Blood profile and Carcass Characteristics of West African Dwarf Goats Fed Groundnut Haulms and Cowpea Husk Supplemented with Brewers’ Dried Grain

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ABSTRACT

Blood profile and carcass characteristics of West African dwarf (WAD) goats fed groundnut haulms and cowpea husk supplemented with brewers’ dried grain, were investigated. Twelve growing WADn goats of average age of 12 months, mean live weight of 12 kg, were randomly allotted to four dietary treatments of T1, T2, T3 and T4 that had groundnut haulms at 10, 30, 70 and 90 % combined with cowpea husks at 90, 70, 30 and 10% basal diet and supplemented with 150g of brewers’ dried grain (BDG). Randomized Complete Block Design (RCBD) was employed in the study. Parameters determined were growth, carcass characteristics and hematological indices. All data obtained were subjected to analysis of variance (ANOVA) using the SAS (2001) package. Means were separated using Duncan’s multiple range test (Duncan, 1955). Effects of the experimental diets on blood profiles of the animals showed PCV values of 39.61 to 41.23 %, haemaglobin (Hb) 5.75 – 6.55g/dl, White blood cells, 9.20 to 11.00, Red blood cell counts (RBC) 4.75 – 6.55mg/dl. Total proteins (mg dl-1), albumin (mg dl-1) and globulin (mg dl-1) of 3.95-4.51, 3.45-4.51 and 3.72-4.32 respectively were obtained in this study. Results of total proteins (g dl-1) showed no significant (P>0.01) differences between treatments T1 – T3. In case of albumin (mg dl-1), there were significant (P<0.01) difference between treatments. Values for globulin (mg dl-1), also showed significant difference (P<0.01) among the treatment. For carcass parameters significant (p<0.05) differences were obtained across treatments with the exceptions of weights of flanks, rack and loins.

Introduction

Meat is one of the most important foods in the world and in some countries; it is considered an essential product with high consumption rate Ana et al. [1]. Differences in carcass, fat and conformation affect meat quality Panea et al. [2]. Carcass dimensions give information on its development, helping in determining the main assessment indices Eniolorunda et al. [3]. Carcass conformation is an important indicator of commercial value because carcass with better conformation has advantages of high lean content, proportion of high price cuts and greater muscle area [Kempten et al.]. A carcass composition determines yield and meat sensorial characteristics. Therefore, carcass economic value is based on its conformation and composition Defa et al. [4]. It is reported Ibhaze et al. [5] that the nutritional and health status of animals cannot be compromised, hence the health status of the animal requires close scrutiny and proper examination of the blood. This fact is further strengthened by Babatunde et al. [6] that haematological parameters are important media used to monitor and evaluate the health and nutritional statuses of animals.

Therefore, the most readily available and fast means of assessing clinical and nutritional effects of feeding trials is by the use of blood analysis Babatunde et al. [6]. Ibhaze [7] further stated that variation in blood parameters of animals are due to several factors such as feeding levels, age, sex, breed and physiological status of the animal. The differences have underlined the need to establish appropriate physiological baseline values for various breeds of livestock in Nigeria which could help in a realistic evaluation of
the management practices, nutrition and diagnosis of their health conditions Opara et al. [8]. Determination of blood chemistry parameters such as glucose, total protein, blood urea nitrogen and cholesterol have been used in assessing nutritional status in cattle Gleen et al. [9]. Reduced feed or protein consumption in goats alters some blood chemistry profiles such as glucose, lipids and proteins Irkham et al. [10]. Aletor et al. [11] reported that the effects of diets on blood and serum chemistry should be of paramount interest since blood transports gases, nutrients and excretory products within the body. Information on blood profile and carcass characteristics of West African dwarf goats fed groundnut haulms and cowpea husk supplemented with brewers’ dried grain is scanty. The research was therefore carried out to bridge this gap.

Materials and Methods

Experimental Site

The experiment was conducted at the Teaching and Research Farm of the Faculty of Agriculture, Adamawa State University, Mubi, Nigeria. Mubi is situated in the northern part of Adamawa State on Latitude 90° 11 I north of the equator and Longitude 13° 45 E east of the Greenwich Meridian at an altitude of 696 m above sea level. It has a land area of 4,728.77 m² and population of 245,460, Mubi region falls within the Sudan Savanna vegetation zone of the country (Saidu and Gadiga).

