

In-Vitro Antifungal Activity of Citrullus Colocynthis Schard Against Aflatoxigenic Aspergillus Flavus and Ochratoxigenic Aspergillus Ochraceus

Nora Nahal Bouderra¹, Khalid Mohamed Khalid Elhussain^{2,3,4*} and Hamid Kadi¹

¹Department of Biology, Faculty of Science of Nature and Life, Algeria

²Department of Hematology, Faculty of Medical Laboratory Sciences, Omdurman Islamic university, Sudan

³Faculty of Medicine, Omdurman Ahlia University, Sudan

⁴International University of Africa, Faculty of Medical laboratory sciences, Sudan

*Corresponding author: Khalid M Khalid Elhussain, Department of Hematology, Faculty of Medical Laboratory Sciences, Omdurman Islamic University, Sudan



ARTICLE INFO

Received: 📅 February 21, 2020

Published: 📅 March 03, 2020

Citation: Nora N B, Khalid Mohamed K E, Hamid K. In-Vitro Antifungal Activity of Citrullus Colocynthis Schard Against Aflatoxigenic Aspergillus Flavus and Ochratoxigenic Aspergillus Ochraceus. Biomed J Sci & Tech Res 26(2)-2020. BJSTR. MS.ID.004316.

ABSTRACT

Background: Resistance to current antibacterial and antifungal drugs are growing global concerns. Traditional medicine is a potential source of new antibacterial and antifungal.

Objective: Aim is to study the effects of aqueous and methanolic of fruits leaves and roots of Citrullus colocynthis Schard on fungal strain. Two methods have been used with this extracts on 2 fungal strains producer of mycotoxins; determination of growth radial technique on solid medium and determination of biomass technique on liquid medium. Aqueous and methanolic extract of fruit, leaves and root of Citrullus colocynthis Schard; these are examined for their antifungal potentiality against Aspergillus flavus and Aspergillus ochraceus.

Result: The maximum percentage growth inhibition 100% obtained from the aqueous extract from fruits of Citrullus colocynthis Shard against Aspergillus Ochraceus. These extracts can be included in the list of herbal medicines due to their high antimicrobial potential.

Keywords: Citrullus colocynthis Schard; Aspergillus flavus; Aspergillus ochraceus; Inhibition; Antifungal

Introduction

Citrullus Colocynthis Schard is a member of the gourd family cucurbitaceae, originally from tropical Asia and Africa, it is now widely distributed in Saharan-Arabian phytogeographic region in Africa and the Mediterranean region [1]. It is small scaped perennial creeping herb with prostate or climbing stem, bearing smooth spherical fruits which are mottled green when young and some white yellow when ripe [2]. In moderate doses a drastic hydragogue, cathartic and diuretic; in

large doses emetic and gastro-intestinal irritant; in small dose it is expectorant and alterative. Physician use this drug extensively as a drastic purgative in ascites and jaundice in various uterine conditions, especially in amenorrhea. Colocynth in the form of the solid extract enters many of the purgative pills of modern pharmacy. It is use full in biliousness, fever, intestinal parasites, constipation, hepatic and abdominal, visceral and cerebral congestions, dropsy, etc. Juice of the fruit

mixed with sugar is a house-hold remedy in dropsy [3]. Citrullus colocynths has very high medicinal value; the plant contains three antitumor ingredients: cucurbitacin B, Cucurbitacin E and the D-glucoside of Beta- sitosterol [4]. The purpose of the present study is to investigate the antifungal activity of fruit (Bark and pulp), roots and leaves extract of Citrullus colocynth is against fungal strains were isolated from grains of stored cereal for human consumption.

Materials and Methods

Plant Material and Sample Collection

The plant used for the present study was collected in September 2011, from Oued Béchar, Béchar is a city in west Sahara, Department, Algeria. The leaves and roots were dried for 20 days in the dark at ambient laboratory temperature (20 to 28°C); the fruits were dried for three months at the same conditions, the grains were be debarrassed debarred, the different part were milled to a fine powder in an electrical mill, and stored in the dark room temperature in closed containers until required.

Qualitative Phytochemical Screening

Each organ of plant (leaves, roots and fruits) was screened for the presence of key families of phytochemicals (Sakar and Tanker, 1991; Trease and Evan, 1984). using the following reagents and chemicals: alkaloids with Mayer's reagents, Flavonoids with metallic magnesium and hydrochloridric acid, Spinosad's for their ability to produce suds, steroids acetic anhydride and concentrated sulphuric acid, Tanin with ferric chloride.

