

Genetic Engineering Application in Animal Breeding-Review

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Abbreviations: AI: Artificial Insemination; BST: Bovine Somatotropin; BT: Bacillus Thuringiensis; DNA: Deoxyribonucleic Acid; ESC: Embryonic Stem Cell; ESR: Estrogen Receptor; FDA: Food and Drug Administration; FECB: Boroola Fecundity; GH: Growth Hormone; GM: Genetically Modified; IVF: In Vitro Fertilization; IVM: In Vitro Maturation; MAS: Marker-Assisted Selection; rBGH: Recombinant Bovine Growth Hormone; rBST: Recombinant Bovine Somatotropin; TBA=To Be Announced; US: United States; VLPs: Virus Like Particles; WHO: World Health Organization

ABSTRACT

Genetic engineering is an integral approach to the development of new diagnostic techniques, drugs for human and animal diseases, foods for human health, development of tissues, and cells for xenotransplantation. The components of vaccine for disease control and nutraceuticals for human health by altering the food components or by introducing the health provide proteins, peptides and other components may be an integral part of human life in the coming days. Genetically engineered animals also offer significant human health and environmental benefits, livestock becomes more efficient for converting feed to animal protein and reducing waste production. Finally, genetic engineering will improve the welfare of animals by imparting resistance to disease and enhancing overall health and wellbeing. Genetic engineering, therefore, includes those in vitro techniques involved in the study of genes and their regulation, various techniques used in gene therapy, and the creation of novel strains of existing microorganisms for medical or industrial use. It also includes a group of techniques used for the modification of organisms. The techniques permit individuals or groups of genes to be isolated from large masses of DNA and produced in virtually unlimited quantities. This is through recombining DNA fragments from one organism and transferring them to another for expression. The hybrid molecule formed when a fragment of DNA from one organism is spliced to another DNA fragment is called recombinant DNA. Genetic engineering in animal production has a growing number of practical benefits, such as in the production of transgenic animals resistant to disease, increasing the productivity of animals, in the treatment of genetic disorders, and the production of vaccines.

Keywords: Gene therapy; Genetic Engineering; Transgenesis; Vaccines

Introduction

Genetic engineering is the name of a group of techniques used for direct genetic modification of organisms or populations of organisms using the recombination of DNA. These procedures are of use to identify, replicate, modify, and transfer the genetic material of cells, tissues, or complete organisms [1,2]. Most techniques are related to the direct manipulation of DNA oriented to the expression of particular genes. In a broader sense, genetic engineering involves the incorporation of DNA markers for selection

(marker-assisted selection, MAS), to increase the efficiency of the so-called 'traditional' methods of breeding based on phenotypic information. The most accepted purpose of genetic engineering is focused on the direct manipulation of DNA sequences. These techniques involve the capacity to isolate cut and transfer specific DNA pieces, corresponding to specific genes [3,4]. The mammalian genome has a larger size and has a more complex organization than in viruses, bacteria, and plants. Consequently, genetic modification of animals, using molecular genetics and recombinant DNA

technology is more difficult and costly than in simpler organisms. In mammals, techniques for reproductive manipulation of gametes and embryos such as obtaining of a completely new organism from adult differentiated cells (cloning), and procedures for artificial reproduction such as in vitro fertilization, embryo transfer, and artificial insemination, are frequently an important part of these processes [5]. The interest in genetic engineering of mammalian cells is based on the idea of, for example, use gene therapy to cure genetic diseases such as cystic fibrosis by replacing the damaged copies of the gene with normal ones in fetuses or infants (gene therapy) [1,6,7]. Hence, this aims to review the advances and applications in genetic engineering in animal breeding, including a description of the methods, some applications, and ethical issues.

Transgenic Animals

Transgenesis is a procedure in which a gene or part of a gene from one individual is incorporated into the genome of another one. Transgenic animals have any of these genetic modifications with potential use in studying mechanisms of gene function, changing attributes of the animal to synthesize proteins of high value, create models for human disease, or to improve productivity or disease resistance in animals [8-10]. In the early 80's, several research groups reported success in gene transfer and the development of transgenic mice [11,12]. The definition of transgenic animals has been extended to include animals that result from the molecular manipulation of endogenous genomic DNA, including all techniques from DNA microinjection to embryonic stem (ES) cell transfer and knockout mouse production [13]. Since the early 1980s, the production of transgenic mice by microinjection of DNA into the pro-nucleus of zygotes has been the most productive and widely used technique. Using transgenic technology in the mouse, such as antisense RNA encoding transgenesis, it is now possible to add a new gene to the genome, increase the level of expression or change the tissue specificity of expression of a gene, and decrease the level of synthesis of a specific protein. Removal or alteration of an existing gene via homologous recombination required the use of ES cells and was limited to the mouse until the advent of nuclear transfer cloning procedures [14].

