Ebola virus in rhesus monkeys. Nature 531(7594): 381-385.
- Sabue Mulangu, Lori E Dodd, Richard T Davey, Olivier Tshiani Mbaya, Michael Proschan, et al. (2019) A randomized, controlled trial of Ebola virus disease therapeutics. N Engl J Med 381(24): 2293-2303.
- Anie C O, Nwabuokei I G, Oghenejobo M, Enwa F O (2015) The antibacterial effect of the leaf extract of *Buchholzia coriacea* (*Capparidaceae*) on gram negative nasal isolates. Scholars Academic Journal of Pharmacy 4(4): 226-231.
- 19. Burkill H M (1985) The useful plants of West Africa. Royal gardens, Kew 319.
- Xu Y, Lou Z, Liu Y, Pang H, Tien P, et al. (2004) Crystal structure of severe acute respiratory syndrome coronavirus spike protein fusion core. J. Biol. Chem 279(47): 49414-49419.
- Li W, Moore M J, Vasilieva N, Sui J, Wong S K, et al. (2003) Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. Nature 426(6965): 450-454.
- 22. Sui J, Li W, Murakami A, Tamin A, Matthews L J, et al. (2004) Potent neutralization of severe acute respiratory syndrome (SARS) coronavirus by a human mAb to S1 protein that blocks receptor association. Proc Natl Acad Sci USA 101(8): 2536-2541.
- 23. Sui J, Li W, Roberts A, Matthews L J, Murakami A, et al. (2005) J Virol 79: 5900-5906.
- 24. Almazan F, Dediego M L, Galan C, Escors D, Alvarez E, et al. (2006) Construction of a severe acute respiratory syndrome corona virus infectious cDNA clone and a replicon to study corona virus RNA synthesis. J Virol 80(21): 10900-10906.
- 25. Anowi F C, Ike C, Ezeokafor, Ebere C (2012) The Phytochemical, Antispamodic and Antidiarrhoea properties of the methanol extract of the leaves of *Buchholzia coriacea* family Capparaceae. International Journal of Current Pharmaceutical Research 4(3): 52-55.
- Cheung O Y, Chan J W, Ng C K, Koo C K (2004) The spectrum of pathological changes in severe acute respiratory syndrome (SARS): Histopathology 45(2): 119-124.
- Chika E, Ikegbunam M, Ugwu C, Araka O, Iroha I, et al. (2012) Evaluation of antibacterial activity of the leave extracts of *Buchholzia coriacea*. Asian Journal of Pharmaceutical and Biological Research 2(4): 204-208.
- Chinese S M E C (2004) Molecular evolution of the SARS corona virus during the course of the SARS epidemic in China. Science 303(5664): 1666-1669.

- 29. Chow K C, Hsiao C H, Lin T Y, Chen C L, Chiou S H, et al. (2004) Detection of severe acute respiratory syndrome-associated corona virus in pneumocytes of the lung. Am J Clin Pathol 121(4): 574-580.
- Chu C M, Cheng V C, Hung I F, Wong M M, Chan K H, et al. (2004) Role of lopinavir/ritonavir in the treatment of SARS: initial virological and clinical findings. Thorax 59(3): 252-256.
- Drosten C, Günther S, Preiser W, Van Der Werf S, Brodt H R, et al. (2003) Identification of a novel corona virus in patients with severe acute respiratory syndrome. New England journal of medicine 348(20): 1967-1976.
- 32. Eickmann M, Becker S, Klenk H D, Doerr H W, Stadler K, et al. (2003) Phylogeny of the SARS corona virus. Science 302: 1504-1505.
- 33. Erhirhie E O, Ekene N E (2013) Medicinal Values on Citrullus lanatus (Watermelon): Pharmacological Review. International Journal of Research in Pharmaceutical and Biomedical Sciences 4(4): 1305-1312.
- Erhirhie E O, Moke G E (2014) Xylopia aethiopica: A review of its ethnomedicinal, Chemical and Pharmacological properties. American Journal of Pharm Tech research 4(6): 22-37.
- Ezekiel O O, Onyeoziri N F (2009) Preliminary studies on the antimicrobial properties of *Buchholzia coriacea* (wonderful kola): African Journal of Biotechnology 8(3): 472-474.
- 36. Franks T J, Chong P Y, Chui P, Galvin J R, Lourens R M, et al. (2003) Lung pathology of severe acute respiratory syndrome (SARS): a study of 8 autopsy cases from Singapore. Hum Pathol 34(8): 743-748.