Extraction Protocol

Aqueous Extract: 5g of different organs powder of plant were added to 50 ml of distilled water; the mixture was allowed to reflux for 30min. After cooling it was filtered and stored to 4°C prior to analysis [5,6].

Hydroethanolic Extract: 5g of different organs powder of plant were added to 50 ml of methanol: distilled water (1V/2V), the mixture was allowed to reflux for 30min. After cooling it was filtered. The filtrate was passed in a rotary evaporator at 65°C to vapor the methanol; the crude extract was stored to 4°C prior to analysis.

Fungal Strains

The fungal strains, A. Flavus and A. ochraceous; were obtained from durum wheat, they are isolate purify and identify in biology laboratory at Bechara university. Confirmation of Aspergillus genera was realized by micro culture method described by [7,8]. Furthermore, identification of A. flavus and A. ochraceous species was carried out by single spore method using three cultures media:

Malt Extract Agar(M.E.A) at 25°C; Glycerol Nitrate Agar (G25N) at 25°C and Czapek Yeast Agar(C.Y.A) at 5°C and 37°C. Using the identification key of Pitt (1973). Observation has been made after the first and second week. The fungal strains were stored in tubes of PDA acidified at 4°C.

Mycotoxigenic Test

A. flavus and A. ochraceous were sowed on Y.E.S (Yeast Extract Sucrose) medium rich on B vitamin complex. After two weeks of incubation at 30±2°C, the biomass was removed by filtration of Y.E.S medium. The filtrate was added then to 180 ml of chloroform and stirred for 30min. After decantation, the organic layers were concentrated to 2ml. Aflatoxin and ochratoxin were determined in each extract by spotting samples onto thin layer chromatography plate. Plate was developed with a toluene/Ethyl acetate/ Formic acid (50:40:10, v/v/v) solvent system. Aflatoxin and ochratoxin standard were spotted on the same plate as reference, aflatoxin and ochratoxins spots were identified using 365nm UV [9]. The presence of Aflatoxins was provided by appearance of blue fluorescence for AFB and green fluorescence for OTA which have the same Rf as control.

Antifungal Activity

Evaluation of biomass or radial grow thing was achieved by counting of spores' number using Malassezia Hemet meter in order to obtain the concentration of 105 spores /ml [10].

Growth Radial Technique on Solid Medium: 1,2,3,4 and 5 ml of fruit, root and leaves extract of Citrullus colocynthis were added to 20 ml of PDA a (Potatoes Dextrose Agar acidified) solid medium; after agitation the select solution were transferred into a Petri plats which were incubated for 7 days at 25±2°C after ensemencing the fungal strain in the center. Mycelial radial growth was measured from the third day of incubation [11]. The inhibition percentage of mycelial growth of each extracts was calculated using the following formula:

$$PI = [(Dt-D)/Dt] \times 100 \quad [11]$$

Where Dt is mean diameter of mycelial growth in control and D is mean diameter of mycelial growth in treatment.

Biomass Technique on Liquid Medium: This technique consists to put different volumes of extract in flasks and completed them with 50ml of PDB a (Potatoes Dextrose Broth Acidified) in order to obtain different concentrations. These liquid cultures were sowed with 30µl of sporal suspension. The flasks were incubated for 14 days at 25±2°C [12] filtration, the filter paper was dried at 60°C during 24 hours [13]. Biomass weight formed P was determined using the following formula of Imtiaj and Lee [14].

$$[P=P_1-P_0]$$

Where P0 is the filter paper weight and P1 is the filter paper and fungal biomass weight after dryness.

Results

Qualitative Phytochemical Screening

Phytochemical screening is usually carried out to screen for and to characterize the constituents available in a given plant sample. All phytochemical constituents tested were identified in *Citrullus colocynthis* fruits, leaves and roots as shown in Table 1.

Mycotoxigenic Test: The mycotoxin separation elaborated by the fungal strains was made on a CCM plate, the spots were revealed under the UV lamp at the wavelength 365nm, the elaboration of Aflatoxin B1 by *A. flavus* and ochratoxin A by *A. ochraceus* in the test are confirmed with CCM plate illustrate in Figures 1 & 2.