Transgenic Methods

Microinjection of DNA and now nuclear transfer are two methods used to produce transgenic livestock successfully. The steps in the development of transgenic models are relatively straightforward. Once a specific fusion gene containing a promoter and the gene to be expressed has been cloned and characterized, sufficient quantities are isolated, purified, and tested in cell culture if possible and readied for preliminary mammalian gene transfer experiments. In contrast with nuclear transfer studies, DNA microinjection experiments were first performed in the mouse [1,15]. While nuclear transfer might be considered inefficient in its current form, major advances in experimental protocols can be anticipated. The

added possibility of gene targeting through nuclear transplantation opens up a host of applications, particularly with regard to the use of transgenic animals to produce human pharmaceuticals. The only major technological advance since the initial production of transgenic farm animals has been the development of methods for the in vitro maturation of oocytes (IVM), in vitro fertilization (IVF), and subsequent culture of injected embryos before transfer to recipient females [14,16].

DNA microinjection

The direct DNA microinjection into the pro-nuclei of embryos was the first technique that led to regular and relatively easy success in mammals [17]. In lower vertebrates and invertebrates, pro-nuclei are not a visible gene. Microinjection must therefore be performed in the cytoplasm, using much larger amounts of DNA. For unknown reasons, the success of this approach is quite variable from one species to another. It proved efficient in several fish species and mainly in salmonids [11,18]. It remains inefficient in the laboratory fish medaka as well as in *Xenopus* and chicken. In these species, foreign DNA usually does not integrate into the genome of the animals. In insects (*Drosophila*) and worms (*Caenorhabditiselegans*), foreign DNA is injected into gonad syncytium [19,20].

The use of transposons

Several transposons have been used successfully. Transposon has been used for years to generate transgenic *Drosophila* [21]. The mariner transposon originally found in *Drosophila* and adapted to different species is efficient to transfer genes in medaka [22], chicken [23], and mouse [24].

DNA transfer into gametes

One logical approach to generate transgenic animals may theoretically consist of introducing foreign DNA in gametes before fertilization. The yield of transgenic animals is usually low and largely unpredictable. Moreover, the integrated DNA is most of the time profoundly rearranged and no more functional. This phenomenon can seemingly be greatly attenuated by selecting the most appropriate ejaculates and by removing DNase by repeated washing and addition of DNase inhibitors [25]. Experiments aiming at transferring foreign genes into sperm precursors either in vivo or in vitro are in the course [25]. Although encouraging, the results obtained by different groups indicate that additional studies are required before this approach may become an attractive alternative to the other methods. The mechanism of gene transfer into epididymal spermatozoa by injection of a DNA- transfectant complex into testis is under study. This method permitted the generation of a limited number of transgenic animals so far [26].

The use of retroviral vectors

A family of vectors capable of infecting chicken primordial germ cells and of generating transgenic animals has been described [27].

Although laborious this method remains the only, which allowed repeatedly to transfer the foreign gene into the chicken. Recent unpublished work carried out in D. Baltimore laboratory has shown that lentiviral vectors transfer foreign genes with quite a high efficiency in a one-cell embryo. This approach appears much simpler than microinjection and might become used extensively in the future [28].

Gene Transfer using embryonic cells

It should be mentioned that in a work published recently, it was reported that appropriate vectors are capable of inducing targeted gene transfer into *Drosophila* by homologous recombination [29]. Two recent studies indicate that chicken cultured primordial germ cells retransferred to embryos can participate in their development and transmit their genes to progeny [30]. ES-like cells from medaka embryo capable of generating chimeric animals have also been described and are potential tools for gene transfer and targeting in fish [31].

Gene transfer By Nuclear Transfer

Interestingly, a recent study showed that the donor genome in bovine embryos generated by nuclear transfer from somatic cells is aberrantly methylated [32]. This may have inactivated genes required for embryo development. In a recent publication, it was reported that cloned sheep having the knocked-out gene did not survive after birth [33,34]. Gene addition by the cloning technique has been extended to goat [35] and cow [36,37] and pig [38]. Three groups obtained cloned pigs independently and by different methods. Gene knock-out was recently achieved in the pig. Moreover, cloning was successful in the rabbit [39].

Recombination of Genes Using Plasmids

Foreign DNA is inserted into host cells by combining the foreign DNA with the DNA of a vector. If the recombinant DNA gets inside a host cell, it can replicate along with the DNA of the host cell. This means that every time the host bacterium undergoes cell division, each new daughter cell receives a copy of the recombinant DNA, thus amplifying the recombinant DNA with each cell division [40]. In order to use a plasmid to insert foreign DNA into a bacterial cell, two steps are required: First, the foreign DNA must be combined with a plasmid. In the second step, a bacterial cell must absorb the recombinant plasmid. For the first step, restriction enzymes and DNA ligase are used. Restriction enzymes are naturally occurring enzymes that cut DNA. Many restriction enzymes are valuable tools in molecular biology. Each restriction enzyme cuts DNA only where a specific sequence of base pairs occurs. The broken bonds between the deoxyribose and phosphate groups that form the "side rails" of the DNA double helix (the phosphodiester linkages) must also be repaired. DNA ligase is the enzyme that catalyzes this reaction [11,40].

