

Dextromethorphan As A NOX2 Inhibitor Mitigates Endotoxemia-Induced Sickness Behavior and Brain Interleukin-1 β Production in Mice

Yi Li^{1,2}, Yubao Wang^{2,3*}, Zhan Zhao^{2,4}, Ran Zhou^{2,4} and Jing Feng^{4*}

¹Department of Respiratory and Critical Care, Tianjin Chest Hospital, Tianjin 300222, China

²Neurobiology Laboratory, National Institute of Environmental Health Sciences, National Institutes of Health, Research Triangle Park, NC 27709, USA

³Institute of Infectious Diseases, The Second Hospital of Tianjin Medical University, Tianjin 300211, China

⁴Department of Respiratory and Critical Care, Tianjin Medical University General Hospital, Tianjin 300052, China

***Corresponding author:** Yubao Wang, Neurobiology Laboratory, National Institute of Environmental Health Sciences, National Institutes of Health, Research Triangle Park, NC 27709, USA; Institute of Infectious Diseases, The Second Hospital of Tianjin Medical University, Tianjin 300211, China

Jing Feng, Department of Respiratory and Critical Care, Tianjin Medical University General Hospital, Tianjin 300052, China



ARTICLE INFO

Received:  September 03, 2021

Published:  September 16, 2021

Citation: Yi Li, Yubao Wang, Zhan Zhao, Ran Zhou, Jing Feng. Dextromethorphan As A NOX2 Inhibitor Mitigates Endotoxemia-Induced Sickness Behavior and Brain Interleukin-1 β Production in Mice. Biomed J Sci & Tech Res 38(5)-2021. BJSTR. MS.ID.006204.

Keywords: Dextromethorphan; Sickness Behavior; Interleukin-1 β ; NADPH Oxidase; Neuroinflammation

Abbreviations: ROS: Reactive Oxygen Species; NO: Nitric Oxide; NOX2: NADPH oxidase; DM: Dextromethorphan

ABSTRACT

Brain acute inflammatory response evoked by peripheral infections can result in both short-term behavior alterations (sickness behavior) and sustained microglial activation-mediated long-term neuronal damage. The proinflammatory factor IL-1 β is essential for not only sickness behavior but also long-lasting microglia activation. However, mechanisms underlying endotoxemia and IL-1 β -induced short-term sickness behavior remain unclear. In the present study, we report that NOX2 plays a critical role in endotoxemia-induced sickness behavior. Sickness behavior measured by animal response scores was significantly relieved in NOX2^{-/-} mice compared to wild type (WT) mice after a peritoneal injection of lipopolysaccharide (LPS). LPS-elicited brain mature IL-1 β levels and prolonged microglial activation were also decreased in NOX2^{-/-} mice. Dextromethorphan, an antitussive drug, was recently found to exert anti-inflammatory and neuroprotective effect through the inhibition of NOX2 activation.

Post-administration of dextromethorphan to WT mice after LPS-elicited endotoxemia significantly mitigated animal response score, brain mature IL-1 β concentration and NOX2 mRNA expression. Moreover, dextromethorphan alleviated microglial activation at 24h after LPS dosing indicated by CD-11b staining. Our findings demonstrate that NOX2 is a novel target for intervening IL-1 β -related sickness behavior during endotoxemia and raises a possibility that dextromethorphan can be used in endotoxemia patients for relieving sickness behaviors and neuroinflammation.

Introduction

Severe systemic infections (sepsis) can cause cytokine storm and lead to acute brain inflammation through the over-production of proinflammatory factors, such as reactive oxygen species (ROS), nitric oxide (NO) and cytokines release by activated

microglia [1]. Acute brain inflammatory responses might result in both short-term behavior alterations (sickness behavior) and chronic neuronal damage via sustained microglial activation [2-5]. Sickness behaviors associated with infections by bacteria, viruses

or fungi are common [6]. Sickness or sickness behavior refers to a series of physiological and behavioral changes, which is thought as a malfunction due to systemic inflammatory response during infections or injury, including fever, fatigue, sleepiness, anorexia, cognitive dysfunction, social withdrawal [7,8]. Although it might be beneficial for individuals to fight the infection induced by pathogenic microorganisms [8], immoderate sickness could further aggravate its harm to the health, even death.

For example, neuronal excitotoxicity would be exacerbation following persistence increased temperature in brain tissue during fever, even destroy the blood-brain-barrier permeability [9,10]. Multiple cytokines such as IL-1 β , IL-6 and TNF α are associated with sickness behavior [7]. IL-1 β seems to be the predominant mediator of sickness behavior in the brain, because blockade of its action by central administration of the IL-1 receptor antagonist significantly attenuates sickness behavior [11]. The propagation of neuroinflammatory cytokines during peripheral immune challenge not only induces sickness behavior but also triggers neuroinflammation characterized by the activation of microglia, the resident macrophage in the brain [12]. Moreover, prolonged neuroinflammation can cause long-term neuronal loss via the self-propelling vicious cycle between injured neurons and overacted microglia [3,5]. Among various pro-inflammatory cytokines, IL-1 β showed a high potent neurotoxicity [13,14].

Our recent study has revealed that the IL-1 β signaling is essential for maintaining endotoxemia-associated microglial activation. Reducing brain IL-1 β levels prevents chronic neuroinflammation and resultant neurodegeneration [15]. In this study, we investigated the role of NADPH oxidase (NOX2) in LPS-elicited sickness behaviors. NOX2, a major superoxide-producing enzyme, plays a critical role in the development of neuroinflammation [5-18]. We hypothesized that NOX2 is essential for sickness behavior and brain IL-1 β maturation during endotoxemia-mediated sepsis. Besides NOX2-/- mice, we also employed a NOX2 inhibitor dextromethorphan (DM), a widely used non-opioid cough suppressant, which exerts anti-inflammation and neuro-protection through the inhibition of microglial NOX2 [19,20]. In this study, we found that genetic knockout of NOX2 or pharmacological inhibition of NOX2 with dextromethorphan diminished sickness behavior in the LPS peritoneal injection mice model.