Table 1: Phytochemical Screening of *Citrullus colocynthis* fruits; leaves and roots.

Phytochemical constituents	Fruits	Leaves	Roots
_ Alkaloid	+	+/-	+/-
_ Tannins	+	+/-	+
_ Saponins	+	+/-	+
_ Flavonoids	+	+/-	+
_ Unsaturated sterols and terpens	+	+	+
_ Sterol and steroid	+	+	+

Key: +: present; -: Absent; +/-: low presence

Table 2: the inhibition percent of fungal strains tested with different concentrations of *Citrullus colocynthis* extracts.

	Aqueous/ methanolic	A. Flavus ip (%)	A. Ochraceus ip (%)
Fruits	Aqueous	88.46 (21mg/ml)	100 (3.3mg/ml)
	Methanolic	100 (25.25mg/ml)	100 (33.66 mg/ml)
Leaves	Aqueous	/	77.77 (18mg/ml)
	Methanolic	100 (10.8mg/ml)	100 (21.6mg/ml)
Roots	Aqueous	100 (10.8mg/ml)	75.55 (13.5mg/ml)
	Methanolic	44.23 (8mg/ml)	100 (6.4 mg/ml)

IP: Inhibition percent

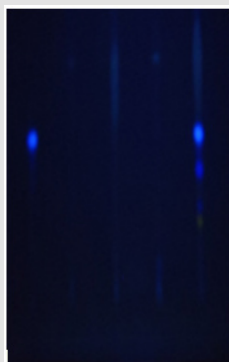


Figure 1: CCM plate of *Aspergillus flavus* strain.

Biomass Technique on Liquid Medium: In fact, the different results collected and recorded in Table 3 show a decrease in fungal biomasses proportional to the concentration of the extracts added

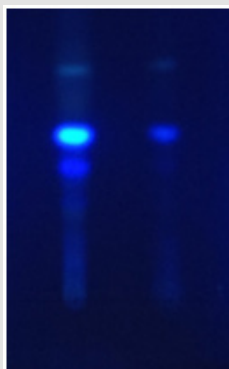
Antifungal Activity

Growth radial technique on solid medium: Different fractions of *Citrullus colocynthis* roots leaves and fruits were found to inhibit the growth of *Aspergillus flavus* and *Aspergillus ochraceus*. Different concentrations of aqueous and hydroethanolic extract for the three part of *Citrullus colocynthis* were tested for their efficacy against fungi of food stored with growth radial technique on solid medium; the result are presented in Figure 3. According to the results obtained it was found that *A. flavus* was 100% inhibited by the methanolic fruit extract and the aqueous root extract, while *A. ochraceus* was 100% inhibited by aqueous extract of fruits and methanolic extracts of leaves and roots. These results are shown on the Table 2.

to the medium with the exception of *A. flavus* with the aqueous extract of the leaves which gave an opposite effect.

Table 3: Fungal biomasses according to the different concentrations of *Citrullus colocynthis* extracts.

	Aqueous Extract			Methanolic Extract		
	Concentrations (mg/ml)	A flavus (mg)	A ochraceus (mg)	Concentrations (mg/ml)	A flavus (mg)	A ochraceus (mg)
Fruits	0	250±08	243±11	0	250±08	243±11
	2.7	191±05	27±07	5.31	195±23	103±14
	5.5	127±10	22±06	11.2	137±11	88±12
	8.1	100±09	21±13	17.8	96±05	54±8
	10.8	96±11	07±11	25.25	89±08	26±11
	13.5	78±15	01±01	33.66	61±14	08±14
Leaves	0	250±08	243±11	0	250±08	243±11
	3.6	100±11	120±04	5.62	89±03	142±16
	7.2	151±06	113±09	12.54	78±07	137±15
	10.8	204±03	78±10	19.77	70±05	130±07
	14.4	253±08	65±10	28.16	58±16	69±08
	18	283±11	65±15	35.66	49±12	63±11
Roots	0	250±08	243±11	0	250±08	243±11
	2.47	83±09	137±05	2.7	98±02	86±03
	5.33	72±16	130±08	6.12	82±05	77±08
	8.05	64±12	125±12	10.47	74±13	59±07
	10.24	59±00	67±06	15.03	56±08	38±05
	12.62	42±09	58±05	18.98	41±07	22±03

Figure 2: CCM plate of *Aspergillus ochraceus* strain.