Future Perspectives of Transgenesis

The techniques for obtaining transgenic animals in species of agricultural interest are still inefficient. Some approaches that may overcome this problem are based on cloning strategies. Using these techniques it is feasible to reduce to less than 50% the number of embryo receptor females, which is one of the most important economic limiting factors in domestic species. It would also facilitate the further proliferation of transgenic animals. Recent results relate these techniques with still low success rates [41], high rates of perinatal mortality, and variable transgenic expression that requires to be evaluated before generalizing their application [8,42]. Considerable effort and time are required to propagate the transgenic animal genetics into commercial dairy herds. Rapid dissemination of the genetics of parental animals by nuclear transfer could result in the generation of mini herds in two to three years. However, the existing inefficiencies in nuclear transfer make this a difficult undertaking. It is noteworthy that the genetic merit of the 'cloned' animals will be fixed, while continuous genetic improvements will be introduced in commercial herds by using artificial insemination breeding programs [43].

In an alternative scenario of herd expansion, semen homozygous for the transgene may be available in four to five years. Extensive breeding programs will be critical in studying the interaction and co-adaptation of the transgene(s), with the background polygenes controlling milk production and composition. Controlling inbreeding and confirming the absence of deleterious traits so that the immediate genetic variability introduced by transgenesis is transformed into the greatest possible genetic progress is equally critical [11,15,43].

Application of Genetic Engineering

Gene Therapy

Gene therapy is a technique that may be used to change the genome (germ-line cells i.e., eggs or sperm) or the somatic cells of particular organs (*in vitro* or *in vivo*) of a developing or already developed organism. Changes made to somatic cells using gene therapy are not inherited by the organism's descendants [44]. Gene therapy uses a vector (most usually a disabled virus) to 'infect' target cells with the desired gene. Genetic engineering has successfully produced germ-line changes in marmoset monkeys. Gene therapy modifications, when conducted on germ-line cells, are also inherited by the organism's descendants. Using such techniques hereditary diseases could be cured and eliminated from the germ-line and the disease potentially eliminated from a species [44,45].

Increases Milk Composition

Milk proteins are coded by unique copy genes that can be altered to modify milk composition and properties. Among the

different applications of milk modification in transgenic animals [12]. To introduce the human lactoferrin into the bovine milk, transgenic cows have been obtained [46]. Lactoferrin is responsible for iron transport and inhibits bacterial growth. To express antibacterial substances in the milk, such as proteases to increase mastitis resistance. The objective is to alter the concentrations of antibacterial proteins such as lysozyme or transferrin in the milk [10,47].

Improving Wool Production

The objectives are to improve the production of sheep wool and to modify the properties of the fiber. Because cysteine seems to be the limiting amino acid for wool synthesis, the first approach was to increase its production through the transfer of cysteine biosynthesis from bacterial genes to sheep genome [12]. This approach did not achieve the efficient expression of these enzymes in the rumen of transgenic sheep. The control of the quality, color, yield, and ease of harvest of hair, wool, and fiber for fabric and yarn production has been an area of focus for genetic engineering in livestock. The manipulation of the quality, length, fineness, and crimp of the wool and hair fiber from sheep and goats has been examined using transgenic methods [48]. Transgenic methods will also allow improvements to fiber elasticity and strength. In the future transgenic manipulation of wool will focus on the surface of the fibers. Decreasing the surface interactions between fibers could decrease the shrinkage of garments made from such fibers [49].

Enhancing growth Rates and Carcass Composition

Using transgenic technology, it is possible to manipulate growth factors, growth factor receptors, and growth modulators. Transgenic sheep and pigs have been used to examine the postnatal growth of mammals. Growth hormone (GH) and IGF genes have been incorporated and expressed at various levels in genetically engineered animals [12]. Frequently the used promoters have not allowed an efficient control of the expression of the transgene. It was assessed that it is necessary to develop more complex constructions that activate or repress the expression of the transgene more precisely. Adams et al., [50] found inconsistent results regarding the effect of a growth hormone construct in sheep on growth and meat quality. Recently, a spectacular transformation was obtained by insertion of a plant gene in pigs generated transgenic pigs that carried the fatty acid desaturation 2 Gene for a 12 fatty acid desaturase from spinach [45].

Vaccine Production

Recombinant vaccines fall into three basic categories: live genetically modified organisms, recombinant inactivated vaccines, and genetic vaccines [51].

Live Genetically Modified Vaccine: The first category of live genetically modified vaccines can be viruses or bacteria with one or

more genes deleted or inactivated, or they can be vaccines carrying a foreign gene from another disease agent, which are referred to as vaccine vectors. Deletion or gene-inactivated vaccines are developed to attenuate the disease agent. Generally, two (double-knockout) or more genes are deleted or inactivated so the vaccine remains table and cannot revert to a pathogenic agent [52]. Foreign genes from disease agents have been inserted into potatoes, soybeans, and corn plants and fed to animals; the expressed proteins from those foreign genes immunized the animals against the disease agent [53].

Recombinant Inactivated Vaccine: Recombinant inactivated vaccines are subunit vaccines containing only part of the whole organism. Subunit vaccines are synthetic peptides that represent the most basic portion of a protein that induces an immune response. Subunit vaccines consist of whole proteins extracted from the disease agent or expressed from cloned genes in the laboratory. Several systems can be used to express recombinant proteins, including expression systems that are cell-free or that use whole cells. Whole-cell expression systems include prokaryotic (bacteria-based) systems such as *E. coli* and eukaryotic (mammalian, avian, insect, or yeast-based) systems. Another type of recombinant subunit vaccines, called virus-like particles (VLPs), can be created when one or more cloned genes that represent the structural proteins of a virus are expressed simultaneously and self-assemble into VLPs. These VLPs are immunogenic. Because subunit vaccines do not replicate in the host, they usually are administered with an adjuvant [11,54].

