

Development of HPLC Method for Estimation of Darunavir Related Substance in Formulation

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ABSTRACT

A simple, specific and validated reverse phase high performance liquid chromatography method was developed for determination of drug related substances (impurity) of Darunavir in formulation. Method development includes optimization of stationary phase (column) and mobile phase flow rate for the resolution of six known impurities and one unknown impurity and Darunavir. Development of HPLC method for estimation of Darunavir related substance in formulation was carried out on Zorbax SB-C8, 250 x 4.6mm, 5µm by using a gradient mode of the mobile phase flow rate at 1ml/min with the same mobile phase as, ACN : Buffer pH 4.0 (Mobile phase B: Mobile Phase A). The stability-indicating capability of the method was proved through the solution state stability, solid state study and it was concluded that purity threshold was found to be greater than purity angle. A good linear relation exists over the range of 50 to 150 % specification level with a correlation coefficient value of 0.9998. Validation of the method was successfully established by performing various validation parameters such as specificity, precision, linearity, accuracy, LOD, LOQ, robustness, ruggedness according to ICH guidelines.

Introduction

International conference for harmonization (ICH) provides regulatory guidelines, such as ICH Q3A guidelines for the identification, quantification of impurities in new drug as well as pharmaceutical formulation. Therefore, it is necessary to conduct the impurity profiling to give documentary evidence that these impurities are present within the specified limits as predefined by the toxicological studies and to limit or to decrease the levels of these impurities in the formulation. For this, various analytical methods are available for the quantitative determination of impurities even at trace levels, which in turn ensures the consistent quality output and also safety & efficacy of drug therapy [1]. Generally forced degradation/stress testing is used to generate the samples for stability-indicating assay methods. Forced degradation/stress testing is defined as “the stability testing of drug substance and drug product under conditions exceeding those used for accelerated stability testing” [2]. Degradation can be achieved by exposing the drug, for extended periods of time, to extremes of pH (HCl or NaOH solutions of different strengths), at elevated

temperature, to hydrogen peroxide at room temperature, UV light, and to dry heat (in an oven) to achieve degradation to an extent of 5–20% [3,4]. Nowadays stability indicating methods had the main focus on the estimation of the amount of drug substance present in the marketed preparations. So, such type of testing methods is known as specific stability testing methods.

These methods involve the estimation of the amount of drug substance in the presence of other impurities, excipients, degradation products without their separations. The presence of other impurities and degradation products of drug substance may be responsible for the adverse effects of drug products. It makes the given drug product unsafe to use clinically [5]. ICH guidelines have guided the preparation of stability testing protocols for registration of new drug substance and product in USA, Japan, and European Union countries. It provides the requirement of three types of stability testing such as long term stability studies, accelerated stability study and intermediate stability study for four climatic zones. ICH guidelines also guide qualification, identification,

and quantification of impurities in drug substances and drug products to determine its thresholds [6,7]. Knowing these USFDA regulatory requirements and need of impurity / drug related impurity profiling [8], Darunavir drug had been selected for our current research work. Various new protease inhibitor drugs used to treat HIV infection. Among them, Darunavir is a second-generation protease inhibitor (PIs), designed specifically to overcome problems (Drug resistance problems) with the older agents in this class [9]. Darunavir drug sold under various brand names

(dose:100mg/400mg) [10]. It acts on the HIV-Aspartyl protease enzyme which is needed by virus to cleave the HIV polypeptide into its functional fragments. The chemical name of darunavir [11] is (3R, 3aS, 6aR)-hexahydrofuro [2,3-b]furan-3-yl(1S,2R)-3-[[[4-amino phenyl] sulfonyl] (2-methylpropylamino) -1 -benzyl-2-hydroxypropyl] carbamate corresponding to the molecular formula $C_{27}H_{37}N_3O_7S$ and a relative molecular mass of 547.68g/mol (Figure 1).

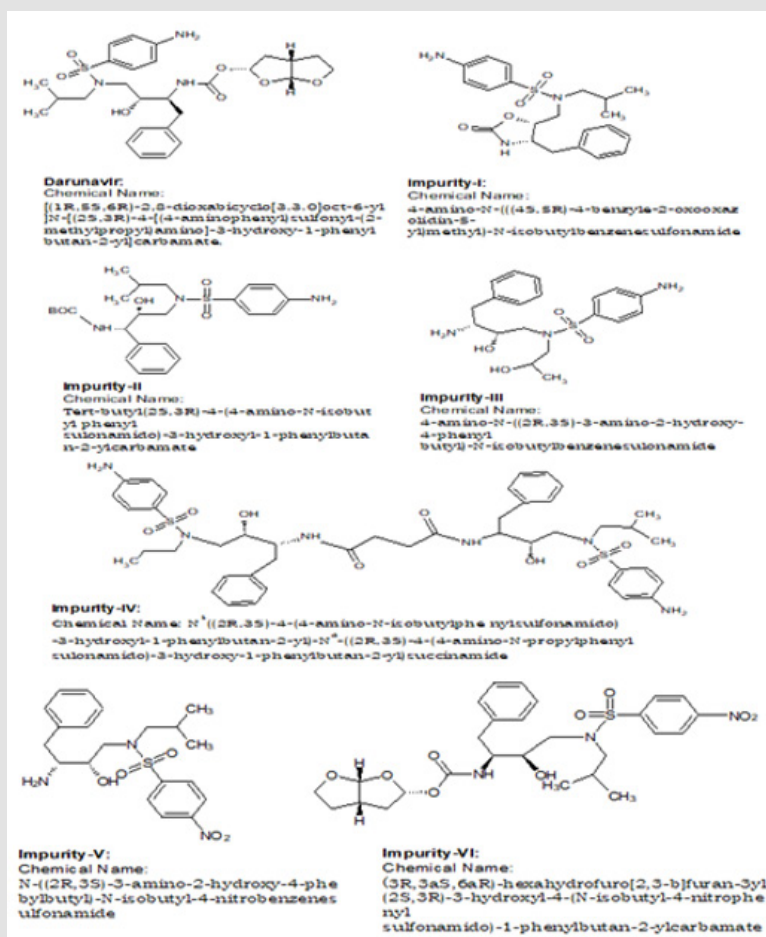


Figure 1: Structure of Darunavir and Darunavir impurities-I,II,III,IV,V and VI.

Darunavir exists as polymorphs in the form of Darunavir amorphous, Solvates and Hydrates. The ethanolate and hydrate forms exist in the form of crystals, whereas the non-solvated form is amorphous. When in the formed crystalline forms exist solvent molecules, water, salt, excipient or impurity has a pseudo polymorphic behavior and it can occur during handling, processing and storing. The tendency of a molecule to form solvates is related to the molecular structure, standards of hydrogen bonding and crystal packing. The solvent serves to stabilize the structure and desolvation process results in the formation of an amorphous form [12]. Literature survey reveals that Stability-indicating HPLC [13], RP-HPLC [14-16], IR [17], LC-MS [18] and UV Spectrophotometric methods [19] are reported for determination of darunavir as ethanolate form but no methods are reported for determination of

darunavir in its amorphous form. Hence the present work describes the development and validation of HPLC method for its assay and related substance determination in amorphous form of the drug.

Materials and Methods

Apparatus

The analysis was carried out on a HPLC-Jasco LC-Net II/ADC equipped with Quaternary Gradient pump PU-2089 plus, multi-wavelength detector MD-2010 plus with manual injector with 20 μ L loop and a reserved phase ACE C₁₈ column (150x4.6mm, 5 μ) with pore size of 100Å was used for chromatographic studies, Jasco v-630 -UV-visible spectrophotometer, pH meter EI, Model No. 1102012, Stability Chamber -THERMOLAB, Model No.:TS

00002008, Membrane filters with cellulose filter paper of 0.45µm, Sonicator model PCI Mumbai, Model No. 3.5L 100H, Weighing balance: Shimadzu AUX220 and Calibrated glassware's were used throughout experimental work.

Chemicals and Solvents

Acetonitrile and Methanol (HPLC Grade), Hydrochloric acid, Sodium hydroxide, Hydrogen peroxide, Pot. Dihydrogen phosphate, O-phosphoric acid of analytical grade. Distilled water was used throughout the experimental study.