Furthermore, the deficiency of NOX2 or the post-treatment of dextromethorphan significantly decreased IL-1 β production in the brain and resultant microglial activation. This study demonstrated that NOX2 plays a critical role in modulating the brain level of IL-1 β and associated sickness behavior. Our data also suggests that dextromethorphan could be used in endotoxemia patients for relieving sickness behaviors and neuroinflammation.

Materials and Methods

Animals and Treatment

The C57BL/6J mice and NOX2-/- mice were obtained from the Jackson Laboratory (Bar Harbor, ME). Housing and breeding of animals were performed humanely with regard for alleviation of suffering following the National Institutes of Health's Guide for the Care and Use of Laboratory Animals (Institute of Laboratory Animal Resources, 2011). LPS (2.5mg/kg) or saline (2.5 mg/kg) were intraperitoneally administrated to 8-week male C57BL/6J mice and NOX2-/- mice initially, and sickness behavior was observed every 3hs after LPS injection until 24h. Dextromethorphan (12.5mg/kg) or saline (12.5mg/kg) were also intraperitoneally injected to C57BL/6J mice two times (3h and 9h after LPS injection) and sickness behavior was accessed every 3hs after LPS injection until 24h.

Sickness Behavior Assessment

We applied behavior test to evaluate the mice's sickness behavior after treatment, including the response, the moving distance, ptosis, piloerection and excretion in eyes.

- a) Response. The toughing frequency until the mice moved. We have four degree: 0. Animal moved freely without stimulation. 1. The movement of mice under one- or two-times' toughing stimulation. 2. Three to five toughing stimulation. 3. More than five stimulations.
- b) Moving distance. We also set four degree to assess moving distance. 0. The animal moved freely more than 20cm without any stimulation. 1. The moving distance is more than 20cm with stimulation. 2. The moving distance is 10cm to 20cm under toughing stimulation. 3. In case of mice moving a few steps with stimulation, they were regarded as grade 3.
- c) Appearance of ptosis, piloerection and excretion in eyes. 0. Negative. 1. Positive.

Immunohistochemistry

Mice were euthanized using fetal plus at the desired time points after injection. Brains were fixed in 4% paraformaldehyde and processed for immune staining as described previously [21,22]. We used the following primary antibodies for immunohistochemistry: antibodies against CD11b (1:400, AbDSerotec, Raleigh, NC). Immuno-staining was visualized by using 3,3'-diaminobenzidine (DAB) and urea-hydrogen peroxide tablets.

Quantification of the immunohistochemistry staining density

The quantification CD11b immunohistological staining in SN was performed by Image J software based on a protocol for

quantifying western blots [23]. Briefly, the image was first converted into the grayscale picture, and the background was adjusted before the quantifying area was selected for the measurement of the total pixels. The relative density of the staining was compared based on the density of the total pixels of a certain brain region (total pixels/area).

Cytokine Analysis

The brain tissues from WT and NOX2^{-/-} mice were measured for mouse IL-1 β (R & D Systems Inc., Minneapolis, MN) according to the manufacturer's instructions.

Real-time RT-PCR Analysis

Total RNA was extracted from the mouse brain by using a RNeasy Mini Kit from QIAGEN (Valencia, CA) to detect the level NOX2 (gp91) according to the previous description [4]. Total RNA was reversely transcribed with MuLV reverse transcriptase and oligo-dT primers. Then SYBR green PCR master mix was used for realtime PCR analysis. The primer sequences were as follow: GAPDH F (5' TTC AAC GGC ACA GTC AAG GC 3'), GAPDH R (5' GAC TCC ACG ACA TAC TCA GCACC 3'), NOX2 F (5' GAT TCAAGA TGG AGG TGG GAC 3'), NOX2 R (5' GGT CAG TGT GAA TGG GTG CC 3'). Real-time PCR amplification was performed using SYBR Green PCR Master Mix and QuantStudio 6 Flex Real-Time PCR System (Applied Biosystems, Foster City, CA, USA) according to manufacturer's protocols. Amplifications were done at 95°C for 10 s, 55°C for

30 s, and 72°C for 30 s for 40 cycles. All samples were tested in duplicate and normalized with GAPDH using the 2- $\Delta\Delta$ Ct method. Fold changes for each treatment were normalized to the control.

Statistical Analysis

All data were represented as mean \pm SEM. Data were analysed using one-way or two-way ANOVA. Tukey's post hoc tests were applied when comparing groups to only the control, or Sidak's post hoc test when comparing between multiple treatment groups. Number of mice used is indicated in each figure legend.

Results

NOX2 Plays a Critical Role in Mediating LPS-Elicited Sickness Behavior

C57BL/6J and NOX2^{-/-} mice received a single injection of LPS (2.5mg/kg; ip) for investigating the role of NOX2 in the development of LPS-induced sickness behavior. Sickness was assessed every 3hs after LPS administration up to 24h. In WT mice, the sickness behavior score showed a time-dependent increase over time, peaked around 9-15h following LPS injection, then declined gradually afterwards, but still remained higher than the base line values at 24h after LPS injection. A similar pattern of change in the sickness behavior score was observed in LPS-injected NOX2^{-/-} mice. However, a significant reduction in sickness behavior score in NOX2^{-/-} mice was observed at 9h, 12h, 15h, compared to WT mice ($p < 0.05$; Figure 1).