Genetic or DNA vaccine: The third category of recombinant vaccines is referred to as genetic vaccines because they are DNA alone. Genetic or DNA vaccines usually are circular pieces of DNA, called plasmids, which contain a foreign gene from a disease agent and a promoter that is used to initiate the expression of the protein from that gene in the target animal [55]. Plasmids can be maintained in bacteria (usually *E. coli*) and have been designed to accept foreign genes for expression in animals. Recombinant plasmids containing a foreign gene are purified from bacteria, and "naked" DNA is injected directly into an animal, intramuscularly, or intradermally [54].

Improving Reproductive Performance

Several genes that may profoundly affect reproductive performance were identified. These included the estrogen receptor (ESR) and the Boroola fecundity (FECB) genes. A specific form of the ESR gene is associated with 1.4 more pigs born per litter than is typical in lines of pigs that do not contain this specific ESR gene type [56]. The introduction of a mutated or polymorphic ESR gene could increase litter size in pigs. A single major gene for fecundity, the FECB gene that allows for increased ovulation rate was identified in Merino sheep [57]. Each copy of this gene increases ovulation rate by approximately 1.5 ova per cycle [58].

Increasing Resistance to Disease

Animal biotechnology offers several approaches to fight diseases in animals. Firstly, through genetic selection, livestock producers can select for certain traits that are associated with disease resistance and populations of animals that are less vulnerable to diseases could be developed. Secondly, through genetic engineering, breeders can integrate disease resistance genes from new sources, allowing for improved animal disease resistance benefits not only livestock producers and their animals, but consumers also benefit as a result of safer animal products in the market and a reduction in the incidence of human transmissible diseases such as avian influenza [59]. Increased disease resistance can be achieved by introducing resistance-conferring gene constructs into animals or by depleting a susceptibility gene or locus from the animal. Hence gain of function (additive), as well as loss of function (deletive, knockout) gene transfer experiments, can be used. Gene transfer experiments are often hampered by the lack of identified major genes or loci responsible for resistance traits [60].

Agricultural Applications

Genetic engineering was originally envisioned to have a multitude of agricultural applications. Recombinant bovine somatotropin (BST) derived from genetically engineered bacteria is one product of genetic engineering that is currently being used in animal agriculture. This protein, which increases milk production in lactating cows, is widely used throughout the U.S. dairy industry. Administering the protein rBST does not modify the DNA of the cow, and they do not become genetically engineered. BST was approved by the FDA in 1993 following extensive testing by numerous medical associations and scientific societies, which revealed no health or safety concerns for consumers [61].

Future of Genetic Engineering

A published report in California Agriculture entitled "Genetic engineering and cloning may improve milk, livestock production" [5]. The detailed potential uses of these biotechnologies and optimistically concluded that "by midcentury most agricultural animals will be genetically engineered to be more efficient and healthier than the current stock, producing healthy products for human consumption in an environmentally friendly system." While the technologies undoubtedly have the potential to deliver such benefits, no genetically engineered food animals are currently on the market, and the U.S. Food and Drug Administration (FDA) continues to call for a voluntary prohibition on the marketing of milk or meat from clones and their offspring. This review examines the scientific, regulatory, ethical, and public acceptance issues faced by the animal biotechnology industry and discusses the implications of the current climate on the future of animal biotechnology. Biotechnology is defined as technology based on biology. From this definition, it is obvious that livestock breeders have been practicing animal biotechnology for many years.

For example, traditional selection techniques involve using observations about the physical attributes and biological characteristics of an animal to select the parents of the next generation. Genetic improvement through selection, based on an increased understanding of population genetics and statistics, has been an important contributor to dramatic advances in agricultural productivity [62]. Many different biotechnologies have been incorporated into livestock breeding programs to accelerate the rate of genetic improvement. These include artificial insemination (AI), sire-testing programs using data collected from thousands of offspring, synchronization of estrus, embryo transfer, cryopreservation of gametes and embryos, and DNA-based marker-assisted selection of genetically superior animals [63].

Effect of Genetic Engineering

Effect on the Environment

Livestock farming is thought to be the major source of steroid hormones found in regional groundwater [64], several streams and rivers, and external surface water [65]. Beef cattle wastes are strongly androgenic [66]. Significant amounts of synthetic and natural hormones and their metabolites are excreted in animal waste [67]. Synthetic hormones excreted by animals are present in manure applied as fertilizer and in feedlot retention ponds, and from there they may be retained in soil or transported to the ground and surface water [68,69]. Lange et al., [70] calculated the number of beef cattle implanted with estrogens and androgens or progesterone, and the percent of applied hormones that reach the environment via cattle excrement. Commonly used androgenic growth promoter trenbolone has been found in groundwater near cattle feedlots, and that this growth promoter has androgenic effects. These numbers represent an increase in estrogens and androgens or progesterone over natural elimination rates.