Experimental

Sample preparations for Development of HPLC Method for Darunavir Related Substance Estimation in Formulation

Preparation of Solutions: (Diluent:- Methanol)

i. **Preparation of Standard Solution of Darunavir:** A standard solution of darunavir 0.24mg/mL was prepared using methanol as diluent. Further 1.0mL of this solution was diluted to 100.0mL in volumetric flask with diluent and mixed. (Concentration of Darunavir: 2.4µg/mL)

ii. **Preparation of Impurity stock solution (One solution for each Impurity):** Standard impurities DAR-I, DAR-II, DAR-III, DAR-IV, DAR-V and DAR-VI were weighed separately and transferred to a series of 50.0mL volumetric flasks and diluent was added, sonicated to dissolve and volume were made up to the mark with diluent and mixed. (Concentration of about each Impurity: 80µg/mL).

iii. **Preparation of Darunavir impurity spiked solution:** An accurately weighed quantity about 120.0mg of Darunavir standard was transferred to 100.0mL volumetric flask, about 50.0mL of diluent was added, sonicated to dissolve and cooled, from each impurity stock solution DAR-I, DAR-II, DAR-III, DAR-IV, DAR-V, DAR-VI 3.0mL was added and volume made up to the mark with diluent. (Concentration of DAR-I, DAR-II, DAR-III, DAR-IV, DAR-V and DAR-VI Impurity 2.4µg/mL and Darunavir 1200 µg/mL).

iv. **Preparation of Buffer Solution pH 4.0:** An accurately weighed quantity about 1.36gm. of potassium dihydrogen phosphate in 1000mL of water and pH adjusted to 4.0 ± 0.05 with Ortho-phosphoric acid, it was then filtered through 0.45µm nylon membrane filter.

v. Preparation of Mobile Phase:

a. Mobile phase A: Potassium dihydrogen phosphate Buffer pH 4.0

b. Mobile phase B: 100% Acetonitrile

vi. **Selection of Mobile Phase:** Mobile phase was selected on trial and error basis. Using following chromatographic parameters,

various trials were carried out to select suitable gradient program of mobile phase for resolution of impurities.

a. Column :- Zorbax SB-C8, 250 x 4.6mm, 5µm.

b. Detection Wavelength :- 265nm

c. Temperature :- 60 °C

vii. Gradient Program: (Table 1)

Table 1.

Time (in min)	Mobile Phase A%	Mobile Phase B%
0	70	30
10	60	40
20	60	40
30	53	47
35	53	47
45	40	60
50	30	70
55	30	70
55.1	70	30
60	70	30

A 20µL SST solution of Darunavir Standard Spiked with Impurities solution was injected and chromatogram recorded.

Study of System Suitability Parameters

Preparation of System Suitability Solution (SST): An accurately weighed quantity about 120.0mg of Darunavir was transferred to 100.0mL volumetric flask, 50.0mL of diluent was added, sonicated to dissolve, cool and 3 mL portion of DAR-I impurity stock solution added and volume was made up to the mark with diluent and mixed (Solution S1). (Concentration of DAR-I Impurity 2.4µg/mL and Darunavir 1200µg/mL).

Sample Preparation for Study of Linearity (Calibration Curve)

Preparation of DAR-I Impurity Stock Solution: An accurately weighed quantity about 3.0mg of DAR-I impurity was transferred to 100mL volumetric flask, 70.0mL of diluent was added, sonicated to dissolve and volume was made up to the mark with diluent and mixed. (Concentration 30.0µg/mL Stock solution).

Darunavir Standard Stock Solution: An accurately weighed quantity about 3.0mg of Darunavir standard was transferred to 100mL volumetric flask; add 70.0mL of diluent, sonicated to dissolve volume made up to the mark with diluent and mixed. (Concentration 30 µg/mL Stock solution)

Sample Preparation for Validation of Method:

Preparation of DAR-I Impurity Stock Solution for Accuracy: An accurately weighed quantity about 3.2 mg of DAR-I impurity was

transferred to 50 mL volumetric flask, 30mL of diluents was added, sonicated to dissolve and volume made up to the mark with diluent mixed. (concentration of 60.7 μ g/mL as a stock solution).

NOTE: Impurity is spiked in 2nd dilution of sample solution.

Preparation of Sample for Linearity & Range: The working concentration level of Darunavir is about 160 μ g/mL, was prepared in diluent.

Results and Discussion

Selection of λ_{\max} by UV Spectroscopy

Standard solution of Darunavir 10 μ g/mL was scanned in the range of 400-200nm in 1.0cm cell against solvent (methanol) blank and spectrum were recorded. The spectra of Darunavir standard solution is shown in Figure 2. The λ_{\max} of Darunavir was found to be 265.0nm, hence selected as detection wavelength for further experimentation.

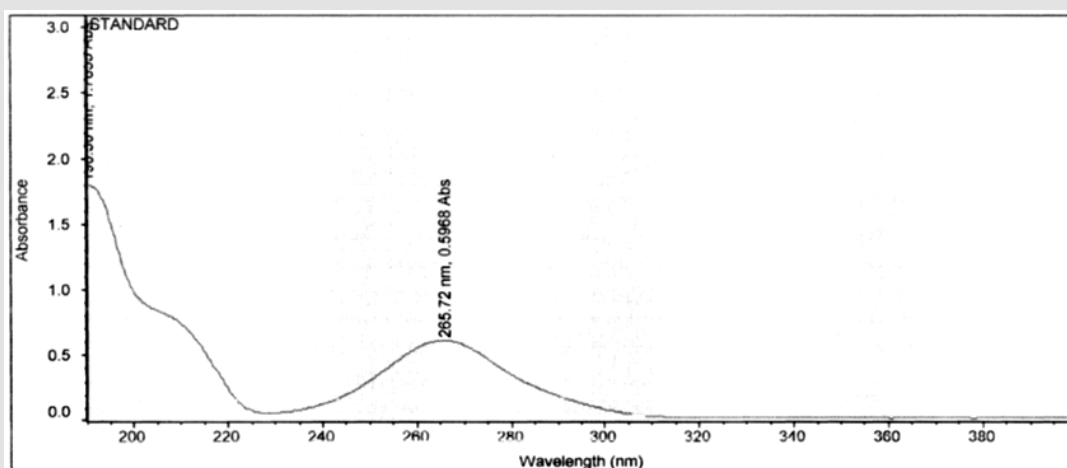


Figure 2: UV spectrum of Darunavir Standard Solution.

Preliminary Selection of Chromatographic Condition:

In order to optimize chromatographic conditions to separate elute and quantify Darunavir one or two parameters (Column and flow rate) were modified at each trials. Standard solution

was prepared in different mobile phases varying in composition and various trials are taken for the selections of chromatographic conditions such as mobile phase, column, flow rate, wavelength, temperature and pH are shown in Table 2. The chromatogram recorded for trail no 3 is shown in Figure 3.

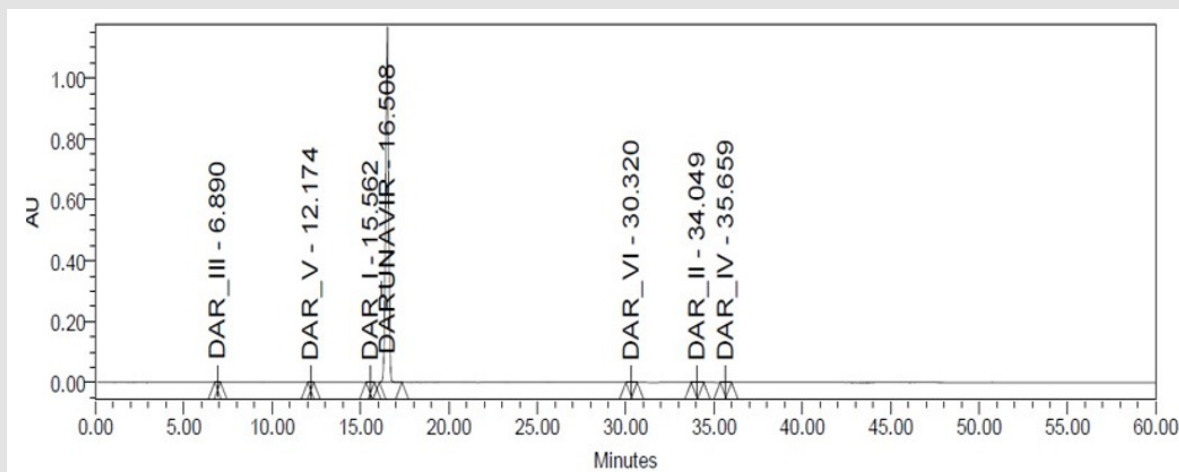


Figure 3: Chromatogram recorded for Darunavir impurity spiked Solution.