Discussion

The traditional use of plants as medicines provide the basis for indicating which essential oils and plant extract may be useful for specific medical conditions [15,16]. The present investigation has explored the use of one such plant; *Citrullus colocynthis* Schard endemic in the south west of Algeria, for testing phytochemical compound and the antifungal activity of this endemic plant. Generally, in the phytochemical screening of any plant one normally identifies secondary metabolites that have accumulated to some extent at specific organ of the plant. These metabolites that are mainly used by the plant for protection against herbivores may have pharmacological activity when tested on animals [17]. Result of phytochemical screening of *Citrullus colocynthis* Schard fruits leaves and roots of showed the presence of saponins, sterols, steroid,

terpene, flavonoids, tannin and alkaloids in different proportion in the tree part of plant. This result is in agreement with finding of [18]. which proved that alkaloids were found in all extracts except the roots, flavonoids were present only in seeds; gallic tannin and coumarins only in leaves, and all of them contained steroids. Ambit, et al. [2] confirmed that three phytochemical constituents were identified in *Citrullus colocynthis* Schard seeds extracts as alkaloid, steroid, glycosides and flavonoids. Extraction of secondary metabolites highly depends on using extractor techniques that depend on the chemical properties of these compounds. Water soluble compounds and proteins can be extracted in water or polar solvents whereas water insoluble compounds can be extracted with organic solvents [19]. Several authors Greyer and Harborne [20]; Yin and Cheng [21] have reported the fact that the extract of

certain spices and herbs of medicinal importance exhibit antifungal property. These natural antifungal agents could be exploited in controlling the growth of fungi consequently inhibiting aflatoxin formation. All fungal strains have inhibited by the extracts tested at different concentration 3,3 mg/ml for *A. ochraceus* with fruits aqueous extract and 10.8 mg/ml for *A. flavus* and *P. expansa* with leaves hydroethanolic extract. It's interesting to mention that the hydro methanolic extracts are the most effective antifungal than aqueous extract because methanol allowed to extract well the less polar compound such as terpene drives [22]. Only *A. flavus* who noted a diameter and greater biomass than the control with the leaves aqueous extract. These data may be due to inoculum quality added on culture medium or strain using aqueous extract as carbon source. For *Citrullus colocynthis* Schard fruits; Belsem et al., 2009 reported the antifungal activity.

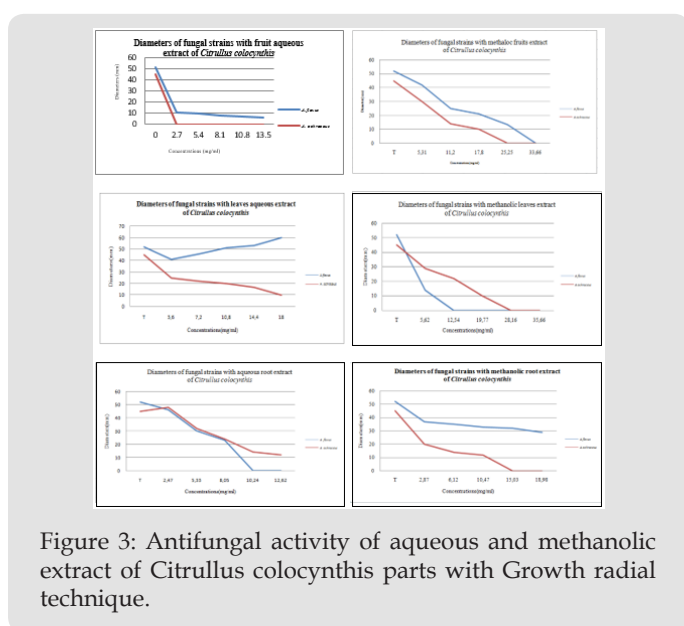


Figure 3: Antifungal activity of aqueous and methanolic extract of *Citrullus colocynthis* parts with Growth radial technique.

These results suggest that the inhibitory effect exhibited by the extract of *Citrullus colocynthis* Schard may be attributable to the secondary metabolites like phenolic compounds and saponins. Activity cannot be imputed to one family of phytochemical only; Alkaloids are commonly found to have antimicrobial properties [23]. Furthermore, tannins have been found in vitro to have various pharmacological properties such as antioxidant, antimicrobial and anti-inflammatory [24] Flavonoids are known to be synthesized by plants in response to microbial infection [25]. Antimicrobial and anticandidal activity may be attributed possibly in combination to various phytochemicals detected during. The extract chemical screening and which are known to cause damage to cell membrane, causing leakage of cellular materials and ultimately the microorganism's death [26-32].