Effects on Human

Humans are potentially exposed to synthetic hormones by consumption of commercial meat products and from environmental exposures related to animal waste [7,63]. Human exposure to both the synthetic and natural hormones causes cancer, reproductive effects, and other endocrine disruption outcomes. Estrogen is carcinogenic, anabolic steroids are reproductive toxicants and trenbolone is an anabolic steroid. TBA, zeranol, and MGA cross the placenta and are detectable in fetal tissues in rabbits [70] and are reflected in humans. Some evidence showed that xenobiotic growth promoters and their metabolites are thought to be genotoxic [71]. Veterinary use of hormones causes postmenopausal women, and pre-pubertal children, leaving them more vulnerable to the effects of exogenous hormone exposure [11,15].

Effects on the Animal Health

The use of Recombinant Bovine Growth Hormone (rBGH) had problems like mastitis, lameness, loss of condition, and lowered

immune system functions, which they attributed to rbST use [11,72].

Limitations of Genetic Engineering

Environmental Impacts

Potential risks for the environment include unintended effects on non-target organisms, ecosystems, and biodiversity. Insect-resistant GM crops have been developed by the expression of a variety of insecticidal toxins from the bacterium *B. thuringiensis* (Bt). Detrimental effects on beneficial insects, or a faster induction of resistant insects (depending on the specific characteristics of the *Bacillus thuringiensis* proteins, expression in pollen and areas of cultivation), have been considered in the area of several insect-protected GM crops [73].

Impacts on Human and Animal Health

Healthcare research is the most well-known purpose for which animals are genetically engineered. Through animal genetic engineering, scientists have made breakthroughs in organ transplantation, cancer research, and other areas. Similarities between the genomes of humans and other animals also suggest that future genetic research on animals will yield additional benefits for humans. In the future, kidney, heart, and lung failure patients will likely benefit from animal organ transplants. Xenotransplantation is the procedure of transplanting organs from one species to another. Although xenotransplantation is not new, its use to solve immunological problems such as transplant rejection began recently. Some believed that animal organ transplantation may be able to solve the organ shortage problem [74]. The application of modern biotechnology to food production presents new opportunities and challenges for human health and development. Recombinant gene technology, the most well-known modern biotechnology, enables plants, animals, and microorganisms to be genetically modified (GM) with novel traits beyond what is possible through traditional breeding and selection technologies [15].

Religious, Cultural, and Ethical Issues

The current and potential impact of rapid developments in biotechnology to effect innovations in medicine and drug development, as well as such diverse areas as crime detection, agriculture, pollution control, and industrial processes, brings into question how these techniques can be used constructively without damaging the cornerstones of religious ethics, namely respect for human life [75]. Revolutionary innovations in genetic engineering, the decoding of the human genome now make it possible for vegetables in our food chain to bear animal transgenes [76]. Public opinion surveys have reported that some people are ethically uncomfortable with the idea of genetically engineering animals. There are two central ethical concerns associated with the genetic engineering of animals. The first has to do with breaching species

barriers or playing God. Proponents of this view suggest that life should not be regarded solely as if it were a chemical product subject to genetic alteration and patentable for economic benefit. The second major ethical concern is that the genetic engineering of animals interferes with the integrity or telos of the animal. Telos is defined as "the set of needs and interests which are genetically based, and environmentally expressed, and which collectively constitute or define the form of life or way of living exhibited by that animal, and whose fulfillment or thwarting matter that animal" [20].

Economic Impact

The requirement of adequate infrastructure is a critical factor for the establishment of biotechnology companies. While research and development budgets for biotechnology research are only beginning to pick up in developing countries, start-up funding for biotechnology companies is still very rare to come by. Financing early stages of business development could be achieved through seed funding, easier access to loans, and venture funds [77].

Recombinant DNA Technology in Ethiopia

Ethiopia is an agrarian country that can have enormous benefit from the applications of biotechnology for increasing its agricultural productivity. The country is at the initial stages of research and development in agricultural biotechnology. Livestock related applications include artificial insemination, molecular diagnostics, vaccine production, and molecular genetic analysis. Infrastructure and skills in recombinant DNA and other cutting edge technologies such as proteomics and bioinformatics are not available. Ethiopia has recently given due emphasis to capacity building in agricultural biotechnology extending from promoting research, development, and education in various public institutions to the set up of an independent agricultural biotechnology research center. The constraints holding back progress in agricultural biotechnology are numerous ranging from poor technical and regulatory capacity to lack of appreciation of opportunities provided by agro-biotechnology by the public and decision-makers [78]. The Ethiopian Ministry of Agriculture is seeking to advance the capacity of various Ethiopian agricultural research institutes in the field of biotechnology. The Ministry requested a set of guidelines that will aid to direct national and international resources to establish a platform of technologies in the areas of molecular genetics, cell and tissue culture, and genetic engineering. These technologies are expected to increase productivity and raise the competitive marketability of Ethiopian agricultural products in national and international trade [11].