Table 2: Trials and Chromatographic observation.

Trial No.	Change in Parameters		Remark
Trial 1	Column	ACE C18, 150 x 4.6mm, 3.0µm	Resolution between impurities and Darunavir was not found to be satisfactory.
	Flow Rate	1.5mL/min	
	Injection Volume	20µL	
	Detector	265nm	
	Column temp	60 °C	
	Sample temp	15 °C	
Trial 2	Column changed to Inertsil C8, 150 x 4.6mm, 3.0µm		Resolution between Dar-I impurity and Darunavir was not optimum
Trial 3	Column changed to Zorbax, C8, 250 x 4.6mm, 5.0µm with flow rate 1.0mL		All the impurity peaks were well resolved with proper peak shape except Dar-I and eluted out within 40 min

Conclusion

From the above chromatographic condition the finalized RP-HPLC parameters are shown in Table 3.

Table 3: Final Optimized chromatographic condition.

Parameters	Condition
Stationary Phase	Zorbax SB-C8, 250 x 4.6mm, 5µm.
Mobile Phase	Gradient program as above
Flow Rate	1.0mL/min
Detection Wavelength	265nm
Pump Mode	Gradient
Injection Volume	20µL
Column Temperature	58-60 °C
Sample Temperature	15 °C

Study of System Suitability Parameters:

The study of chromatogram Figure 3 reveals that the API and all the impurities were well resolved except DAR-I Impurity which elutes very close to Darunavir API peak, hence resolution of the same was studied. System suitability parameters were measured to

verify the system performance. For this sample solution prepared which contains DAR-I Impurity was 2.4µg/mL and Darunavir 160µg/mL of concentration and five replicate injections of 20µL injected. The chromatogram recorded is shown in Figure 4 and Observations are recorded in Table 4.

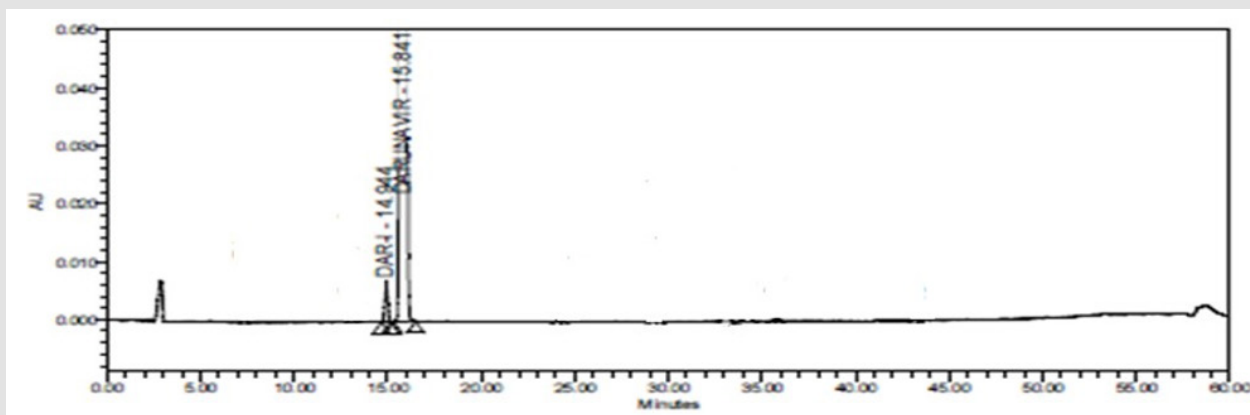
**Figure 4:** Chromatogram of Darunavir System suitability Solution.

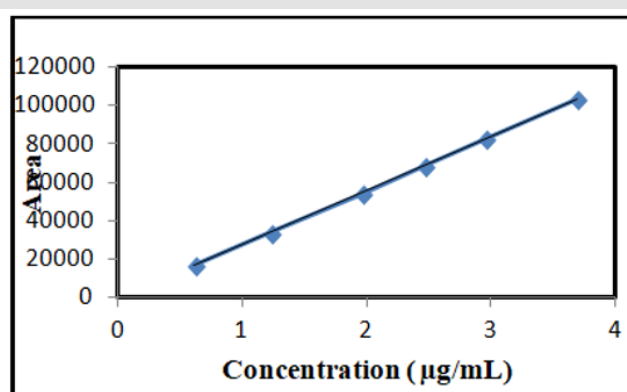
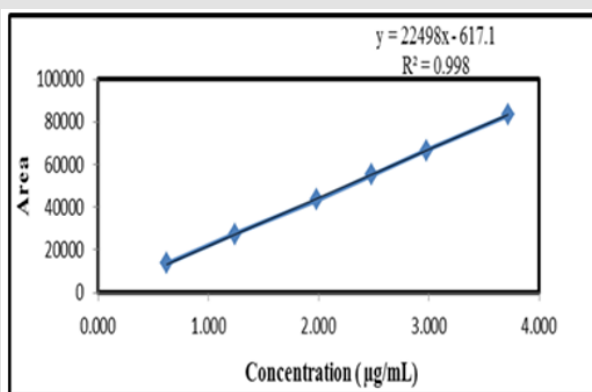
Table 4: Results of System Suitability Parameters.

Sr. No.	Wt. of std. Taken(mg)	Darunavir	Imp Dar-I
		AUC (mAU)	
1	301.2	51415	15914
2		51692	15945
3		51947	15987
4		51864	15908
5		51855	15928
	Mean	51591	15936
	%RSD	0.9	0.19
	Theoretical Plates	37274	44025
	Asymmetry	1.1	0.95
	Resolution	3.08	---

Study of Linearity (Calibration Curve)

Preparation of Linearity Stock Solution: The dilutions were prepared for Linearity Level (%) from 50% to 150% in such a way that we get final solutions having concentration in the range

of 1 to 4 µg/mL and observations recorded & graph plotted as concentration of drug Vs peak area are shown in Figures 5a&5b for drug and impurity 1. The correlation coefficient for Darunavir and DAR-I Impurity was found to be 0.998 and 0.999 respectively.

**Figure 5:** Plot of Calibration curve of

- Darunavir
- Impurity 1.

Application of Proposed Method to Formulation

The optimized gradient program and chromatographic parameters were applied to check the presence of unknown peaks in diluents and standard solution of Darunavir. For this, 20 µL injection of diluent (blank) and standard solution prepared earlier

were injected in system. The chromatogram was shown in Figures 6(a)&(b). The observations from Figures 6a&6b are recorded in Table 5. From the figure and observation it was concluded that no additional peaks were seen in the chromatogram of diluents and standard solution. Hence the proposed method was further applied to formulations.

Table 5: Peak name, Retention time, Area and Asymmetry.