- Gorbalenya A E, Snijder E J, Spaan W J M (2004) severe acute respiratory syndrome corona virus phylogeny: toward consensus. J Virol 78: 7863-7866.
- Calvin J Gordon, Egor P Tchesnokov, Joy Y Feng, Danielle P Porter, Matthias Götte (2020) The antiviral compound remdesivir potently inhibits RNA-dependent RNA polymerase from Middle East respiratory syndrome corona virus. J Biol Chem 295(15): 4773-4779.
- Guan Y, Zheng B J, He Y Q, Liu X L, Zhuang Z X, et al. (2003) Isolation and characterization of viruses related to the SARS corona virus from animals in southern China. Science 302(5643): 276-278.
- Rita Humeniuk, Anita Mathias, Huyen Cao, Anu Osinusi, Gong Shen, et al. (2020) Safety, tolerability, and pharmacokinetics of remdesivir, an antiviral for treatment of COVID-19, in healthy subjects. Clin Transl Sci.
- Jacomy H, Fragoso G, Almazan G, Mushynski W E, Talbot P J, et al. (2006): Human cor- onavirus OC43 infection induces chronic encephalitis leading to disabilities in BALB/ C mice. Virology 349(2): 335-346.
- Jeffers S A, Tusell S M, Gillim Ross L, Hemmila E M, Achenbach J E, et al. (2004) CD209L (L-SIGN) is a receptor for severe acute respiratory syndrome coronavirus. Proc Natl Acad Sci USA 101(44): 15748-15753.
- 43. Jia H P, Look D C, Shi L, Hickey M, Pewe L, et al. (2005) ACE2 receptor expression and severe acute respiratory syndrome coronavirus infection depends on differentiation of human airway epithelia. J Virol 79(23): 14614-14621.
- 44. Ksiazek T G, Erdman D, Goldsmith C S, Zaki S R, Peret T, et al. (2003) A novel coronavirus associated with severe acute respiratory syndrome. N Engl J Med 348(20): 1953-1966.
- 45. Kuiken T, Fouchier R A, Schutten M, Rimmelzwaan G F, van Amerongen G, et al. (2003) Newly discovered coronavirus as the primary cause of severe acute respiratory syndrome. Lancet 362(9380): 263-270.
- Lemmens R H, Schmelzer G H, Gurib Fakim A (2013) PROTA *Buchholzia coriacea* Engl. In: (Plant Resources of TropicalAfrica / Ressources végétales de l'Afrique tropicale).

- Lu Y, Gong E C, Zhang Q Y, Gu J, Li X W, et al. (2005) Expression of SARS-CoV in various types of cells in lung tissues. Beijing Da Xue Xue Bao 37(5): 453-457.
- Mansuri Z, Shah B, Zafar M K (2020) Remdesivir and potential interactions with psychotropic medications: a COVID19 perspective. Prim Care Companion CNS Disord 22(3): 20
- 49. Martina B E, Haagmans B L, Kuiken T, Fouchier R A, Rimmelzwaan G F, et al. (2003) Virology: SARS virus infection of cats and ferrets. Nature 425(6961): 915.
- Memish Z A, Mishra N, Olival K J, Fagbo S F, Kapoor V, et al. (2013) Middle East respiratory syndrome coronavirus in bats, Saudi Arabia. Emerg Infect Dis 19(11): 1819-1823.
- 51. Nazeruddin G M, Shirish S P, Samir S S (2011) Pharmacological review of *Tridax procumbens L*. Der Pharmacia Sinica 2(4): 172-175.
- 52. Nwachukwu M I, Duru M K C, Amadi B A, Nwachukwu I O (2014) Comparative Evaluation of Phytoconstituents, Antibacterial Activities and Proximate Contents of Fresh, Oven Dried Uncooked and Cooked Samples of *Buchholzia coriacea* Seed and Their Effects on Hepatocellular Integrity. International Journal of Pharmaceutical Science Invention 3(6): 41-49.
- 53. Nweze N E, Anene B M, Asuzu I U (2011) Investigation of the antitrypanosomal activity of *Buchholzia coriacea* seed extract against a field strain of Trypanosoma congolense. African Journal of Traditional, Complementary and Alternative Medicines 8(5): 175-180.
- 54. Osterhaus A D, Fouchier R A, Kuiken T (2004) The aetiology of SARS: Koch's postulates fulfilled. Philos Trans R Soc Lond B Biol Sci 359(1447): 1081-1082.
- Peiris J S M, Lai S T, Poon L L M, Guan Y, Yam L Y C, et al. (2003) Coronavirus as a possible cause of severe acute respiratory syndrome. The Lancet 361(9366): 1319-1325.