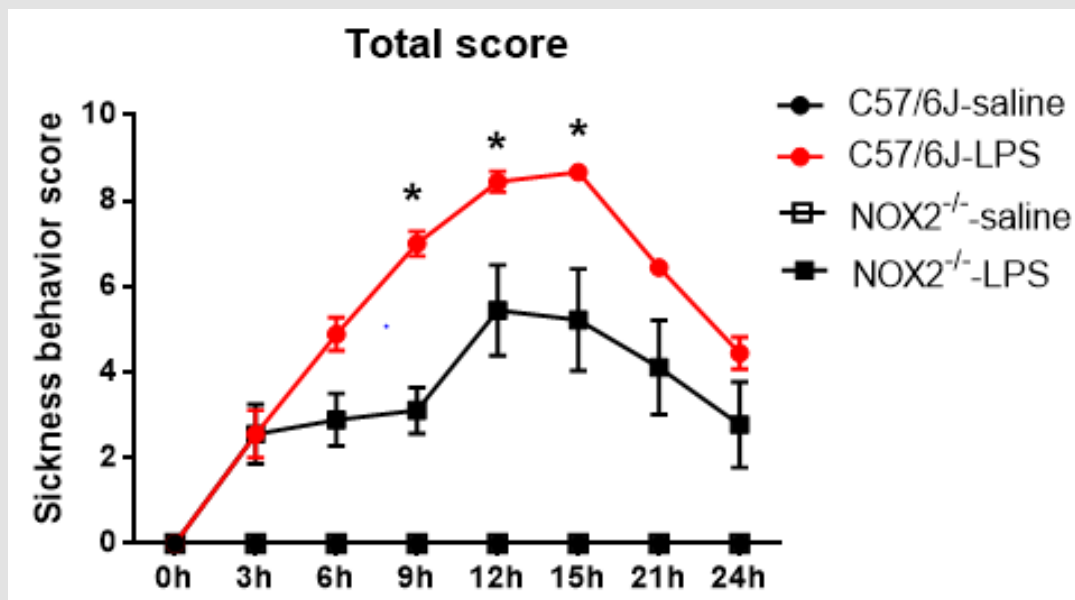


Figure 1: The total scores of sickness behavior in different groups after LPS administration. NOX2^{-/-} mice (N=9) displayed attenuated sickness behavior after LPS challenge compared to WT mice (N=9). Sickness behavior scores were measured every 3hs until 24h after LPS i.p. injection (2.5mg/kg) in WT and NOX2^{-/-} mice. * $p < 0.05$, the mean differences in total score between two groups was observed at 9h ($p < 0.0001$), 12h ($p = 0.0405$) and 15h ($p = 0.0382$) after LPS treatment. Repeated-measure ANOVA and multi-variate ANOVA to test the differences over time and differences between groups.

The absence of NOX2 Suppresses Brain Levels of IL-1 β

As mentioned previously, increase in brain IL-1 β plays a critical role in mediating sickness behavior. Our previous study indicated that the production of mature brain IL-1 β in brain peaked around 9h after LPS injection [15]. Since the sickness behavior analysis showed high scores at 9h upon the peritoneal LPS stimulation (Figure 1), we measured mature IL-1 β concentrations in brain

tissues at 9h in both WT and NOX2-deficient mice. Results showed that level of IL-1 β in brain tissues of WT mice was higher after LPS injection than that in vehicle-treated mice. Notably, the level of IL-1 β in brain tissues from NOX2 $^{-/-}$ mice after LPS administration was significantly lower than that of LPS-treated WT mice (Figure 2). This result indicates that NOX2 is essential in mediating LPS-induced increase in the IL-1 β production.

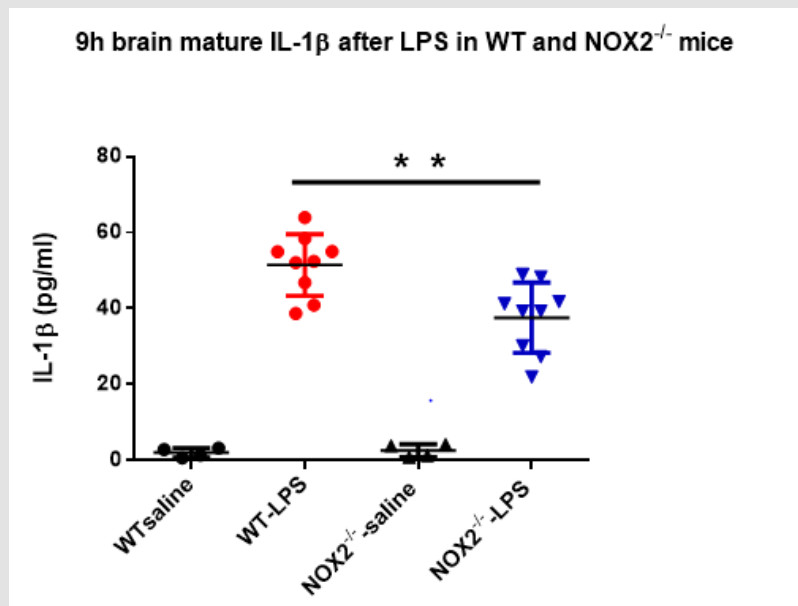


Figure 2: The brain mature IL-1 β at 9h after LPS in WT and NOX2 $^{-/-}$ mice. Mature IL-1 β concentrations in brain tissue were measured by ELISA at 9h after LPS treatment and they were decreased in NOX2 $^{-/-}$ mice compared to WT mice after LPS injection (2.5mg/kg) (n=4-9/group). **p<0.05, the differences of IL-1 β concentrations between LPS-treated WT and NOX2 groups. Two-way ANOVA followed by Sidak's multiple comparison test. (treatment \times group, F(1,22)=5.152, p=0.0334; effect of groups, F(1,22)=4.434, p=0.0469; effect of treatment, F(1,22)=177.3, p<0.0001).

The Deficiency of NOX2 relieves LPS-Elicited Activation of Microglia in The Substantia Nigra

The reduction of brain mature IL-1 β prevents sustained microglial activation induced by endotoxemia [15]. Therefore, we examined microglial activation in the absence of NOX2. Because sepsis-associated neuroinflammatory responses involve multiple brain regions and the substantia nigra (SN) was found a good area in our previous study to observe the persistence of microglial activation [3,4], we chose the SN in this study to evaluate the degree

of microglial activation by immune-staining analysis. Typical morphological changes reflecting microglial activation were observed in WT mice 24h after LPS injection. Activated microglia exhibited significant enlargement of cell bodies, irregular shapes. This morphological activation was significantly reduced in NOX2 deficient mice, characterized by a small and round shape compared with WT mice. Analysis by gray quantification showed pattern changes similar to those of morphological observations (Figure 3a & 3b).

