

Preliminary Blood Compatibility Comparison of Silk Fibroin Dissolved by different Solvents



Ling zhang¹, Mengyao Ding¹, Guangzhou Song¹, Wei Tian¹, Helei Li¹, Honggen Yi¹, Yin Yin², Fenglin Dong³ and Jiannan Wang^{1*}

¹National Engineering Laboratory for Modern, China

²Laboratory Animal Research Centre, China

³Department of Ultrasonography, China

Received:  November 29, 2018; **Published:**  December 17, 2018

***Corresponding author:** Jiannan Wang and Ling zhang National Engineering Laboratory for Modern Silk, College of Textile and Clothing Engineering, Suzhou Industrial Park, Suzhou, Jiangsu Province, 215123, China

Abstract

Silk fibroin possesses good cytocompatibility and biodegradability, was a potential material for wound haemostasis. Ca^{2+} acts as a coagulation factor in cascade pathway for thrombin generation, it can be chelated to silk fibroin molecule. We used three neutral salt solvents (CaCl_2 , $\text{Ca}(\text{NO}_3)_2$ and LiBr) to dissolve silk fibers and prepare regenerated silk fibroin materials, preliminarily explored blood compatibility. Results showed that the regenerated silk fibroin materials prepared by calcium salt dissolving could shorten thrombin time and plasma recalcification time, compared with non-calcium salt reagent LiBr .

Keywords: Silk Fibroin; Calcium Salt; Lithium Salt; Hemolysis; Blood Coagulation

Introduction

Haemorrhage still remains a leading cause of early death and infectious complications in trauma or combat wounds continue to challenge caregivers. Various materials (such as gelatine, fibrin glue, chitosan, collagen, and inorganic microporous materials) have been developed as haemostatic materials [1-5]. On market currently, gelatine and chitosan have attracted several clinically proven products for wound haemostasis. Collagen is also considered a promising biomaterial for wound haemostasis [1,5], but it does not have the ability to induce the rapid hemostasis required for surgical applications [6,7]. Chitosan, as a haemostatic material, is able to stop low pressure arterial bleeds, but is flimsy to maintain hemostasis after intravenous administration of fluids, leading to the risk of rebleeding [8]. After modification, the haemostatic efficiency of chitosan can be improved [9,10]. Collagen, gelatine and chitosan do not have physiological haemostatic functions.

In the coagulation mechanism, Ca^{2+} is indispensable in cascade pathway for thrombin generation. On a wound, Ca^{2+} interacts with platelet surface phospholipids to activate thrombin, and then digest fibrinogen into fibrin, resulting in coagulation formation. So,

calcium alginate has been paid more attention to the applications in embolization [11-13]. However, the cytocompatibility of calcium alginate is not satisfactory. Therefore, it is desired to develop a haemostatic materials with comprehensive ability, combining physical properties, good cytocompatibility and physiologically haemostatic properties. Silk fibroin is a natural animal protein, rich in sources, however, the developments on the haemostatic function of SF materials are lacking. Silk fibroin possesses good cytocompatibility and biodegradability, has been proved for cells adhere and proliferate [14-16], also has been considered as potential tissue engineering scaffolds and drug delivery carriers [16-21]. Interestingly, silk fibroin has a large number of functional groups, such as $-\text{COOH}$ of glutamic acid or aspartic acid, $-\text{NH}_2$ of lysine and $-\text{OH}$ of serine or Tyrosine [22-24], which can interact with Ca^{2+} by electrostatic interaction and coordination bonds. In the study, we prepared silk fibroin porous haemostatic materials by different solvents, and investigated Hemolysis and clotting capability, expecting to expand the application of silk fibroin in haemostatic materials.