Name	Retention Time	Area (mAU)	Asymmetry	Theoretical Plate
Darunavir	16.047	20795	0.92	48250

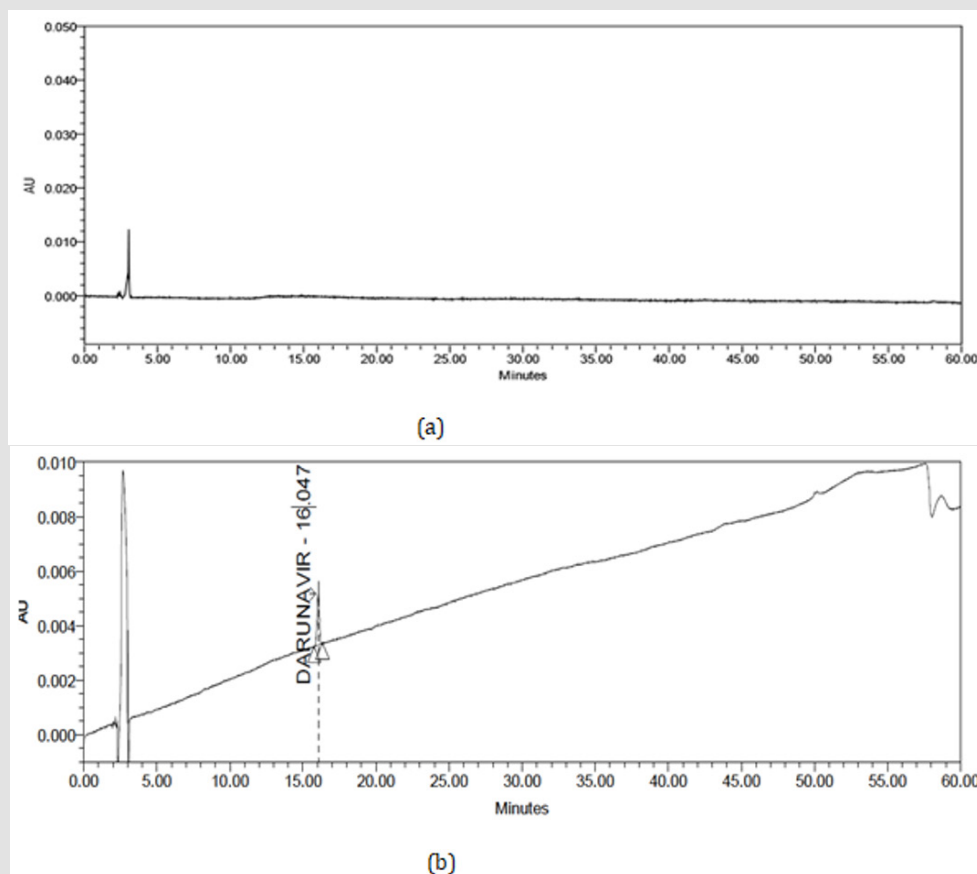


Figure 6:

- (a) Chromatograms of Diluent (blank) solution
- (b) Chromatograms of Darunavir Standard solution.

Validation

Specificity: (Identification, Interference & Peak Purity)

A 20 μ L volume of Blank (Diluent), Darunavir Standard solution, impurity mixture solution and sample solution spiked with known impurities at specification limit were injected into the HPLC system. There was no interference from the blank at the retention time of

Darunavir and impurity peaks. The chromatogram of impurity spiked sample reveals that the impurities were well separated and also the peak purity data reveals that there was no co-eluting peaks and no interference of impurities at the retention time of analyte peak. The observed chromatograms are shown in Figures 7a-7k. The observations of spiked sample for specificity are shown in the Table 6.

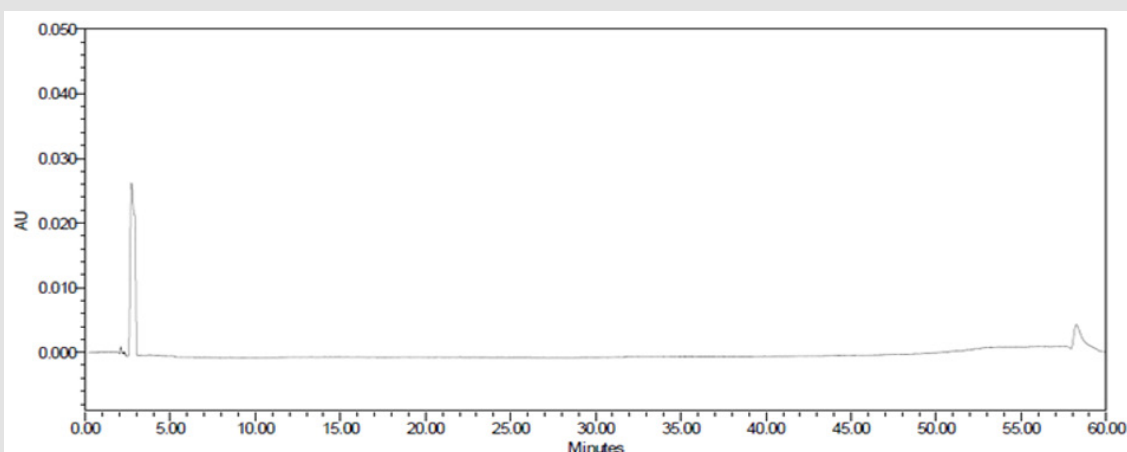


Figure 7(a): Chromatogram of Blank (Diluent) solution.

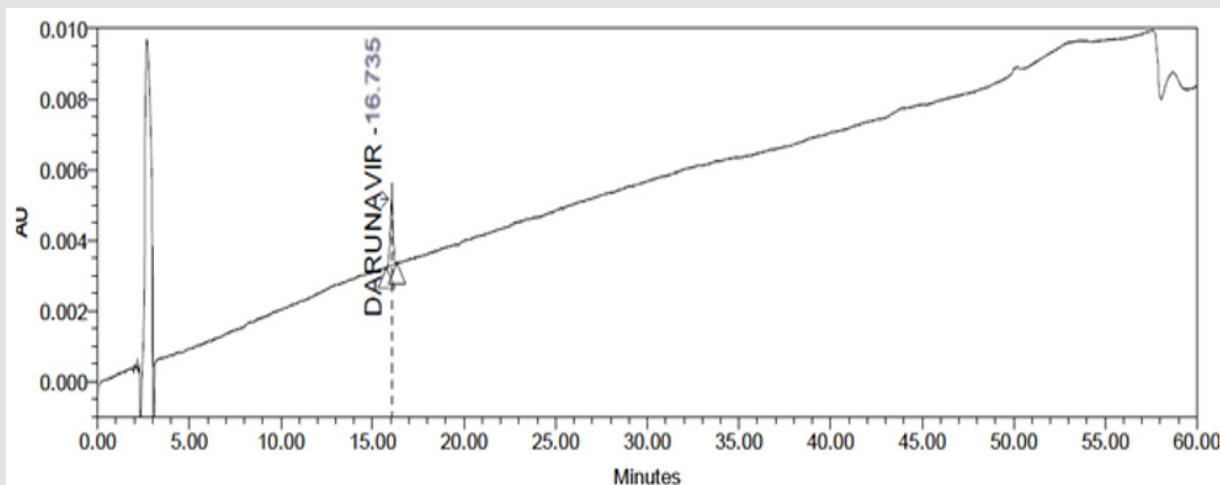


Figure 7(b): Chromatograms of Darunavir Standard Solution.

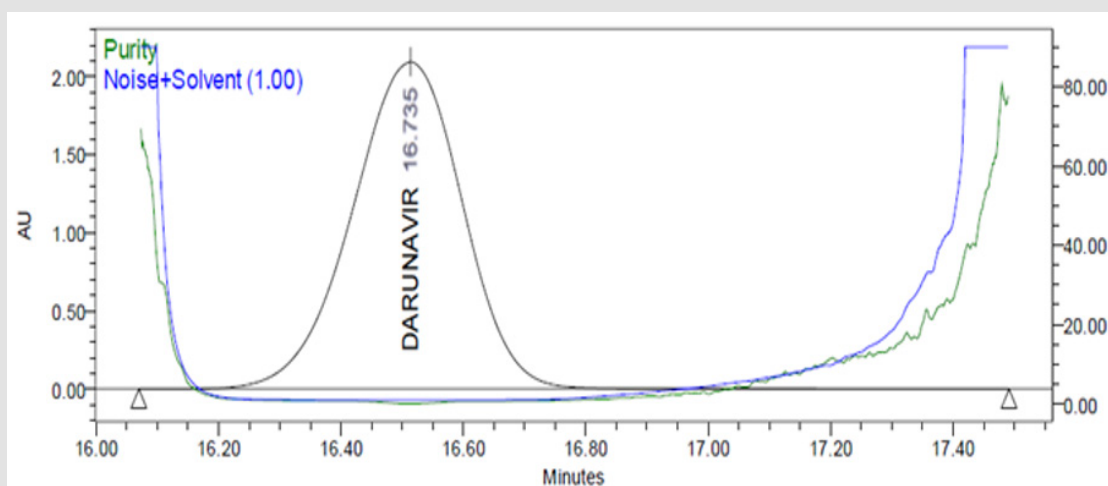


Figure 7(c): Peak Purity Plot of Darunavir Standard Solution.