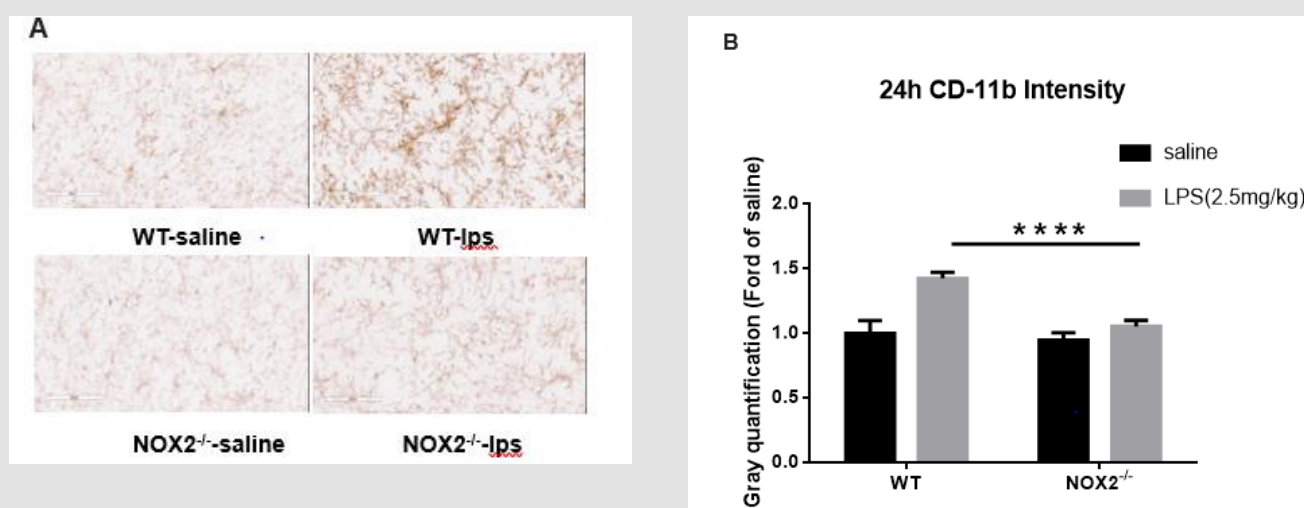


Figure 3: Microglia activation in SN was detected with CD11b IHC at 24h after LPS (2.5mg/kg) treatment.

(a) Attenuated microglial activation in NOX2^{-/-} mice compared to WT mice. Following a single injection of LPS (2.5 mg/kg; ip) or saline vehicle, mice were perfused 9 h thereafter for microglial CD-11b immunostaining (n = 3/group). Representative images at 9h were shown (Figure 3A). Scale bar = 300 μ m.

(b) CD-11b density was quantified. Results are expressed as folds of vehicle control. Histograms represent degree of microglial activation quantified by measuring the density of CD-11b staining with ImageJ. ****p<0.0001, the differences of gray quantification between LPS-treated WT and NOX2^{-/-} groups. Two-way ANOVA followed by Sidak's multiple comparison test. (treatment \times group, F(1,68)=20.45, p<0.0001; effect of groups, F(1,68)=35.93, p<0.0001; effect of treatment, F(1,68)=56.21, p<0.0001).

Targeting NOX2 as a potential therapy for sepsis-related sickness behaviors

Results from NOX2-deficient mice suggest a potential target for developing drug therapy for sepsis-related sickness behaviors. As proof of principle, we employed dextromethorphan (DM), which displays potent anti-inflammatory property via inhibition of microglial NOX2 activity [20,24]. At 3h and 9h after mice received a LPS injection, DM was intraperitoneally injected and sickness behavior was accessed every 3hs for 24hrs. In LPS treated mice, the sickness behavior score increased gradually up to 15h after LPS injection, then it decreased significantly until the end of observation. Analysis of ANOVA showed that DM significantly attenuated sickness behavior at 6h,9h, and 12h compared to mice with LPS treatment alone (p<0.05; Figure 4).

DM reduces LPS-elicited the IL-1 β production, NOX2 mRNA level and diminishes the microglial activation

LPS-elicited increase in IL-1 β production from brain tissues and microglial activation were determined in WT mice. Similar to the results from NOX2-deficient mice, DM treatments effectively reduced LPS-induced IL-1 β production (Figure 5a). NOX2 mRNA levels increased significantly following initial LPS treatment. While the NOX2 mRNA expression in DM post-treatment groups at different timepoints (6h and 12h) were lower than LPS-treated alone (Figure 5b). Pathologically, DM diminished the activation of nigral microglia as well (Figure 6a & 6b). Taken together, these results suggest a potential use of DM in ameliorating the severity of sickness behaviors and neuroinflammation in endotoxemia-related patients.