- Pene F, Merlat A, Vabret A, Rozenberg F, Buzyn A, et al. (2003) Coronavirus 229E-related pneumonia in immunocompromised patients. Clinical Infectious Diseases Official Publication Infectious Diseases Society of America 37(7): 929-932.
- Poon L L, Chu D K, Chan K H, Wong O K, Ellis T M, et al. (2005) Identification of a novel coronavirus in bats. J Virol 79(4): 2001-2009.
- Prabhu K S, Lobo R, Shirwaikar A A, Shirwaikar A (2009) Ocimum gratissimum: A Review of its Chemical, Pharmacological and Ethnomedicinal Properties. The Open Complementary Medicine Journal 1: 1-15.
- 59. Andrea J Pruijssers, Amelia S George, Alexandra Schäfer, Sarah R Leist, Lisa E Gralinksi, et al. (2020) Remdesivir potently inhibits SARS-CoV-2 in human lung cells and chimeric SARSCoV expressing the SARS-CoV-2 RNA polymerase in mice. bioRxiv.
- Rickert K, Martinez R R, Martinez T T (1999) Pharmacist knowledge of common herbal preparations. Proceedings of the Western Pharmacology Society's 42: 1-2.

- 61. Roberts A, Paddock C, Vogel L, Butler E, Zaki S, et al. (2005a) Aged BALB/c mice as a model for increased severity of severe acute respiratory syndrome in elderly humans. J Virol 79(9): 5833-5838.
- 62. Roberts A, Vogel L, Guarner J, Hayes N, Murphy B, et al. (2005b) severe acute respiratory syndrome corona virus infection of golden Syrian hamsters. J Virol 79(1): 503-511.
- Rota P A, Oberste M S, Monroe S S, Nix W A, Campagnoli R, et al. (2003) Characterization of a novel coronavirus associated with severe acute respiratory syndrome. Science 300: 1394-1399.
- 64. Smuts H (2008) Human coronavirus NL63 infections in infants hospitalised with acute respiratory tract infections in South Africa. Influenza Other Respir. Viruses 2(4): 135-138.
- Stadler K, Masignani V, Eickmann M, Becker S, Abrignani S, et al. (2003) SARS—beginning to understand a new virus. Nature Reviews Microbiology 1(3): 209-218.
- 66. To K F, Tong J H, Chan P K, Au F W, Chim S S, et al. (2004) Tissue and cellular tropism of the coronavirus associated with severe acute respiratory syndrome: an in-situ hybridization study of fatal cases. J Pathol 202(2): 157-163.
- 67. Vabret A, Mourez T, Gouarin S, Petitjean J, Freymuth F, et al. (2003) An outbreak of coronavirus OC43 respiratory infection in Normandy, France. Clin Infect Dis 36(8): 985-989.
- van der Hoek L, Pyrc K, Jebbink M F, Vermeulen Oost W, Berkhout R J, et al. (2004) Identification of a new human coronavirus. Nat. Med 10: 368-373.
- Walsh E E, Shin J H, Falsey A R (2013) Clinical impact of human coronaviruses 229E and OC43 infection in diverse adult populations. J Infect Dis 208: 1634-1642.
- Woo P C, Lau S K, Chu C M, Chan K H, Tsoi H W, et al. (2005) Characterization and complete genome sequence of a novel coronavirus, coronavirus HKU1, from patients with pneumonia. J Virol 79: 884-895.
- 71. Woo P C Y, Lau S K P, Lam C S F, Lau C C Y, Tsang A K L, et al. (2012) Discovery of seven novel mammalian and avian coronaviruses in the genus deltacoronavirus supports bat coronaviruses as the gene source of alphacoronavirus and betacor- onavirus and avian coronaviruses as the gene source of gammacoronavirus and deltacoronavirus. J Virol 86: 3995-4008.
- 72. Wu F, Zhao S, Yu B, Chen Y M, Wang W, et al. (2020b) A new coronavirus associated with human respiratory disease in China. Nature 579: 265-269.
- Yount B, Curtis K M, Fritz E A, Hensley L E, Jahrling P B, et al. (2003) Reverse genetics with a full-length infectious cDNA of severe acute respiratory syndrome coronavirus. Proc Natl Acad Sci USA 100(22): 12995-13000.
- 74. Zhou F, Yu T, Du R, Fan G, Liu Y, et al. (2020a) Clinical course and risk factors for mortality of adult inpatients with COVID-19 in Wuhan, China: a retrospective cohort study. Lancet 395: 1054-1062.

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