Conclusion and Recommendation

Animal genetic engineering is here to stay. Genetic engineering plays an essential role in healthcare research and food production. It has resulted in huge profits for a variety of industries. Genetic

engineering can also assist in sustaining species that are facing possible extinction. The current state and federal laws on animal rights suggest that all justifiable animal genetic engineering should be permitted. Free market environmentalism will test this hypothesis by allowing individuals, interest groups, and government agencies to use the market to indicate the optimal level for genetic engineering regulation. Genetic engineering if executed judiciously may provide practical benefits to mankind, as we have seen from other fundamental advances in life science. These benefits may be in terms of improvement in human health through the production of novel replacement proteins, drugs, vaccines, research models and tissues for the treatment and prevention of human disease, genetically engineered animals for improvement of environment and human health, improved in food production traits enabling them to help meet the global demand for more efficient, higher quality and lower-cost sources of food. It may also be beneficial to animal health, well-being, and animal welfare and some beneficial high-value industrial products can also be produced such as spider silk used for medical and defense purposes. However, the practical benefits of this technology have not yet reached to the consumers due to the broader gap between myths and the reality of genetic engineering technology. Therefore, there is an urgent need to formulate the regulatory framework with predictable, rigorous, science-based monitoring, and authentication of the technology to deliver practical benefits through the science of genetic engineering.

References

- Izquierdo Rojo M (2001) Genetic engineering and gene transfer: Madrid: Pirámide, Spain
- Karp G (2002) Cell and Molecular Biology, Concepts and Experiments. 3rd. New York: J Wiley 21(785): 48.
- Klug WS, Cummings MR (2006) Concepts of genetics: Pearson Education, Upper Saddle River, NJ, USA.
- Lewin B, Lewin B (2000) Genes VII: Oxford University Press, New York, USA.
- Murray J, Anderson G (2000) Genetic engineering and cloning may improve milk, livestock production. California Agriculture 54(4): 57-65.
- Coutelle C, Rodeck C (2002) On the scientific and ethical issues of fetal somatic gene therapy. Gene Therapy 9(11): 670-673.
- (2001) International Human Genome Sequencing Consortium. Initial sequencing and analysis of the human genome. Nature 409: 860-921.
- Houdebine LM (2002) Transgenesis to improve animal production. Livestock Production Science 74(3): 255-268.
- Chien KR (1996) Genes and physiology: molecular physiology in genetically engineered animals. The Journal of clinical investigation 97(4): 901-909.
- Felmer R (2004) Animales transgénicos: pasado, presente y futuro. Archivos de medicina veterinaria 36(2): 105-117.
- Asaye M, Biyazen H, Girma M (2014) Genetic engineering in animal production: Applications and prospects. Biochemistry and Biotechnology Research 2(2): 12-22.
- Murray JD, Anderson G, Oberbauer A, Mc Gloughlin M (1999) Transgenic animals in agriculture. CABI Publishing.
- Cameron E, Harvey M, Onions D (1994) Transgenic science. British Veterinary Journal 150(1): 9-24.
- Rao D, Nra, Iga, Itovic (2000) Introduction to transgenesis, an Overview. Proceedings of the 7th International Conference on Goats.
- Murray JD, Maga EA (2016) Genetically engineered livestock for agriculture: a generation after the first transgenic animal research conference. Transgenic research 25(3): 321-327.
- Wall R (2002) New gene transfer methods. Theriogenology 57(1): 189-201.
- Krimpenfort P, Rademakers A, Eyestone W, Van der Schans A, Van den Broek S, et al. (1991) Generation of transgenic dairy cattle using 'in vitro' embryo production. Bio/technology 9(9): 844-847.
- Devlin R, Biagi C, Smailus D (1997) Production and evaluation of transgenic salmonids. Proceedings of a Conference on transgenic Animals in Agriculture, Granlibaken, California, USA.
- Thierry Mieg D, Naert K, Bonnerot C (1997) Genetic transformation of *Caenorhabditis elegans*. Transgenic Animal Generation and Use: Harwood Academic Publishers Amsterdam, pp. 137-151.
- Van Eenennaam A (2008) Genetically Engineered Animals: An Overview. Department of Animal Science, University of California, Davis, California, USA.
- Kayser K (1997) Gene transfer in *Drosophila melanogaster*. Transgenic Animal Generation and Use: Harwood Academic Publishers, Amsterdam, Netherlands, pp. 133-137.
- Hackett PB, Ekker SC, Largaespada DA, Mc Ivor RS (2005) Sleeping Beauty transposon-mediated gene therapy for prolonged expression. Advances in genetics 54: 189-232.
- Sherman A, Dawson A, Mather C, Gilhooley H, Li Y, et al. (1998) Transposition of the *Drosophila* element mariner into the chicken germ line. Nature biotechnology 16(11): 1050-1053.
- Dupuy AJ, Clark K, Carlson CM, Fritz S, Davidson AE, et al. (2002) Mammalian germ-line transgenesis by transposition. Proceedings of the National Academy of Sciences 99(7): 4495-4499.
- Baccetti B, Spadafora C (2000) Proceedings of the workshop on sperm-mediated gene transfer: advances in sperm cell research and applications, Siena, Italy, 23-5, Molecular reproduction and development 56(2): 227-330.
- Sato M, Ishikawa A, Kimura M (2002) Direct injection of foreign DNA into mouse testis as a possible in vivo gene transfer system via epididymal spermatozoa. Molecular Reproduction and Development 61(1): 49-56.
- Ronfort C (1997) The use of retroviral vectors for gene transfer into bird embryo. Transgenic Animals: Generation and Use.
- Lois C, Hong EJ, Pease S, Brown EJ, Baltimore D (2002) Germline transmission and tissue-specific expression of transgenes delivered by lentiviral vectors. Science 295(5556): 868-872.
- Bernards A, Hariharan IK (2001) Of flies and men-studying human disease in *Drosophila*. Current opinion in genetics & development 11(3): 274-278.
- Petitte J, Yang Z, Liu G, Kulik M, Borwornpinyo S (2002) Primordial germ cells, embryonic stem cells and transgenic poultry. Transgenic Research 11(1): 79-80.
- Collodi P, Fan L (2002) Embryo cell cultures for cell-mediated gene transfer and the production of transgenic zebrafish. Transgenic Research 11(1).
- Kang Y K, Koo D B, Park J S, Choi Y H, Chung A S, et al. (2001) Aberrant methylation of donor genome in cloned bovine embryos. Nature genetics 28(2): 173-177.
- Denning C, Burl S, Ainslie A, Bracken J, Dinnyes A, et al. (2001) Deletion of the α (1, 3) galactosyl transferase (GGTA1) gene and the prion protein (PrP) gene in sheep. Nature biotechnology 19(6): 559-562.