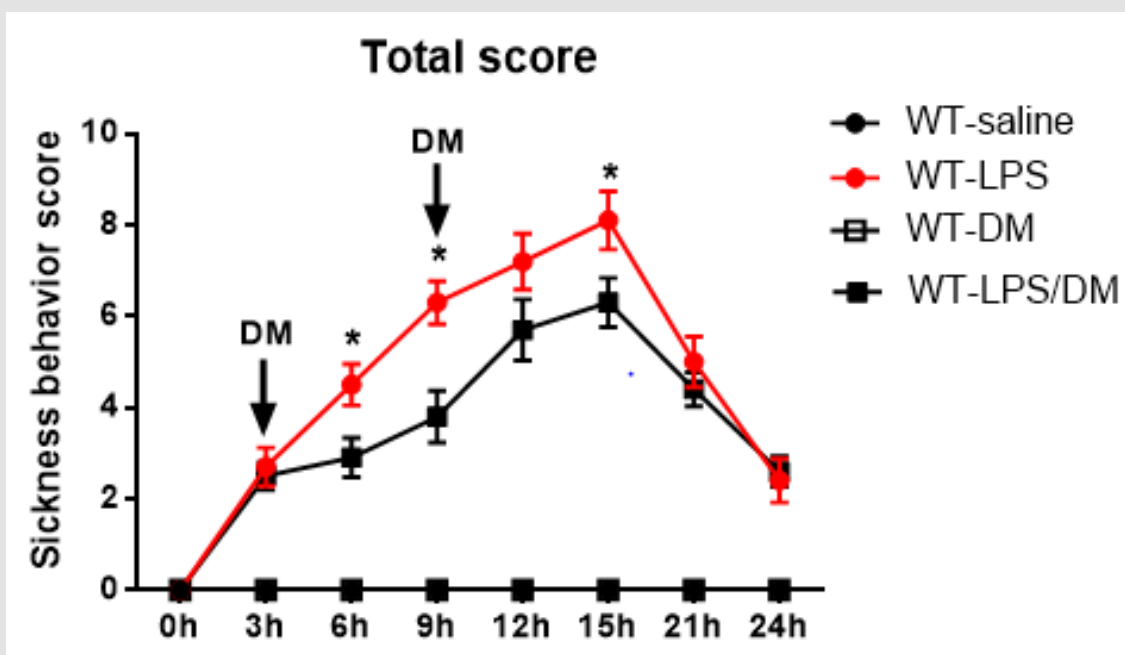


Figure 4: Post-treatment of dextromethorphan (12.5mg/kg) mitigated LPS-induced sickness behavior in WT mice (N=5-10 for each group). Sickness behavior scores were measured every 3hs until 24h after LPS i.p. injection (2.5mg/kg) and indicated administration of dextromethorphan. * $p < 0.05$, the mean differences in total score between two groups was observed at 6h ($p = 0.0459$), 9h ($p = 0.0411$) and 15h ($p = 0.0483$) after initial LPS treatment. Repeated-measure ANOVA and multi-variate ANOVA to test the differences over time and differences between groups.

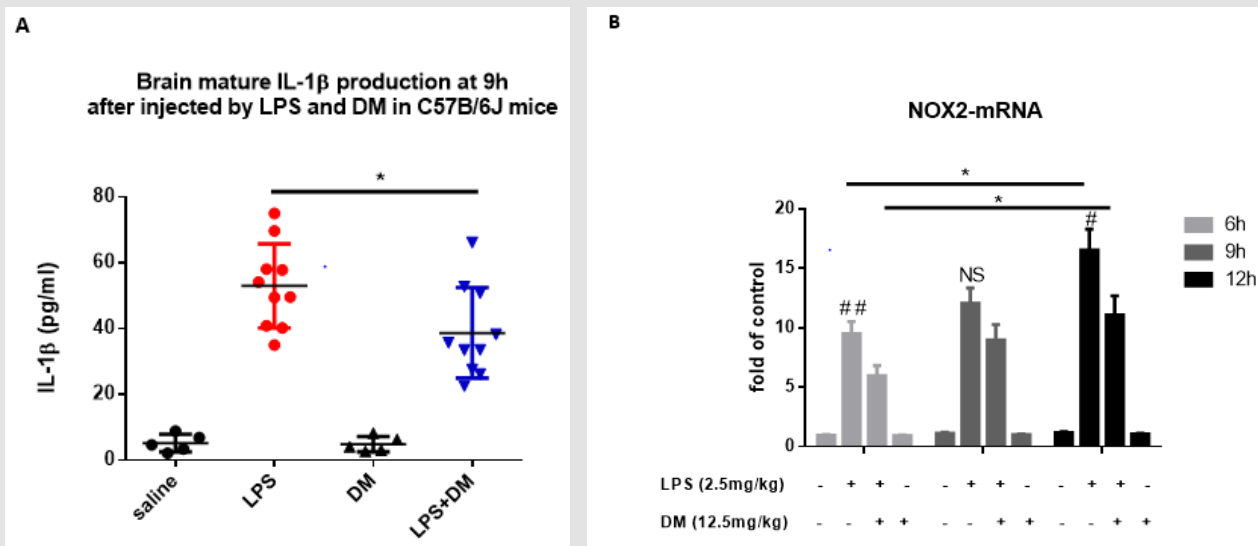


Figure 5: Post-treatment of dextromethorphan alleviated brain mature IL-1 β levels and NOX2 mRNA expression.
 a) Post-treatment of dextromethorphan alleviated brain mature IL-1 β levels. (LPS 2.5mg/kg) (n=5-10/group). * $p < 0.05$, the differences of IL-1 β concentrations between LPS-treated and post-treated dextromethorphan groups. One-way ANOVA followed by Tukey's multiple comparison test. ($F = 32.6$, $p < 0.0001$).
 b) (NOX2 mRNA levels were measured by qPCR at 6h, and 12h after LPS treatment. DM was post-added to mice at 3h and 9h after LPS treatment, respectively. # $P < 0.05$, ## $P < 0.01$ for the differences of NOX2 mRNA level between LPS-treated and post-treated DM groups at 6h and 12h, NS for 9h. One-way ANOVA followed by Tukey's multiple test ($F(3,32)_{6h} = 31.42$, $F(3,32)_{9h} = 29.57$, $F(3,32)_{12h} = 32.66$). ** $P < 0.001$, the differences of NOX2 mRNA level over time in both LPS-treated groups and post-treated DM groups. Repeated-measure ANOVA and multivariate ANOVA test the difference over time and differences between groups.