Materials and Methods

Preparation of Silk Fibroin Solution

Raw silk was treated with boiling Na_2CO_3 solution to remove sericin, as described previously [16]. After drying, degummed silk fibre was dissolved in a ternary solvent $\text{CaCl}_2 \cdot \text{CH}_3\text{CH}_2\text{OH} \cdot \text{H}_2\text{O}$ (mole ratio, 1:2:8) at 70 °C or 9.3 M of LiBr at 60 °C or molten $\text{Ca}(\text{NO}_3)_2$ at 85 °C and stirred until completely dissolved. After dialyzing against deionized water, concentrating and filtering the mixed solution to obtain silk fibroin aqueous solution (6%, w:w).

Preparation of Silk Fibroin Materials

6% of silk fibroin aqueous solution were mixed with polyethylene glycol diglycidyl ether (Sigma, USA) at a weight ratio of 1.0: 0.8(w/w) and stirred to ensure entirely mixing. The mixture was cast into 6-well cell culture plate, then prefrozen at -80 °C for 8h and freeze-dried for 24h to prepare porous materials. After impregnation in deionized water at 4 °C for 72h to remove unreacted residues, porous materials were freeze-dried again. Regenerated silk fibroin materials were named to SF- CaCl_2 , SF-LiBr, SF- $\text{Ca}(\text{NO}_3)_2$, respectively. Commercial calcium alginate porous material was used as a control.

Hemolysis Assay

Specimens were equilibrated in saline (0.9%,w:v; NaCl) at 37 °C for 30min. Whole blood from a healthy rabbit (2.5kg, New Zealand; the Laboratory Animal Research Centre, Soochow University, China) was collected into sterile sodium citrate buffer [25], then the specimens were soaked in diluted rabbit blood (0.2ml in 10ml saline) to incubate at 37 °C for 60min. Distilled water and physiological saline were used as positive and negative controls, respectively. After centrifuging at 2500rpm for 5min, the absorbance of the supernatant was recorded at 545nm using a Synergy HT microplate reader (BIO-TEK, USA). The Hemolysis percentage was calculated according to the following equation, where A_1 , A_2 , and A_3 are the absorbance of the negative control, sample, and positive control, respectively. All samples were analysed in triplicate.

$$\text{Hemolysis rate (\%)} = \frac{A_2 - A_1}{A_3 - A_1} \times 100\%$$

Coagulation Time Testing

Fresh blood from a rabbit (2.5kg, New Zealand) was collected and mixed with sterile sodium citrate. Specimens were placed in mixture solution and incubated at 37 °C for 30min. After removing samples, the Platelet-Poor Plasma (PPP) supernatant was collected by centrifuging at 3000rpm for 5min. Activated Partial Thrombin Time (APTT), Prothrombin Time (PT) and Thrombin Time (TT) were determined according to the instructions supplied with the relevant kits (Nanjing Jian cheng Bioengineering Institute, China). Briefly, APTT assay, 50 μL PPP supernatant was mixed with 50 μL APTT reagent and incubated at 37 °C for 3min, followed by the addition of 50 μL preheated CaCl_2 (0.03mol/L) and mixing; PT assay, 50 μL PPP supernatant was incubated at 37 °C for 3min, then mixed with 100 μL preheated PT reagent; TT assay, 100 μL PPP supernatant was incubated at 37 °C for 3min and mixed with 100 μL TT reagent.

After the addition of the activators in the above procedures, the time taken for the first clotting to appear was recorded instantly. All samples were analysed in quintuplicate.

Plasma Recalcification Time (PRT) Testing

Whole blood in sodium citrate buffer was centrifuged at 1500rpm for 15min to obtain Platelet-Rich Plasma (PRP) supernatant. Specimens were pre-equilibrated with saline for 24h and further incubated with PRP at 37 °C for 3min, then added 0.025M CaCl_2 to accelerate recalcification at 37 °C. PRP supernatant without materials was used as control group. The time taken for fibrin clots to appear was recorded as the PRT. All samples were analysed in triplicate.

Statistics

Data are presented as mean \pm standard deviation of the mean. Comparisons of means were performed using one-way Analysis of Variance (ANOVA), followed by the independent sample t-test using SPSS 17.0 statistical software. A p-value lower than 0.05 was considered as statistically significant.