34. Wells K (2001) Toward knockout sheep. *Nature biotechnology* 19(6): 529-530.
35. Reggio BC, James AN, Green HL, Gavin WG, Behboodi E, et al. (2001) Cloned transgenic offspring resulting from somatic cell nuclear transfer in the goat: oocytes derived from both follicle-stimulating hormone-stimulated and nonstimulated abattoir-derived ovaries. *Biology of Reproduction* 65(5): 1528-1533.
36. Arat S, Rzucidlo SJ, Gibbons J, Miyoshi K, Stice SL (2001) Production of transgenic bovine embryos by transfer of transfected granulosa cells into enucleated oocytes. *Molecular Reproduction and Development: Incorporating Gamete Research* 60(1): 20-26.
37. Zakhartchenko V, Mueller S, Alberio R, Schernthaner W, Stojkovic M, et al. (2001) Nuclear transfer in cattle with non-transfected and transfected fetal or cloned transgenic fetal and postnatal fibroblasts. *Molecular Reproduction and Development: Incorporating Gamete Research* 60(3): 362-369.
38. Dai Y, Vaught TD, Boone J, Chen SH, Phelps CJ, et al. (2002) Targeted disruption of the $\alpha 1$, 3-galactosyltransferase gene in cloned pigs. *Nature biotechnology* 20(3): 251-255.
39. Chesné P, Adenot PG, Viglietta C, Baratte M, Boulanger L, et al. (2002) Cloned rabbits produced by nuclear transfer from adult somatic cells. *Nature biotechnology* 20(4): 366-369.
40. (2006) Austin Community College. *Laboratory Manual*, (12th Edn). Texas, United States of America, USA.
41. Edwards J, Schrick F, Mc Cracken M, Van Amstel S, Hopkins F, et al. (2003) Cloning adult farm animals: a review of the possibilities and problems associated with somatic cell nuclear transfer. *American Journal of Reproductive Immunology* 50(2): 113-123.
42. Samiec M, Skrzyszowska M (2005) Advantages and perspectives of using somatic cloning for production of transgenic animals. *Medycyna Weterynaryjna* 61(1): 24-28.
43. Karatzas CN (2003) Designer milk from transgenic clones. *Nature biotechnology* 21(2): 138-139.
44. (2010) What is gene therapy? *Gene Therapy Net*.
45. Sasaki E, Suemizu H, Shimada A, Hanazawa K, Oiwa R, et al. (2009) Generation of transgenic non-human primates with germline transmission. *Nature* 459(7246): 523-527.
46. Van Berkel PH, Welling MM, Geerts M, Van Veen HA, Ravensbergen B, et al. (2002) Large scale production of recombinant human lactoferrin in the milk of transgenic cows. *Nature biotechnology* 20(5): 484-487.
47. Kerr D, Wellnitz O (2003) Mammary expression of new genes to combat mastitis. *Journal of Animal Science* 81(15_suppl_3): 38-47.
48. Powell B, Walker S, Bawden C, Sivaprasad A, Rogers G (1994) Transgenic sheep and wool growth: possibilities and current status. *Reproduction, fertility and development* 6(5): 615-623.
49. Bawden C, Dunn S, Mc Laughlan J, Nesci A, Powell B, et al. (1999) Transgenesis with ovine keratin genes: expression in the sheep wool follicle for fibres with new properties. *Transgenic Res* 8: s474.
50. Adams N, Briegel J, Ward K (2002) The impact of a transgene for ovine growth hormone on the performance of two breeds of sheep. *Journal of Animal Science* 80(9): 2325-2333.
51. Ellis RW (1999) New technologies for making vaccines. *Vaccine* 17(13-14): 1596-1604.
52. Uzzau S, Marogna G, Leori GS, Curtiss R, Schianchi G, et al. (2005) Virulence attenuation and live vaccine potential of aroA, crp cdt cya, and plasmid-cured mutants of *Salmonella enterica* serovar Abortusovis in mice and sheep. *Infection and immunity* 73(7): 4302-4308.
53. Streatfield S (2005) Plant-based vaccines for animal health. *Revue scientifique et technique (International Office of Epizootics)* 24(1): 189-199.
54. Jackwood M, Hickie L, Kapil S, Silva R, Osterrieder K, et al. (2008) Vaccine development using recombinant DNA technology. *Council Agric Sci Technol* 38: 1-11.
55. Rodriguez F, Whitton JL (2000) Enhancing DNA immunization. *Virology* 268(2): 233-238.