Conclusion

The obtained results might be considered sufficient to prove that *Citrullus colocynthis* Schard extract can be considered

a good antimicrobial agent; it can be used to con an antifungal overcoat against the strain that a major problem of producing mycotoxin, especially in stored foods. The results are only the first step of antimicrobial activity; further studies for the isolation and identification of the active principal and to the evaluation of possible synergism among extract component for their antimicrobial activity. Investigations are in progress to determine the degree of toxicity of these extract.

References

- Feinbrun, Dothan N (1978) Flora Palaestina- Part III Jerusalem: The Israeli Academy of Sciences and Humanities pp. 380.
- Shah CS, Qadry JS (1985) A Textbook of pharmacognosy (5th Edn.). BS Shah, Prakashan, Pankore Naka, Ahmed abed, India, pp. 284.
- Usman M, Abdulhakeem B, Syed Waseemuddin A, Iquba A, Husam B (2003) Antibacterial screening of *Citrulluscolocynthis*. Journal of pharmaceutical science 16(1): 1-6.
- Rajkiran Chauhan, Urvija Garg, Tak RK (2011) Corrosion inhibition of aluminium in acid media by *Citrulluscolocynthis* extract. Journal of chemistry 8(1): 85-90.
- Kadi H, Moussaoui A, Benmahdi H, Lazouni HA, Benyahia A, et al. (2011) Antibacterial activity of ethanolicdaqueous extract of *PunicagranatumL* bark. Journal of applied pharmaceutical science 1(10): 180-182.
- Haris C (1989) Introduction to modern microbiology black wall scientific publication pp. 179.
- Barnett HL (1972) Illustred general of imperfection fungi. Burgess publishing company, (3rd Edn.). Minnesota, USA.
- (1975) Association of Official Analytic Chemists, Natural poison 26, offitial methods of analysis. Association of Official Analytic Chemists Washington, DC, USA, p. 24.
- Serghat SA, Mouria A, Ouazzanitouhami A, Badoc A, Douira A (2004) Effet de quelques fongicides sur le développement in vitro de *pyriculariagrisea* et *helminthosporium oryzae*. Bull Soc Pharm 143: 7-18.
- Singh P, Kumar A, Dubey NK, Gupta R (2009) Essential Oil of *Aeglemarmelos* as a Safe Plant-Based Antimicrobial Against Postharvest Microbial Infestations and Aflatoxin Contamination of Food Commodities. Journal of food science 74 (6): 302-307.
- Tubajika KM (2006) Efficacy of alkyl dimethyl benzyl ammonium chlorid on supression of *phyalospora vaccinii* in laboratory assays. Journal of food protection 69(10): 2460-2464.
- Dhandhukia PC, Thakkar VR (2007) Standardization of growth and fermentation criteria of *Lasiodiplodiatheobromae* for production of jasmonic Acid. African journal of Biotechnology 6(6): 707-712.
- Ambi A, Abdurrahman EM, Sule MI, Pateh UU, Abdurrahman YR, et al. (2007) Phytochemical screening and histopathological studies on the seeds of *colocynthis Cirullus inalbinos* rats. Nigerian journal of pharmaceutical sciences 6(2): 7-12.
- Hoffman DL (1987) The herb user's guide. Thorsons publishing group. Welling borough, UK.
- Lawless J (1995) The illustrated encyclopedia of essential oils. Element books Ltd, Shaftesbury, UK.
- Nahal-Bouderba N, Kadi H, Moghtet S, Meddah B, Moussaoui A (2012) Antibacterial activity and phytochemical screening of *Oleaeuropaea* leaves from Algeria. The open conference proceeding journal 3: 66-69.
- Kran KD, Diallo HA, Kouadio YJ (2009) Activités antifongiques de l'extrait de *Chromolaenaodorata* (L) King & Robins sur deux isolats de *Fusarium oxysporum* (EFSm) responsables du jaunissement mortel des feuilles des bananiers. Journal of Applied Biosciences 24: 1488-1496.