Results and Discussion

Hemolysis of Silk Fibroin

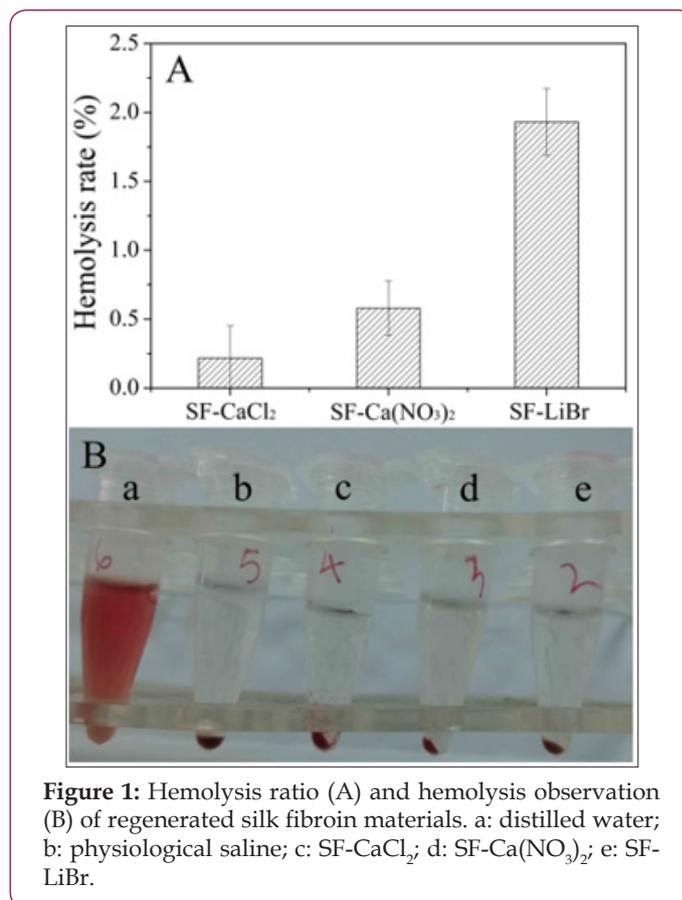


Figure 1: Hemolysis ratio (A) and hemolysis observation (B) of regenerated silk fibroin materials. a: distilled water; b: physiological saline; c: SF- CaCl_2 ; d: SF- $\text{Ca}(\text{NO}_3)_2$; e: SF-LiBr.

Lithium salt (such as LiBr and LiSCN) and calcium salt (such as CaCl_2 and $\text{Ca}(\text{NO}_3)_2$) are recognized solvents used to prepare regenerated silk fibroin materials [16,26,27]. In this work, we used LiBr, CaCl_2 and $\text{Ca}(\text{NO}_3)_2$ to dissolve silk fibers. Figure 1 showed the

haemolytic properties of regenerated silk fibroin porous materials. The Hemolysis ratios of regenerated silk fibroin materials prepared by different dissolution methods were all less than 2% (Figure 1A) and were therefore non-hemolytic [28]. All samples did not damage the red blood cells any more than did the negative control (physiological saline) (Figure 1B). In contrast, the positive control (distilled water) caused extensive damage to the red blood cells, which released large quantities of haemoglobin as a result.

Coagulation Time Analysis of Silk Fibroin

Coagulation system is complex involving various factors that act on both intrinsic and extrinsic activation pathways. Herein, APTT, PT and TT were investigated to evaluate clotting capability of regenerated silk fibroin porous materials dissolved by different solvents (Figure 2). The PT value of SF-CaCl₂ and SF-Ca(NO₃)₂ were shorter than that of SF-LiBr, similar to calcium alginate (Figure 2A). Thromboplastin is the key factor in blood coagulation, in the PT test, there was no obvious difference in the results between samples due