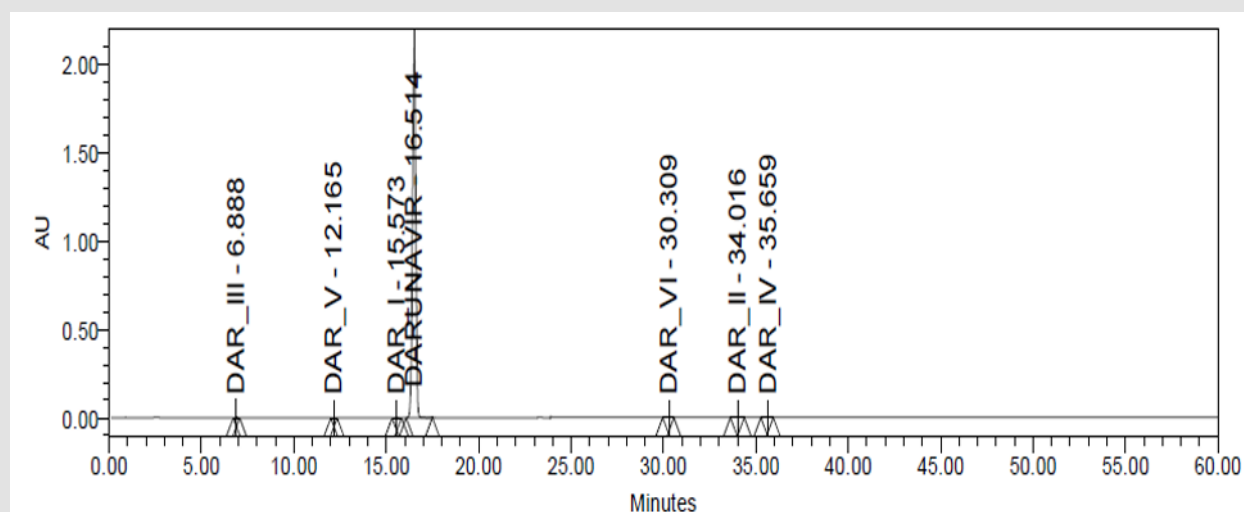


Figure 7(d): Chromatograms of Darunavir Spiked Solution.

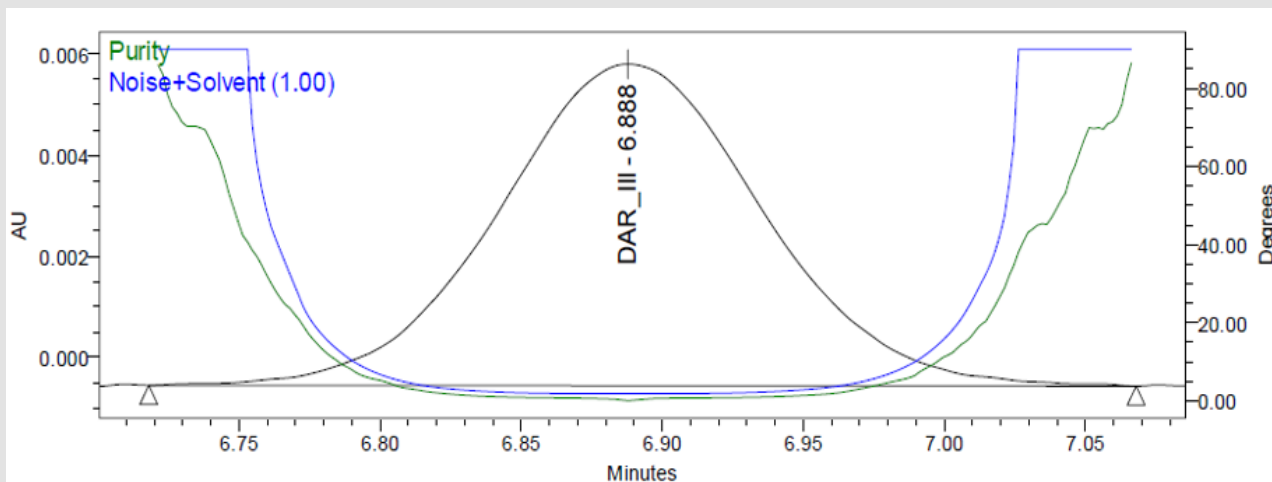


Figure 7(e): Peak Purity Plot of Impurity Dar-III Solution.

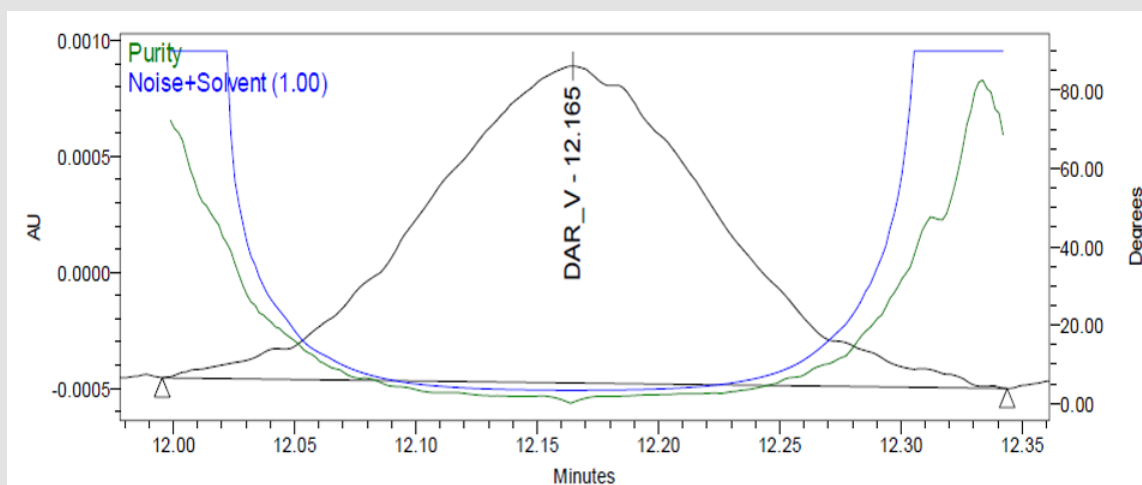


Figure 7(f): Peak Purity Plot of Impurity Dar-V Solution.

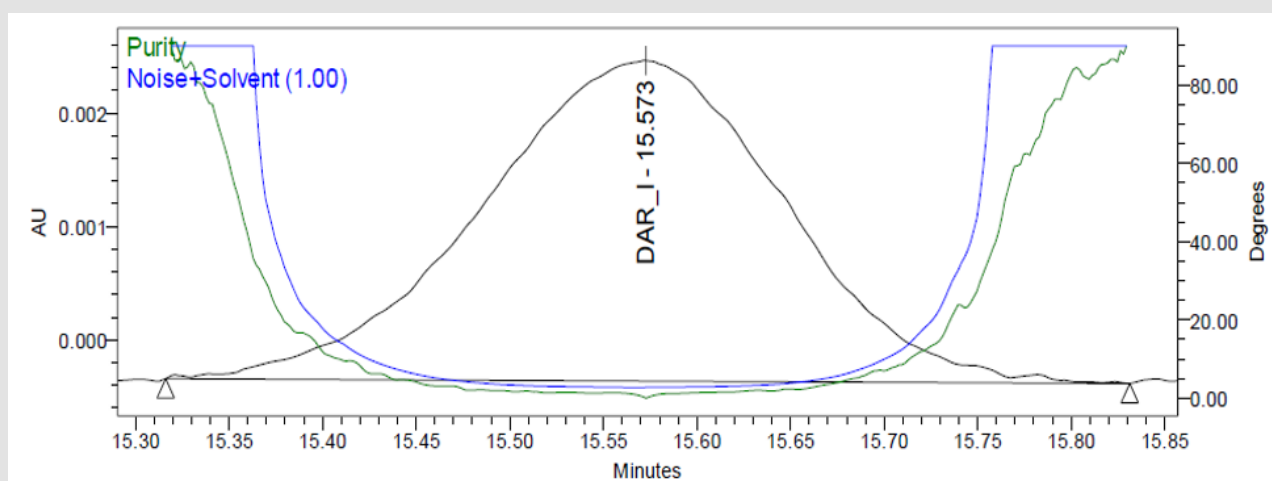


Figure 7(g): Peak Purity Plot of Impurity Dar-I Solution.