18. Cseke, Setzer W, Vogler B, Kirakosyan A, Kaufman P (2006) Traditional, analytical and preparative separation of natural products. Natural products from plants, CRC press/Taylor and francis, Boca Raton, Fla, USA, pp. 263-318.
19. Grayer RJ, Harborne JB (1994) A Survey of antifungal compounds from higher plants. *Phytochemistry* 37: 19-42.
20. Yin MC, Cheng WS (1998) Inhibition of *Aspergillus niger* and *Aspergillus flavus* by some herbs and spices. *Journal of food protection* 61(1): 123-125.
21. Emmanuel MA, Moudachirou M, Akakpo AJ, Quetin Leclercq J (2002) Activités antibactériennes in vitro de *Cassia alata*, *Lantana camara* et *Mitracarpus cabers* sur *Dermatophilus congolensis* isolé au Bénin. *Faculté des Sciences de la santé, Université d'Abomey Calavi Cotonou, Bénin, Revue Pathologie Infectieuse communication* 55(3): 183-187.
22. Omulokoli E, Khan B, Chhabra S (2007) Antiplasmodial activity of four Kenyan medicinal plants. *Journal of Ethnopharmacology* 56(2): 133-137.
23. Soro S, Ouattara D, Guédé NZ, Coffi K (2010) Effet Inhibiteur in Vitro et in Vivo de l'extrait de Poudre et de l'huile Essentielle de *Xylopia aethiopica* (Dunal) A Rich (Annonaceae) sur *Fusarium oxysporum* fsp *Radicis-lycopersici* (Forl), Champignon Parasite des Cultures de Tomate. *European Journal of Scientific Research* 39(2): 279-288.
24. Fogliani B, Raharivelomanana P, Bianchini JP, Bouraima Madjébi S, Hnawia E (2005) Bioactive ellagitannins from *Cunoniaceae*, an endemic *Cunoniaceae* from New Caledonia-*Phytochemistry* 66: 241-247.
25. Mshvildadze V, Favel A, Delmas F, Elias R, Faure R, et al. (2000) Antifungal and antiprotozoal activities of saponins from *Hedera colchica*. *Pharmazie* 55(4): 325-326.
26. Abdel ghani SB, Weaver L, Zidan ZH, Hussein MA, Keevil CW, et al. (2008) Microwave-assisted synthesis and antimicrobial activities of flavonoid derivative. *Bioorganic and medical chemistry letters* 18(2): 518-522.
27. Belsem M, Marzouk Z, Décor R, Edziri H (2009) Antibacterial and anticandidal screening of Tunisian *Citrullus colocynthis* Schrad From Medenine. *Journal of Ethnopharmacology* 125(2): 344-349.
28. Curtay, Jung (2010) Jus de grenade fermenté. (2nd Edn.). Macro pitteur, Belgique p. 10-20.
29. Hibar K, Daami-Remadi M, Jabnoun-Khiareddine H, Znaidi I, Elmahjoub M (2006) Effet des extraits de compost sur la croissance mycélienne et l'agressivité du *Fusarium oxysporum* f. *radicislycopersici*. *Biotechnol Agron Environ* 10(2): 101-108.
30. Imtiaj A, Lee TS (2007) Screening of antibacterial and antifungal activities from Korean wild mushrooms. *World journal of agricultural sciences* 3(3): 316-321.
31. Murray PR, Baron EJ, Pfaller MA, Tenover FC, Tenover RH (1999) *Manual of clinical microbiology*. ASM: Washington, USA, 6: 51-59.
32. Pitt JI (1973) An appraisal of identification methods for *Penicillium* species: Novel taxonomic criteria based on temperature and water relations. *Mycology* 65(5): 1135-1157.
33. Ramirez C (1982) *Manual and Atlas of Penicillia*. Elsevier biomedical press, New York, USA.

ISSN: 2574-1241

DOI: 10.26717/BJSTR.2020.26.004316

Khalid Mohamed Khalid Elhussain. Biomed J Sci & Tech Res



This work is licensed under Creative Commons Attribution 4.0 License

Submission Link: <https://biomedres.us/submit-manuscript.php>



Assets of Publishing with us

- Global archiving of articles
- Immediate, unrestricted online access
- Rigorous Peer Review Process
- Authors Retain Copyrights
- Unique DOI for all articles

<https://biomedres.us/>