to the addition of sufficient thromboplastin and appropriate Ca²⁺. The APTT reflected the clotting ability depending on Ca²⁺ activating. Although silk fibroin dissolved with calcium salt contained a small amount of chelated Ca²⁺ and a little CaCl₂ reagent were added, it is far less than that of calcium alginate. Therefore, the APTT values of silk fibroin materials were much longer than that of calcium alginate (Figure 2B). In the TT test by adding prothrombin without Ca²⁺, the TT values for silk fibroin materials dissolved by calcium salt were significantly shorter than that by LiBr, while slightly longer than that of calcium alginate (Figure 2C). In this test, Ca²⁺ is also indispensable and played a significant role to activate platelets to release thrombin activating factor, thereby converting prothrombin into thrombin. This Ca²⁺ was only derived from plasma or materials, so the TT values of SF-CaCl₂ and SF-Ca(NO₃)₂ were shorter than that of SF-LiBr with significant differences, although there was only a small amounts of chelated Ca²⁺ in calcium salt dissolved silk fibroin materials.

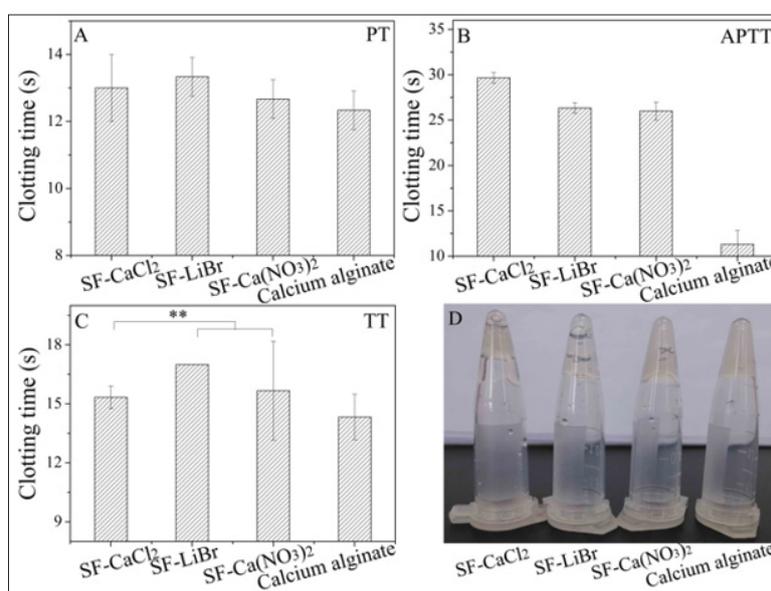


Figure 2: Clotting capability (A-C) and clotting observation (D) of regenerated silk fibroin materials. **p<0.01.

Recalcification of Silk Fibroin

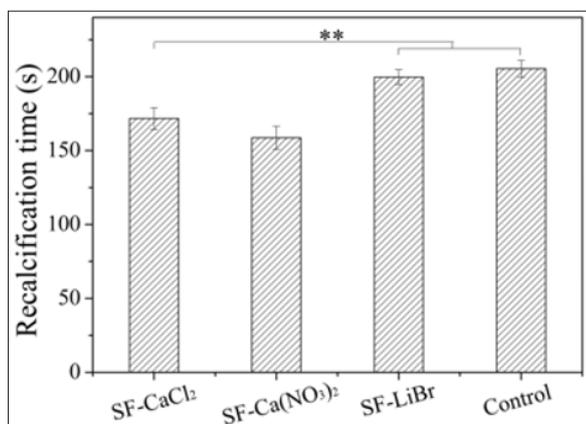


Figure 3: Recalcification time of regenerated silk fibroin materials **p<0.01.

Plasma recalcification time is a convenient measure of the endogenous anticoagulation properties of biomaterials. The recalcification time of regenerated silk fibroin materials dissolved with CaCl₂ and Ca(NO₃)₂ was significantly shorter than that with LiBr (Figure 3) and that of control. The results also because of remained bivalent Ca²⁺, which chelated with -NH₂ or -OH of silk fibroin, activated rich platelets to release thrombin activating factor and promote fibrin formation, resulting in shorter recalcification time of regenerated silk fibroin prepared by calcium salt dissolving.