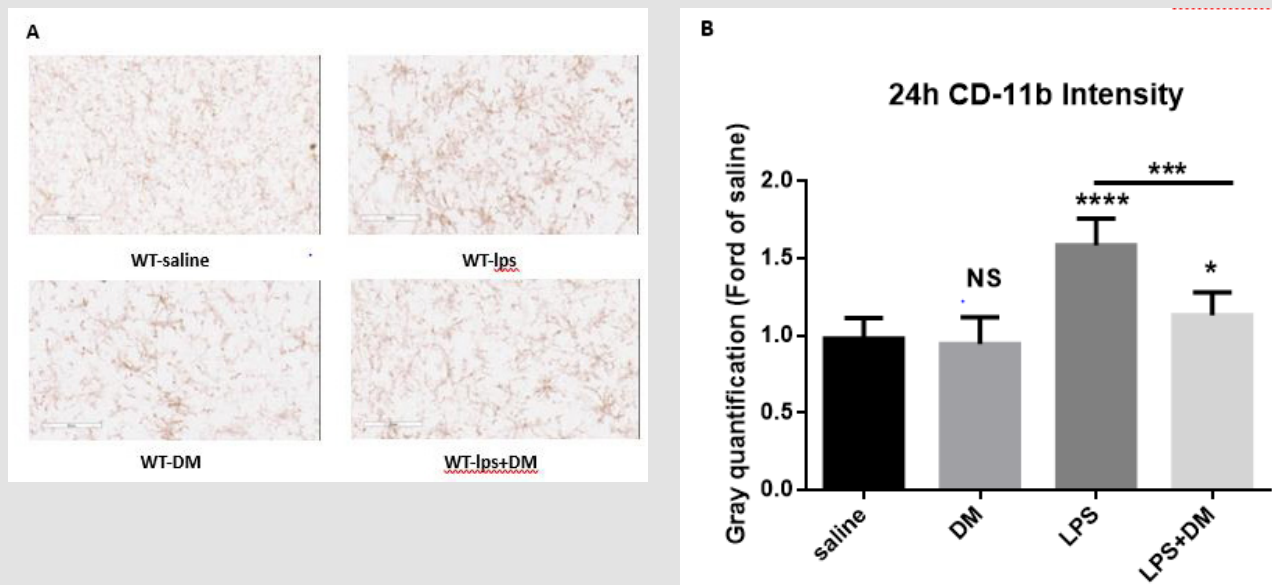


Figure 6: Post-treatment of dextromethorphan alleviated brain microglia activation in SN.

a) The microglia activation in SN was detected with CD11b IHC at 24h after LPS treatment. (n = 3/group) (LPS 2.5mg/kg, DM 12.5mg/kg). Representative images at 9h were shown (Figure 6A). Scale bar = 300 μ m.

b) CD-11b density was quantified. Results are expressed as folds of vehicle control. Histograms represent degree of microglial activation quantified by measuring the density of CD-11b staining with ImageJ. One-way ANOVA followed by Tukey's multiple comparison test. (F=62.73, p<0.0001). ****p<0.0001, ***p<0.001, the differences of gray quantification between LPS-treated and post-treated dextromethorphan groups. ****p<0.0001, *p<0.05 and NS compared to vehicle group.

Discussion

This study provides the first evidence indicating that microglial NOX2 plays a critical role in endotoxemia-related sickness behaviors. Mechanistically, inhibition of microglial activation and subsequent reduction in IL-1 β production through the inhibition of NOX2 underlie the amelioration of LPS-elicited sickness behaviors. This conclusion was based on both genetic knockout of NOX2 gene and pharmacological inhibition by DM. This study also suggests that microglial NOX2 can be a therapeutic target for developing anti sickness behaviors drug. In present study, we found that mice with NOX2 subunit knockout exhibited lower sickness behavior score than WT mice after LPS administration. Furthermore, compared to WT mice following LPS treatment, reduced activation of microglia (24h after LPS treatment) and decreased production of pro-inflammatory IL-1 β (9h after LPS treatment) were both observed in this study. Mechanistically, enhanced production of NOX2-generated superoxide and its related ROS is regarded as the major source of pathological oxidative stress in CNS leading to neuron damage, inflammation, and dysfunction.

In the current study, the NOX2 deficiency reduced acute production of IL-1 β and activation of microglia, indicating that the NOX2 deficiency could attenuate the LPS-induced acute symptom via reducing acute neuroinflammation. After elucidating the role

of microglial NOX2 in mediating LPS-induced sickness behaviors, as a proof of principle we further provide evidence showing high efficacy of DM in ameliorating sickness behaviors. In addition, post-treatment with DM following LPS also attenuated microglia activation, IL-1 β production, as well as NOX2 mRNA in vivo. Over decades, numerous studies demonstrate that DM, a non-opioid cough suppressant, has neuroprotective effects, which is attributed to its antagonistic effect on NMDA receptor [25-27]. However, we previously reported that anti-inflammatory effect of DM is attributed by inhibiting preventing microglia over-activation [28] induced by LPS and MPTP-elicited neurotoxin in neuron-glia culture and in vivo studies [29].

Furthermore, these mechanistic studies revealed that DM reduced the NOX2-generated superoxide free radicals from microglia, which in turn reduced in production of iROS and cytokines. The ill patients always have representative characters in behavior and/or emotion, including fever, fatigue, anorexia, fragmented sleep, losing interesting in physical and social environment, even showing emotion disorders such as depression. Over past two decades, a term of "sickness behavior" is suggested to describe such changes mentioned above in ill individuals or animals. Many studies have clarified the underlying mechanism that pro-inflammatory cytokines, such as IL-1 α , IL-1 β , TNF- α , IL-

6, play a critical role in sickness behavior and CNS dysfunction [30,31]. Despite moderate inflammation or sickness behavior is beneficial for coping with foreign microorganism-elicited infection, the uncontrolled cytokine storm or serious sickness behavior could impair the immunity system, interfere with inflammatory responses and produce further damage to patients.

Therefore, from clinical perspective, it is desirable for ill patients to control or relieve excessive inflammation and sickness behavior after proper anti infection therapy. Our results suggested that DM and NOX2 deficiency not only inhibited activation of microglia, reduced sickness behavior, but also suppressed NOX2 gene expression and production of IL-1 β , which is also a critical pro-inflammatory cytokine in neuroinflammation. IL-1 β plays a significant role in the initiation of inflammatory response in CNS and transition to chronic neuroinflammation induced by various stimulation [32]. IL-1 β could impair the integrity of BBB, leading to infiltration of peripheral immune cells into brain, and aggravating the neuroinflammation [33]. Meanwhile, IL-1 β activated microglia and astrocytes, which in turn generate other pro-inflammation cytokines, such as IL-18, TNF- α , IL-6 in CNS. Furthermore, the IL-1 β indirectly recruits leukocytes by augmenting the expression of chemokines.

These mechanistic studies indicated the IL-1 β is a critical inflammatory cytokine in both acute and chronic inflammatory response in CNS. Thus, this study not only demonstrates a beneficial effect of inhibiting NOX2 in mitigating the sickness behavior symptoms during acute inflammation, but also can be beneficial in preventing the transition from acute to chronic neuroinflammation. Over the years, a number of studies have reported that DM has neuroprotective effects [25,34-36]. The mechanism of neuroprotective activity has been attributed to DM's potential antagonistic effect on the NMDA receptor complex. Recently, studies indicated that DM also play neuroprotective roles in cognition deficit following systemic inflammation [37], together with previous reports [19], suggest that DM achieves these extraordinary protective effects through the inhibition of NADPH oxidase. Our study demonstrated DM and NOX2 deficiency attenuate the microglial activation during acute inflammation, meanwhile, the reduced IL-1 β production and NOX2 gene expression in brain also was observed. Subsequently, the sickness behavior induced by LPS relieved following acute inflammation.