56. Rothschild M, Jacobson C, Vaske D, Tuggle C, Short T, et al. (1994) A major gene for litter size in pigs. *Proceedings of the 5th world congress on genetics applied to livestock production*, Guelph: University of Guelph, Canada.
57. Piper L, Bindon B, Davis G (1985) The single gene inheritance of the high litter size of the Booroola Merino. *Land RB and Robinson D W. Genetics of Reproduction in Sheep (C)*, Butterworths, London, UK, 19(8): 5.
58. Gottlieb S, Wheeler MB (2007) Genetically engineered animals and public health compelling benefits for health care, nutrition, the environment, and animal welfare.
59. Pohlmeier B, Van Eenennaam A (2009) Potential effects of biotechnology on animal health and well-being. *Applications of Animal Biotechnology in Animal Health* p. 1-5.
60. Müller M, Brem G (1998) Transgenic approaches to the increase of disease resistance in farm animals. *Revue scientifique et technique (International Office of Epizootics)* 17(1): 365.
61. Bauman D (1999) Bovine somatotropin and lactation: from basic science to commercial application. *Domestic animal endocrinology* 17(2-3): 101-116.
62. Dekkers JC, Hospital F (2002) The use of molecular genetics in the improvement of agricultural populations. *Nature Reviews Genetics* 3(1): 22-32.
63. (2002) National Research Council. *Animal biotechnology: science-based concerns*: National Academies Press.
64. Peterson E, Davis R, Orndorff H (2000) 17 β -Estradiol as an Indicator of Animal Waste Contamination in Mantled Karst Aquifers. *Journal of Environmental Quality* 29(3): 826-834.
65. Kolpin DW, Furlong ET, Meyer MT, Thurman EM, Zaugg SD, et al. (2002) Pharmaceuticals, hormones, and other organic wastewater contaminants in US streams, 1999– 2000: A national reconnaissance. *Environmental science & technology* 36(6): 1202-1211.
66. Durhan EJ, Lambright CS, Makynen EA, Lazorchak J, Hartig PC, et al. (2006) Identification of metabolites of trenbolone acetate in androgenic runoff from a beef feedlot. *Environmental health perspectives* 114(Suppl 1): 65-68.
67. Kolok AS, Sellin MK (2008) The environmental impact of growth-promoting compounds employed by the United States beef cattle industry: History, current knowledge, and future directions. *Reviews of environmental contamination and toxicology*: Springer p. 1-30.
68. Khan B, Lee LS, Sassman SA (2008) Degradation of synthetic androgens 17 α -and 17 β -trenbolone and trenbolone in agricultural soils. *Environmental science & technology* 42(10): 3570-3574.
69. Khan S, Roser D, Davies C, Peters G, Stuetz R, et al. (2008) Chemical contaminants in feedlot wastes: concentrations, effects and attenuation. *Environment international* 34(6): 839-859.
70. Lange I, Daxenberger A, Meyer H, Rajpert De Meyts E, Skakkebaek N, et al. (2002) Quantitative assessment of foetal exposure to trenbolone acetate, zeranol and melengestrol acetate, following maternal dosing in rabbits. *Xenobiotica* 32(8): 641-651.
71. Metzler M, Pfeiffer E (2001) Genotoxic potential of xenobiotic growth promoters and their metabolites. *Note. Apmis* 109(2): 89-95.
72. An H, Butler LJ (2009) Update on rBST use in the California dairy industry. *Agricultural and Resource Economics Update* 12(5): 5-7.

73. Sears MK (2003) Impact of *Bacillus thuringiensis* corn pollen on monarch butterfly populations: A risk assessment. ACS Publications.
74. West C (2005) Economics and Ethics in the Genetic Engineering of Animals. *Harv JL & Tech* 19: 413.
75. Curran G, Koszarycz Y (2004) Genetic engineering: Creating an ethical framework. *Australian eJournal of Theology* 2: 1-13.
76. Brunk C, Coward H (2009) *Acceptable genes? religious traditions and genetically modified foods*: SUNY Press.
77. Tonukari NJ (2004) Fostering biotechnology entrepreneurship in developing countries. *African Journal of Biotechnology* 3(6): 299-301.
78. Abraham A (2009) Agricultural biotechnology research and development in Ethiopia. *African Journal of Biotechnology* 8(25).

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