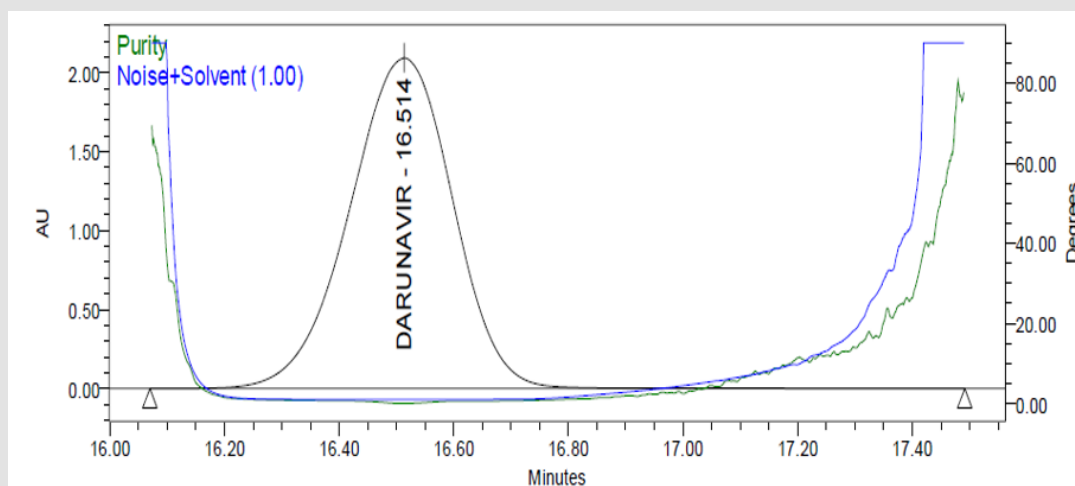


Figure 7(h): Peak Purity Plot of Darunavir Solution.

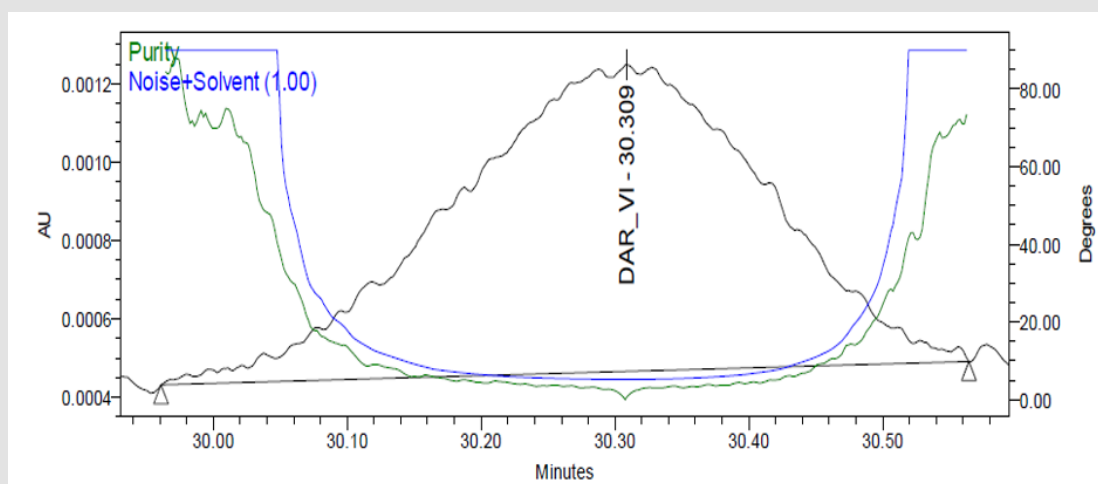


Figure 7(i): Peak Purity Plot of Impurity Dar-VI Solution.

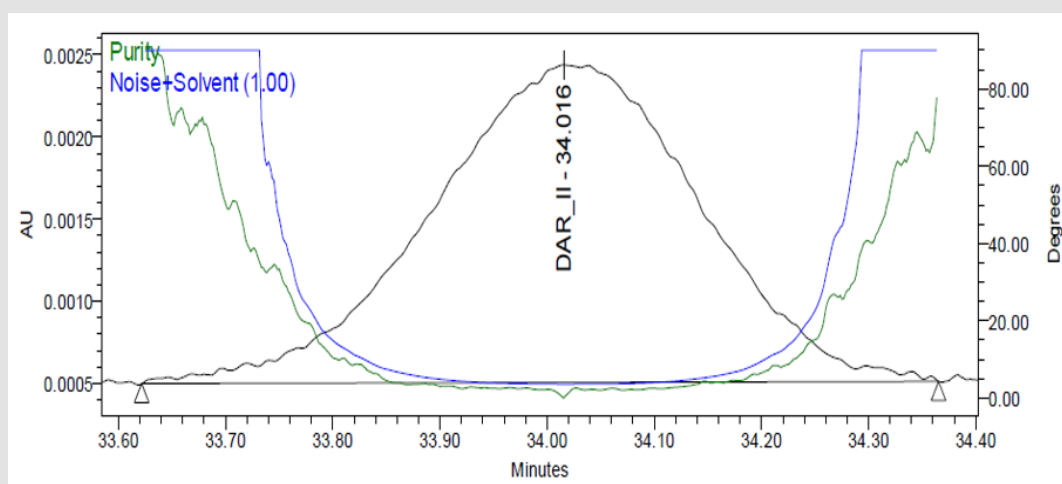


Figure 7(j): Peak Purity Plot of Impurity Dar-II Solution.

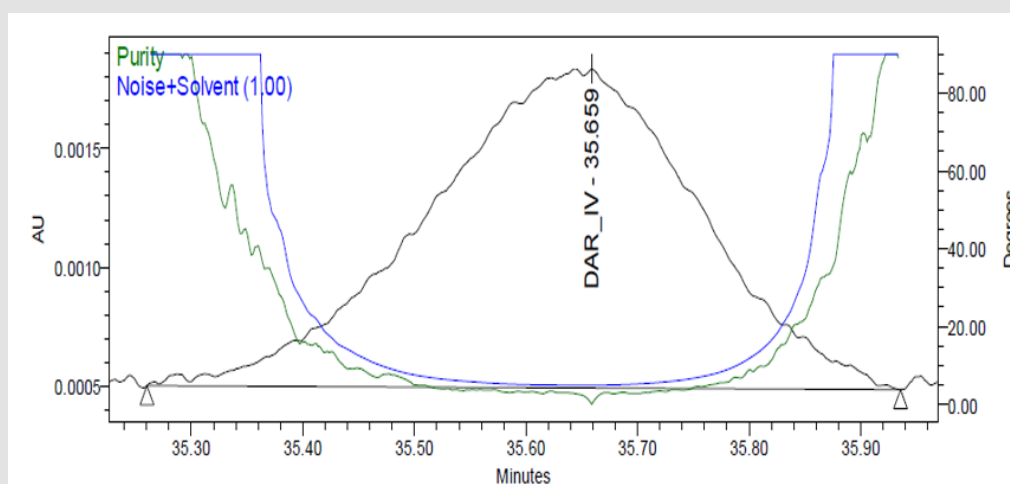


Figure 7(k): Peak Purity Plot of Impurity Dar-IV Solution.

Table 7: Observation of Specificity.(Identification, Interference and Peak purity).

Component	Retention time	RRT	Purity angle	Purity threshold
Blank	-	-	-	-
Individual ID				
DAR-I	14.688	NA	1.592	9.037
DAR-II	32.408	NA	2.589	15.331
DAR-III	7.263	NA	0.970	6.082
DAR-IV	33.385	NA	2.077	9.393
DAR-V	12.714	NA	1.817	9.656
DAR-VI	28.657	NA	4.025	16.122
Darunavir Standard Solution				
Darunavir Standard	16.006	NA	1.413	2.466
Darunavir - Spiked Sample				
DAR-III	6.888	0.41	1.212	2.296
DAR-V	12.165	0.73	3.209	4.438
DAR-II	15.573	0.94	2.492	3.752
Darunavir	16.514	1.00	0.429	1.006
DAR-VI	30.309	1.83	5.306	6.714
DAR-II	34.016	2.05	3.677	4.884
DAR-IV	35.659	2.15	5.068	6.584

Accuracy

Accuracy (Recovery) for DAR-I Impurity: Accuracy was performed by injecting three replicates each of sample spiked at four levels LOQ, 50%, 100% and 150% of the corresponding limit concentration of known impurities like DAR-I Impurity. The limit concentration for DAR-I Impurity (0.20%) each level prepared in triplicates. Prepared 12 sample spiked with known impurities like DAR-I Impurity at 4 different levels, each level in triplicate from 3x LOQ, 3x 50%, 3x 100% and 3x 150% of the limit concentration.