Experimental Animals and Their Management

Twelve West African Dwarf bucks of average age of 12 months and mean live weight of 12 kg were sourced from the local markets in and around Mubi. Their ages were determined through their dental formulae. They were then housed individually in pens measuring 1.5m² and 1.5m high. The floor of the house was of concrete and covered with wood shavings to absorb moisture from the animals’ dung and urine. The animals were quarantined for two weeks during which they were fed the experimental diets for adaptation and dewormed with Albendazol. At the end of the adaptation period, the animals were tagged, randomly allocated to treatments and balanced on weight basis for all the treatments. They were weighed to get the initial weights before embarking on data collection.

Experimental Diets

There were four treatments with each treatment replicated three times, making a total of twelve experimental animals. The treatments were T1, T2, T3, and T4 that received groundnut haulms at 10, 30, 70 and 90 % combined with cowpea husks at 90, 70, 30 and 10% basal diet which was supplemented with 200g of brewers’ dried grain (BDG) across treatments. Randomized Complete Block Design (RCBD) was employed in the study Table 1.

Table 1: Experimental Diets.

<table>
<thead>
<tr>
<th>Treatments</th>
<th>T1</th>
<th>T2</th>
<th>T3</th>
<th>T4</th>
</tr>
</thead>
<tbody>
<tr>
<td>GNHH (%)</td>
<td>10</td>
<td>30</td>
<td>70</td>
<td>90</td>
</tr>
<tr>
<td>CPHK (%)</td>
<td>90</td>
<td>70</td>
<td>30</td>
<td>10</td>
</tr>
</tbody>
</table>

Parameters Measured

Parameters determined were proximate compositions of feed ingredients, daily dry matter intakes, and daily weight changes, digestibility of treatment diets, feed efficiencies, nutrients utilization and hematological indices. Proximate compositions were determined using the methods described by AOAC (2005). Metabolizable were determined using bomb calorimeter. In determining the feed and nutrients utilization, the diets under investigation were given to the animals in known amounts. Each of the animals was restrained in steel metabolism cages designed to collect urine and feces separately. Urine was collected in 10N H₂SO₄ and the volume for each animal was measured and sampled daily for a period of fourteen days. Fecal outputs were also collected and weighed daily. The proximate compositions of feces were determined using the methods described by AOAC (2005). Protein and Nitrogen levels in feed, faeces and urine were determined by standard Kjeldahl procedure (AOAC, 2005). Two samples were collected from each treatment. Hematological parameters were determined by drawing 10mls of blood from jugular veins of each animal into a sample bottle containing anticoagulant, Ethylene Diamine Tetra acetic Acid (EDTA).

These samples were used for the determination of the packed cell volume (PCV), Haemoglobin (Hb) concentration, Red blood cell counts (RBC) and White blood cell counts (WBC). The Total serum protein, Serum albumin, globulin and blood glucose were determined in the blood serum. The serum was obtained by allowing the blood to coagulate in order to separate the serum and cells. Packed cell volume was measured for each animal using the micro hematocrit method. Haemoglobin concentration was also measured using the Sahli’s (acid haematin) method (Oni et al.). Red blood cells were determined with the aid of Neubaur counting chamber (Haemocytometer). Blood smears were used for total white blood cell (WBC) counts determination. Serum samples were also taken and used in the determination of serum urea, total cholesterol, total protein and albumin as described by Fasuyi et al. All data obtained were subjected to analysis of variance (ANOVA) using the SAS (2001) package. Means were separated using the Duncan’s multiple range test (Duncan).

Results and Discussion

Effects of the experimental diets on blood profiles of the experimental animals are presented in Table 2. The PCV values which indicates the relative proportion of plasma and red blood cells of 39.61 to 41.23 % obtained in this study were higher than 22 – 31 % reported by Daramola et al. [12] for healthy female WAD goats and compared favorably with 36.9 - 40.12 % obtained by Taiwo et al. [13]. The haemoglobin (Hb) of 5.75 - 6.55g/dl obtained in this study were not far from the 9.9 g/dl reported by Opara et al. [8] for healthy female WAD goats, indicating the absence of microcytic hypochromic anaemia caused by iron deficiency and improper utilization for the formation of haemoglobin Olafoadeh
Effects of the diets on carcass characteristics of goats are shown in Table 3. The weights of blood ranged from 0.93 (T<sub>1</sub>) to 1.14 Kg (T<sub>2</sub>). The values were statistically significant (p<0.05) but did not follow any definite trend. The Hides/skin, non carcass, digestive organs, fore and hind limbs weights were significantly (p<0.05) different across treatments. These values in this study were similar to those reported by Ukanwoko et al. [17], when they fed West African dwarf goats with cassava leaf meal based diets. The similarities in this study may be due to the inclusion of the gut, head, legs and internal organs such as heart, kidney, lungs, spleen and liver in the dressed carcass. Cassey et al. [18] had reported that dressing percentage can be influenced by many factors such as fleece and hide weight, alimentary trait size and fill, slaughtering procedure and portioning of body fat. Also the dressing method can affect the dressing percentage because parts which are considered as offal may not be considered offal in some dressing methods. For instance, Fasae et al. considers hot carcass weight to be carcass weight that includes head, gastrointestinal tract but minus legs [19]. However, Ukanwoko et al. reported warm carcass weight to exclude all internal organs, skin, head, feet as well as the visceral and abdominal fat. This compared to the former will give a lower dressing percentage. Effects of the diets on whole sale cuts of growing West African dwarf goats showed that the weight for the breast, shoulder, legs and shanks were significantly (p<0.05) different. However, the weights for flanks, racks, loins and total fat deposit showed no significant (p>0.05) differences. Treatment T<sub>2</sub> which received the highest level of groundnut haulms had the highest values for all the parameters.