This finding is similar with the protective effect of DM in chronic neuroinflammation [18,20]. Several studies have investigated the role of a variety of pro-inflammatory cytokines such as TNF- α , IL-1, IL-6, IL-8, IL-10 and IFN- γ and reactive oxygen species like superoxide and iROS in the pathogenesis of sepsis or infection. To relieve inflammation in septic or infected patients via blocking single cytokine (e.g., IL-1 β receptor antagonist), while the crosstalk in cytokine network makes this effort uncertain and controversy

[38-40]. The present study offers a novel approach by using DM to attenuate infection-related sickness behavior, which selectively targets NOX2. Furthermore, the findings from this study provide a potential therapeutic viewpoint: DM would be an alternative drug for septic or endotoxemia-related sickness behavior besides antibiotics and other life supportive therapy, such as ventilation, oxygen, fluid replacement therapy. In conclusion, DM attenuated sickness behavior in mice induced by LPS via inhibiting NOX2 activation and suppressing the production of IL-1 β in brain. The present findings demonstrated that NOX2 is a novel therapeutic target for sickness behavior. Furthermore, this study also suggests that owing to its excellent safety record and high efficacy, DM has a great potential to be tested in clinical settings.

Acknowledgments

We thank Dr. Hong, Jau-Shyong from the Neurobiology Laboratory at NIEHS for the help of experiments design and critical reading of the manuscript.

Funding

This research was supported by the Intramural Research Program of the NIH, National Institute of Environmental Health Sciences, USA (ZIA ES090082-22); as well as grants from the National Natural Science Foundation of China (81270144, 81570844, 81970083 and 30800507 to J.F.)

Availability of Data and Materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' Contributions

YL contributed to the study design, data collection, statistical analysis, data interpretation and manuscript preparation. ZZ and RZ contributed to data collection and statistical analysis. YBW and JF contributed to the study design, data collection and manuscript preparation. All authors have read and approved the final manuscript.

Competing Interests

The authors declare that they have no competing interests.

References

1. Sonnevile R, Verdonk F, Rauturier C, Klein IF, Wolff M, et al. (2013) Understanding brain dysfunction in sepsis. *Annals of Intensive Care* 3(1): 15-25.
2. Sonnevile R, den Hertog HM, Güiza F, Gunst J, Derese I, et al. (2012) Impact of hyperglycemia on neuropathological alterations during critical illness. *The Journal of Clinical Endocrinology and Metabolism* 97(6): 2113-2123.
3. Gao HM, Jiang J, Wilson B, Zhang W, Hong JS (2002) Microglial activation-mediated delayed and progressive degeneration of rat nigral dopaminergic neurons: relevance to Parkinson's disease. *Journal of neurochemistry* 81(6): 1285-1297.