DAR-I Impurity stock solution was prepared of a concentration 60.7µg/mL as a stock solution.

NOTE: Impurity is spiked in 2nd dilution of sample solution.

Procedure

Weighed 20 tablets and average weight of tablet calculated, crushed into fine powder and mixed well. Weighed and transferred powder equivalent to 800mg into 100mL volumetric flask, 70mL of diluent was added, sonicated for 30 minutes with intermittent shaking and allowed it to cool, volume made up to the mark with diluent and mixed. Further 15mL of this solution was diluted to 100mL volumetric flask, added DAR-I impurity stock solution mentioned in above table and volume made up to the mark with diluent and mixed. Filter the sample solution through 0.45µ nylon membrane filter.

Precision

The precision of an analytical procedure is usually expressed as the variance, standard deviation, relative standard deviation or coefficient of series of measurements (Table 7).

Table 7: Accuracy for DAR-I impurity.

Level %	Amount Added (µg/mL)	Response Area (mAU)	Amount recovered (µg/mL)	% Recovery	Mean recovery %
LOQ	05.11	16667	05.20	101.8	100.2
		16171	05.05	98.8	
		16381	05.11	100.0	
50	10.22	32585	10.17	99.5	99.4
		32703	10.21	99.9	
		32378	10.11	98.9	
100	20.43	65614	20.48	100.2	100.4
		65892	20.57	100.7	
		65656	20.50	100.3	
200	30.65	97715	30.51	99.5	99.8
		98290	30.69	100.1	
		98076	30.62	99.9	

System Precision for Related Substance: System precision was performed by single injection of Blank (Diluent) and six replicate injections of DAR standard solution were injected on the system. The results obtained from the observation of chromatogram was shown in Table 8.

Table 8: Accuracy for DAR-I impurity.

Injection	For Related Substance
	Area (mAU)
1	51415
2	51692
3	51947
4	51864
5	51855
6	50774
Mean	51591
SD ±	442.635
%RSD	0.9

Method Precision

Method Precision was performed by injecting six samples solution prepared by spiking DAR-I impurity (0.20%) & Darunavir

for unknown impurities at limit concentration (0.20%) to the Darunavir tablets were prepared and injected on the HPLC. The observations are shown in Table 9.

Table 9: Method precision for Known Impurities & % Unknown impurities.

v Sample. No.	% Known Impurities		% Unknown Impurities 0.2% Spiked
	DAR-I	Total Impurities	Darunavir
Unspiked sample-1	ND	0.20	NA
Unspiked sample-2	ND	0.21	
Unspiked sample-3	ND	0.19	
Sample-1	0.20	0.40	0.20
Sample-2	0.20	0.40	0.20
Sample-3	0.20	0.40	0.20
Sample-4	0.20	0.40	0.20
Sample-5	0.20	0.40	0.20
Sample-6	0.20	0.40	0.20
Mean	0.20	0.40	0.20
% RSD	0.0	0.0	0.0

Intermediate Precision (Ruggedness) For Related Substance: The observation and results is shown in Table 10.

Table 10: Intermediate precision for known and unknown impurities.

Sample. No.	% Known Impurities		% Unknown impurities 0.2% spiked
	DAR-I	Total impurities	Darunavir
Unspiked sample-1	ND	0.15	NA
Unspiked sample-2	ND	0.16	
Unspiked sample-3	ND	0.17	
Sample-1	0.19	0.35	0.20
Sample-2	0.19	0.34	0.20
Sample-3	0.19	0.34	0.19
Sample-4	0.19	0.34	0.20
Sample-5	0.19	0.35	0.19
Sample-6	0.19	0.35	0.20
Mean	0.19	0.35	0.20
% RSD	0.0	1.6	2.6

Linearity & Range

It was performed in the range of LOQ to 150% of the Darunavir Sample. The working conc. level of Darunavir is about 30µg/mL. The range is proposed to be from LOQ to 150 % of 30µg/mL. Sample preparations are prepared as per earlier procedure in Development of HPLC Method for Estimation of Darunavir Related Substance in Formulation for study of linearity (calibration Curve). Then each solution was injected and chromatographed. The observation of linearity of drug is recorded in Table 11 and graph was plotted as

Concentration (µg/mL) w.r.t limit concentration v/s area shown in Figure 8a-8b. The correlation coefficient for Darunavir and Dar-I impurity was found to be 0.998 and 0.999 respectively.

Limit of Detection and Limit of Quantitation

Based on determination of prediction linearity, six replicate injections were made for LOQ. Six replicates of above predicted LOD and LOQ solution were injected on system and calculated % RSD are summarized in Table 11.

Table 11: Precision for LOD and LOQ for Darunavir & DAR-Impurity.

Sr. No.	LOD		LOQ	
	Response Area (mAU)		Response Area (mAU)	
	Darunavir	DAR-I impurity	Darunavir	DAR-I impurity
Conc. (µg/mL)	0.039	0.058	0.617	0.613
Conc. of sample (%)	0.0033	0.0048	0.051	0.051
1	814	1754	13660	17110
2	1045	1557	13954	17266
3	1006	1480	13921	17529
4	842	1953	14651	18394
5	874	1541	13912	17477
6	1079	1477	14121	17516
Mean	943	1627	14037	17549
% RSD	12.0	11.6	2.4	2.5

Forced Degradation

In forced degradation study a drug substance or the drug product is exposed to an environmental vigorous than the normal i.e. at high temperature, high humidity over the period of time, hydrolysis of drug with base or acid and photo stability of drug are called as accelerated stability conditions. Force degradation studies are performed to get an idea of how drug substance or product

degrades, degenerate and behaves under changing condition, which helps in developing the stability indicating method of analysis. The ICH Guideline Q1A suggests the following condition to the employed.

Solution State Analysis:

- Acid Degradation
- Base Degradation

c. Peroxide Degradation

Solid State Analysis

- Photolytic Degradation (UV Light)
- Humidity Studies
- Thermal Degradation

Solution State Stability

For Related substance: For the solution stability study for the related substances, sample prepared to get final concentration of 1200 µg/mL Concentration solution of Darunavir. The amount of standard drug undegraded was calculated using the following formula:

The amount of standard drug undergarded was calculated using the formula

$$\% \text{ Drug undegradation} = \frac{Au(\text{expose}) \times W(\text{unexposed})}{As(\text{unexposed} \times Wstd(\text{exposed}))} \times \text{Potency} \quad (1)$$

Where,

Au = Peak area of standard (exposed), As = Peak area of standard (unexposed), Wstd = Weight of Standard

Calculation formula for Mass balance:

$$\text{Mass balance} = \frac{(\text{Assay of degraded sample} + \text{total impurities generated})}{(\text{Assay of control sample} + \text{total impurities present})} \times 100 \quad (2)$$

Acid Hydrolysis

It was performed by adding 15mL of 1N HCL & kept on bench top at room temperature for 20 Min. and neutralized 1N alkali. After neutralization, the standard solutions were prepared following general procedure as described earlier. The Sample solution of assay and sample solution for related substance solutions were injected after 20min and chromatographed separately using optimize chromatographic conditions.

Base degradation: It was performed by adding 15mL of 1 N NaOH & kept on bench top at room temperature for 14 h and neutralized 1N alkali. After neutralization, the sample solution was prepared following general procedure as described earlier. The Sample solution of assay and sample solution for related substance solution were injected after after 14h in 1 N NaOH and chromatographed separately using optimize chromatographic conditions.