Table 3: Effects of Diets on carcass characteristics of goats.

<table>
<thead>
<tr>
<th>Parameters</th>
<th>T&lt;sub&gt;1&lt;/sub&gt;</th>
<th>T&lt;sub&gt;2&lt;/sub&gt;</th>
<th>T&lt;sub&gt;3&lt;/sub&gt;</th>
<th>T&lt;sub&gt;4&lt;/sub&gt;</th>
<th>SEM</th>
<th>Sig Lev</th>
</tr>
</thead>
<tbody>
<tr>
<td>Live weight (Kg)</td>
<td>1.275&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.295&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.175&lt;sup&gt;c&lt;/sup&gt;</td>
<td>1.290&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.23</td>
<td>*</td>
</tr>
<tr>
<td>Blood weight (Kg)</td>
<td>0.95&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.08&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.95&lt;sup&gt;a&lt;/sup&gt;</td>
<td>1.14&lt;sup&gt;a&lt;/sup&gt;</td>
<td>0.05</td>
<td>*</td>
</tr>
<tr>
<td>Hides/skin (Kg)</td>
<td>2.90a</td>
<td>2.95a</td>
<td>2.78&lt;sup&gt;a&lt;/sup&gt;</td>
<td>2.78b</td>
<td>0.67</td>
<td>*</td>
</tr>
<tr>
<td>Non carcass (Kg)</td>
<td>2.77b</td>
<td>2.95&lt;sup&gt;a&lt;/sup&gt;</td>
<td>3.20a</td>
<td>3.22a</td>
<td>0.72</td>
<td>*</td>
</tr>
<tr>
<td>Digest. organs (Kg)</td>
<td>3.23a</td>
<td>2.33b</td>
<td>3.15a</td>
<td>3.05a</td>
<td>0.69</td>
<td>*</td>
</tr>
<tr>
<td>Fore/hind limbs (Kg)</td>
<td>2.98a</td>
<td>2.98a</td>
<td>2.77b</td>
<td>2.94a</td>
<td>0.43</td>
<td>*</td>
</tr>
<tr>
<td>Breast (Kg)</td>
<td>0.88c</td>
<td>0.85c</td>
<td>0.95b</td>
<td>1.10a</td>
<td>0.03</td>
<td>*</td>
</tr>
<tr>
<td>Shoulder (Kg)</td>
<td>1.61c</td>
<td>1.91a</td>
<td>2.02a</td>
<td>1.72b</td>
<td>0.04</td>
<td>*</td>
</tr>
<tr>
<td>Legs (Kg)</td>
<td>1.11b</td>
<td>1.21a</td>
<td>1.32a</td>
<td>1.26a</td>
<td>0.03</td>
<td>*</td>
</tr>
<tr>
<td>Shanks (Kg)</td>
<td>0.97b</td>
<td>0.98b</td>
<td>1.05a</td>
<td>1.08a</td>
<td>0.02</td>
<td>*</td>
</tr>
<tr>
<td>Flanks (Kg)</td>
<td>0.92</td>
<td>0.94</td>
<td>0.95</td>
<td>0.96</td>
<td>0.04</td>
<td>NS</td>
</tr>
<tr>
<td>Racks/Loins (Kg)</td>
<td>1.30</td>
<td>1.32</td>
<td>1.32</td>
<td>1.28</td>
<td>0.05</td>
<td>NS</td>
</tr>
<tr>
<td>Fat (g)</td>
<td>10.45</td>
<td>10.65</td>
<td>10.35</td>
<td>10.23</td>
<td>1.16</td>
<td>NS</td>
</tr>
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</table>
References

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