4. Chen SH, Oyarzabal EA, Sung YF, Chu CH, Wang Q, et al. (2015) Microglial regulation of immunological and neuroprotective functions of astroglia. *Glia* 63(1): 118-131.
5. Gao HM, Hong JS (2008) Why neurodegenerative diseases are progressive: uncontrolled inflammation drives disease progression. *Trends Immunology* 29(8): 357-365.
6. Dantzer R, O'Connor JC, Freund GG, Johnson RW, Kelley KW (2008) From inflammation to sickness and depression: when the immune system subjugates the brain. *Nature Reviews Neuroscience* 9(1): 46-56.
7. Dantzer R (2009) Cytokine sickness behavior, and depression. *Immunology and Allergy Clinics of North America* 29(2): 247-264.
8. Hart BL (1988) Biological basis of the behavior of sick animals. *Neuroscience and Biobehavioral Reviews* 12(2): 23-37.
9. Suehiro E, Fujisawa H, Ito H, Ishikawa T, Maekawa T (1999) Brain temperature modifies glutamate neurotoxicity in vivo. *Journal of Neurotrauma* 16(4): 285-297.
10. Sharma HS, Hoopes PJ (2003) Hyperthermia induced pathophysiology of the central nervous system. *International Journal of Hyperthermia* 19(3): 325-343.
11. Konsman JP, Veeneman J, Combe C, Poole S, Luheshi GN, et al. (2008) Central nervous action of interleukin-1 mediates activation of limbic structures and behavioural depression in response to peripheral administration of bacterial lipopolysaccharide. *The European Journal of neuroscience* 28: 2499-2510.
12. Moon ML, McNeil LK, Freund GG (2011) Macrophages make me sick: how macrophage activation states influence sickness behavior. *Psychoneuroendocrinology* 36(10): 1431-1440.
13. Gayle DA, Ling Z, Tong C, Landers T, Lipton JW, et al. (2002) Lipopolysaccharide (LPS)-induced dopamine cell loss in culture: roles of tumor necrosis factor- α , interleukin-1 β , and nitric oxide. *Brain Research Developmental Brain Research* 133: 27-35.
14. Ferrari CC, Pott Godoy MC, Tarelli R, Chertoff M, Depino AM, et al. (2006) Progressive neurodegeneration and motor disabilities induced by chronic expression of IL-1 β in the substantia nigra. *Neurobiology of Disease* 24: 183-193.
15. Zhao Z, Wang Y, Zhou R, Li Y, Gao Y, et al. (2020) A novel role of NLRP3-generated IL-1 β in the acute-chronic transition of peripheral lipopolysaccharide-elicited neuroinflammation: implications for sepsis-associated neurodegeneration. *J Neuroinflammation* 17(1): 64-82.
16. Block ML, Zecca L, Hong JS (2007) Microglia-mediated neurotoxicity: uncovering the molecular mechanisms. *Nature Review Neuroscience* 8(1): 57-69.
17. Qin L, Liu Y, Wang T, Wei SJ, Block ML, et al. (2004) NADPH oxidase mediates lipopolysaccharide-induced neurotoxicity and proinflammatory gene expression in activated microglia. *Journal of Biological Chemistry* 279(2): 1415-1421.
18. Zhang W, Wang T, Qin L, Gao HM, Wilson B, et al. (2004) Neuroprotective effect of dextromethorphan in the MPTP Parkinson's disease model: role of NADPH oxidase. *FASEB Journal* 18: 589-591.
19. Li G, Liu Y, Tzeng NS, Cui G, Block ML, et al. (2005) Protective effect of dextromethorphan against endotoxic shock in mice. *Biochemical Pharmacology* 69(2): 233-240.
20. Liu Y, Qin L, Li G, Zhang W, An L (2003) Dextromethorphan protects dopaminergic neurons against inflammation-mediated degeneration through inhibition of microglial activation. *The Journal of Pharmacology and Experimental Therapeutics* 305(1): 212-218.
21. Wang QS, Chu CH, Li Q, Chen SH, Wilson B, et al. (2014) exacerbates dopaminergic neurodegeneration through neurokinin-1 receptor-independent activation of microglial NADPH oxidase. *Journal of Neuroscience* 34: 12490-12503.
22. Wang Q, Qian L, Chen SH, Chu CH, Wilson B, et al. (2015) post-treatment with an ultra-low dose of NADPH oxidase inhibitor diphenyleneiodonium attenuates disease progression in multiple Parkinson's disease models. *Brain* 138: 1247-1462.
23. Gassmann M, Grenacher B, Rohde B, Vogel J (2009) Quantifying Western blots: pitfalls of densitometry. *Electrophoresis* 30: 1845-1855.
24. Chechneva OV, Mayrhofer F, Daugherty DJ, Pleasure DE, Hong JS, et al. (2011) Low dose dextromethorphan attenuates moderate experimental autoimmune encephalomyelitis by inhibiting NOX2 and reducing peripheral immune cells infiltration in the spinal cord. *Neurobiology Disease* 44(1): 63-72.
25. Haberecht MF, Mitchell CK, Lo GJ, Redburn DA (1997) N-methyl-D-aspartate-mediated glutamate toxicity in the developing rabbit retina. *Journal of Neuroscience Research* 47: 416-426.
26. Chaplan SR, Malmberg AB, Yaksh TL (1997) Efficacy of spinal NMDA receptor antagonism in formalin hyperalgesia and nerve injury evoked allodynia in the rat. *Journal of Pharmacology Experimental Therapeutics* 280(2): 829-838.
27. Plesan A, Hoffmann O, Xu XJ, Wiesenfeld Hallin Z (1999) Genetic differences in the antinociceptive effect of morphine and its potentiation by dextromethorphan in rats. *Neuroscience Lett* 263: 53-56.
28. Liu Y, Qin L, Li G, Zhang W, An L, et al. (2003) Dextromethorphan protects dopaminergic neurons against inflammation-mediated degeneration through inhibition of microglial activation. *Journal of Pharmacology Experimental Therapeutics* 305: 212-218.
29. Zhang W, Wang T, Qin L, Gao HM, Wilson B, et al. (2004) Neuroprotective effect of dextromethorphan in the MPTP Parkinson's disease model: role of NADPH oxidase. *FASEB J* 18: 589-591.
30. Dantzer R, Bluthé RM, Gheusi G, Cremona S, Layé S, et al. (1998) Molecular basis of sickness behavior. *Ann N Y Acad Sci* 856: 132-138.
31. Henry CJ, Huang Y, Wynne AM, Godbout JP (2009) Peripheral lipopolysaccharide (LPS) challenge promotes microglial hyperactivity in aged mice that is associated with exaggerated induction of both pro-inflammatory IL-1 β and anti-inflammatory IL-10 cytokines. *Brain, Behavior and Immunity* 23(3): 309-317.
32. Cannon JG (2000) Inflammatory Cytokines in Nonpathological States. *News in Physiological Sciences* 15: 298-303.
33. Alvarez JI, Dodelet Devillers A, Kebir H, Ifergan I, Fabre PJ, et al. (2011) The Hedgehog pathway promotes blood-brain barrier integrity and CNS immune quiescence. *Science* 334: 1727-1731.
34. Choi DW (1987) Dextrorphan and dextromethorphan attenuate glutamate neurotoxicity. *Brain Research* 403(2): 333-336.
35. Steinberg GK, George CP, DeLaPaz R, Shibata DK, Gross T (1988) Dextromethorphan protects against cerebral injury following transient focal ischemia in rabbits. *Stroke* 19: 1112-1118.
36. Britton P, Lu XC, Laskosky MS, Tortella FC (1997) Dextromethorphan protects against cerebral injury following transient, but not permanent, focal ischemia in rats. *Life Sciences* 60: 1729-1740.
37. Huang WY, Liu KH, Lin S, Chen TY, Tseng CY, et al. (2020) NADPH oxidase 2 as a potential therapeutic target for protection against cognitive deficits following systemic inflammation in mice. *Brain Behavior and Immunity* 84: 242-252.
38. Van der Poll T, van de Veerdonk FL, Scicluna BP, Netea MG (2017) The immunopathology of sepsis and potential therapeutic targets. *Nat Rev Immunol* 17(7): 407-420.
39. Deutschman CS, Tracey KJ (2014) Sepsis: current dogma and new perspectives. *Immunity* 40(4): 463-475.
40. Vincent JL, Sun Q, Dubois MJ (2002) Clinical trials of immunomodulatory therapies in severe sepsis and septic shock. *Clin Infect Dis* 34(8): 1084-1093.

ISSN: 2574-1241

DOI: 10.26717/BJSTR.2021.38.006204

Yubao Wang, Jing Feng. Biomed J Sci & Tech Res



This work is licensed under Creative Commons Attribution 4.0 License

Submission Link: <https://biomedres.us/submit-manuscript.php>



Assets of Publishing with us

- Global archiving of articles
- Immediate, unrestricted online access
- Rigorous Peer Review Process
- Authors Retain Copyrights
- Unique DOI for all articles

<https://biomedres.us/>