Peroxide degradation: It was performed by adding 15mL of 30

% hydrogen peroxide solution & heated at 70 °C in water bath for 5 h, allowed to cool and volume made up to the mark with diluent and mixed. The Sample solution of assay and sample solution for related substance solution were prepared following general procedure as described earlier. The sample solution was injected and chromatographed separately using optimize chromatographic conditions. From the chromatographic observations of Acid degradation, Alkali degradation and peroxide degradation chromatograms, AUC were noted, % undegraded drug (Assay) and mass balance was calculated using the formula 1 and 2 is shown in Table 12.

Solid State Analysis

Photolytic degradation (UV Light): According to ICH Guideline sample should be exposed to light to providing an overall illumination of not less than 200 watt hrs. /square meter. The stability chamber was calibrated using appropriate UV meter for UV Light study and the study was carried out for 7 days.

Exposure period of UV light was calculated by the following formula:

$$200 \times 1000000 / \text{Intensity} / 10000 \text{ hrs}$$

Darunavir sample solution was prepared by general procedure as described earlier and it was performed by exposing the sample solutions under UV and white light for 1.2 million lux hours and an integrated near ultraviolet energy of not less than 2000 watt/square and volume made up to the mark with diluents and mixed. The sample solution was prepared following general procedure. After photolytic degradation sample solution for assay and for related substance solution were injected and chromatographed separately using optimize chromatographic conditions.

Thermal degradation

It was performed by heating the solution in the oven at 50 °C for 24 h, allowed to cool and volume made up to the mark with diluent and mixed. The sample solution was prepared following general procedure. The sample solution for assay and for related substances were injected and chromatographed separately using optimize chromatographic conditions. From the chromatographic observations of Photolytic degradation (UV Light) and thermal degradation chromatograms, AUC were noted, % undegraded drug (Assay) and mass balance was calculated using the formula 1 and 2 is shown in Table 12. Purity Threshold was found to be greater than purity angle.

Table 12: Observation of Forced degradation.

Condition	Impurity Name	RRT	% Content	Purity Angle	Purity Threshold	Peak Purity	% Assay	% Mass Balance
Control	DAR-I	NA	ND	NA	NA	NA	101.1	NA
	Max. Unknown	2.63	0.06	4.319	5.768	Passes		
	Total Impurities	NA	0.2	NA	NA	NA		

Acid_1N_ HCL_20 Min_RT	DAR-I	NA	ND	NA	NA	NA	86.9	97.5
	Max. Unknown	0.88	8.5	0.346	0.420	Passes		
	Total Impurities	NA	11.9	NA	NA	NA		
Base_1N_ NaOH_14 HRS_RT	DAR-I	0.56	0.08	0.260	0.461	Passes	88.9	95.5
	Max. Unknown	0.88	1.5	1.214	1.510	Passes		
	Total Impurities	NA	7.8	NA	NA	NA		
Peroxide_ 30%.5 HRS_70 °C	DAR-I	NA	ND	NA	NA	NA	102.5	101.4
	Max. Unknown	2.63	0.05	8.111	16.559	Passes		
	Total Impurities	NA	0.2	NA	NA	NA		
Photolytic study	DAR-I	NA	ND	NA	NA	NA	102.6	NA
	Max. Unknown	2.63	0.04	6.754	16.939	Passes		
	Total Impurities	NA	0.1	NA	NA	NA		
Heat 24 HRS_50 °C	DAR-I	NA	ND	NA	NA	NA	103.2	102.1
	Max. Unknown	2.63	0.06	4.845	6.197	Passes		
	Total Impurities	NA	0.2	NA	NA	NA		

Note: ND = Not Detected, NA= Not Applicable

Stability in Analytical solution

The standard and sample solution was prepared as per described in general procedure for preparation of solutions and

kept sample for 24 h at sample temperature condition (15 °C) & room temperature (25 °C) after 24 hrs sample was injected time to time continuously to check the solution stability. The results obtained are shown in Table 13.

Table 13: Stability in analytical Standard and Sample solution (15 °C) and (25 °C).

Time (in h)	Standard and Sample solution (15 °C)				Standard and Sample solution (25 °C)			
	Standard Solution Stability		Sample Solution Stability		Standard Solution Stability		Sample Solution Stability	
	Response (area) (μV)	Cumulative % RSD	% Assay	Absolute % Difference	Response (area) (μV)	Cumulative % RSD	% Assay	Absolute % Difference
Initial	997789.2	NA	98.5	NA	992334	NA	98.5	NA
12	1006991	1.1	99.4	-0.90	993466	1.0	99.4	-0.90
20	968751	1.5	98.1	0.40	987251	1.0	99.0	-0.50
24	995670	1.4	100.2	-1.70	1000622	1.0	97.6	0.90

Filter Compatibility

The Standard and Sample solution was prepared as per described in general procedure for preparation of solutions and

filtered through different types of filters (PVDF, Nylon, PTFE & Teflon + Glass) and injected on to the HPLC. The centrifuged sample and filtrates of different filters were analyzed for their equivalency.

The observations are shown in Table 14.

Table 14: Study of Filter compatibility.

Condition	Standard		% Impurity			
Time	% Recovery	Absolute difference	DAR-I	Absolute difference	Total impurities	Absolute difference
Centrifuged	99.4	NA	0.20	NA	0.40	NA
0.45μ Nylon Filter	98.9	0.49	0.20	0.00	0.40	0.00
0.45μ PVDF Filter	99.3	0.06	0.20	0.00	0.40	0.00
0.45μ PTFE Filter	99.1	0.30	0.20	0.00	0.40	0.00
0.45μ PTFE + Glass Filter	99.1	0.23	0.20	0.00	0.39	0.01

Robustness

The robustness of an analytical procedure is measured by making small but deliberate variations in method parameters and observing the effect of these variations on the system suitability parameters by injecting standard and sample solution.

- Change in chromatographic conditions:
- Change in wavelength (± 5 nm)
- Change in flow rate (± 0.1 ml/min)
- Change in column temperature (± 5 °C)

e. Change in buffer pH (± 0.2 unit).

The observations are shown in Table 15a & 15b.

Table 15(a): Robustness for Darunavir Standard solution.

Changes in Parameters	Values	Retention Time of Darunavir	Theoretical Plates	Symmetry Factor	%RSD	Resolution Between Darunavir and DAR-I Impurity
*Control	As per method	16.865	37274	1.1	0.9	2.9
Wavelength (265nm)	260nm	16.864	37717	1.1	0.8	2.9
	270nm	16.864	37133	1.1	1.3	2.9
Flow Rate (1.0mL/Min)	0.9mL/min	18.080	34227	1.1	0.6	2.8
	1.1mL/min	15.770	33424	0.9	1.2	2.7
Column temp. (60 °C)	55°C	17.543	316466	0.9	0.4	2.2
	65°C	16.169	32455	1.0	0.6	3.2
pH (4.0)	3.8 pH	17.114	40260	1.0	1.5	3.2
	4.2 pH	17.107	39725	1.0	0.9	3.1

Table 15(b): Robustness for Darunavir Spiked Sample.

Changes in Parameters	Values	DAR-I	Absolute Difference	Total Impurities	Absolute Difference	Resolution Between Darunavir and DAR-I Impurity
*Control	As per method	0.20	NA	0.40	NA	2.9
Wavelength (265nm)	260nm	0.20	0.00	0.39	0.01	2.9
	270nm	0.20	0.00	0.41	-0.01	2.9
Flow Rate (1.0mL/Min)	0.9mL/min	0.20	0.00	0.39	0.01	2.8
	1.1mL/min	0.20	0.00	0.40	-0.01	2.7
Column temp. (60 °C)	55°C	0.19	0.01	0.39	0.01	2.2
	65°C	0.20	0.00	0.39	0.01	3.2
pH (4.0)	3.8 Ph	0.19	-0.01	0.36	-0.01	3.2
	4.2 Ph	0.19	-0.01	0.36	-0.01	3.1

Conclusion

From all above studies, we can concluded that the proposed HPLC method can be used successfully for estimation of Assay and Related Substance in Darunavir. It can also be useful in detecting the degradation of unknown impurities in tablet formulation. Further this method may be applied to preparative HPLC for qualification of unknown impurities which might be generated during forced degradation studies. The results obtained by HPLC method for determination of Darunavir are reliable, accurate and precise. The method can be employed for routine quality control analysis of Darunavir in tablet dosage form.

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