Conclusion

Silk fibroin molecules can chelate with Ca²⁺, a small amount of chelated Ca²⁺ remained in the silk fibroin materials by calcium salt dissolving and dialyzing. Results showed that the regenerated silk fibroin materials prepared with calcium salt dissolution had a prominent advantage for shortening clotting time compared with non-calcium salt reagent. The study provided an idea to develop silk

fibroin haemostatic materials by using a more suitable preparation method, such as regulating the Ca²⁺ concentration.

Acknowledgement

This work was supported by National Natural Science Foundation of China [Nos 51473108, 51873141], Natural Science Foundation of Jiangsu Province of China [No BK20181192] and College Natural Science Research Project of Jiangsu Province of China [No 18KJA540001].s

References

- Alpaslan C, Alpaslan GH, Oygur T (1997) Tissue reaction to three subcutaneously implanted local hemostatic agents. *Br J Oral Maxillofac Surg* 35(2): 129-132.
- Carraway JW, Kent D, Young K, Cole A, Friedman R, et al. (2008) Comparison of a new mineral based hemostatic agent to a commercially available granular zeolite agent for hemostasis in a swine model of lethal extremity arterial hemorrhage. *Resuscitation* 78(2): 230-235.
- Sambasivan CN, Cho SD, Zink KA, Differding JA, Schreiber MA, et al. (2009) A highly porous silica and chitosan-based hemostatic dressing is superior in controlling hemorrhage in a severe groin injury model in swine. *Am J Surg* 197(5): 576-580.
- Al Qahtani K (2011) Initial experience with hemostatic fibrin glue as adjuvant during drainless parotidectomy. *Saudi Dent J* 23(2): 67-71.
- Manon Jensen T, Kjeld NG, Karsdal MA, Thromb J (2016) Collagen-mediated hemostasis. *J Thromb Haemost* 14(3): 438-448.
- Gazzeri R, Galarza M, Fiore C, Calloveni G, Alfieri A (2015) Use of tissue-glue-coated collagen sponge (Tachosil) to repair minor cerebral dural venous sinus lacerations: technical note. *Neurosurgery* 11 Suppl 2: 32-36.
- Wong C, Liu Y, Yip J, Chand R, Wee JL, et al. (2009) CEACAM1 negatively regulates platelet-collagen interactions and thrombus growth in vitro and in vivo. *Blood* 113(8): 1818-1828.
- Kheirabadi BS, Scherer MR, Estep JS, Dubick MA, Holcomb JB (2009) Determination of efficacy of new hemostatic dressings in a model of extremity arterial hemorrhage in swine. *J Trauma* 67(3): 450-460.
- Bochicchio G, Kilbourne M, Kuehn R, Keledjian K, Hess J, et al. (2009) Use of a modified chitosan dressing in a hypothermic coagulopathic grade V liver injury model. *Am J Surg* 198(5): 617-622.
- Chan LW, Kim CH, Wang X, Pun SH, White NJ, et al. (2016) PolySTAT-modified chitosan gauzes for improved hemostasis in external hemorrhage. *Acta Biomater* 31: 178-185.
- Jarvis PM, Galvin DAJ, Blair SD, Mccollum CN (1987) How does calcium alginate achieve haemostasis in surgery? *J Thromb Haemost* 58(01): 080.
- Xu GZ, Cheng L, Zhang QT, Sun YL, Chen CL, et al. (2016) In situ thiolated alginate hydrogel: Instant formation and its application in hemostasis. *J Biomater Appl* 31(5): 721-729.
- Forster REJ, Thurmer F, Wallrapp C, Lloyd AW, Macfarlane W, et al. (2010) Characterisation of physico-mechanical properties and degradation potential of calcium alginate beads for use in embolization. *J Mater Sci Mater Med* 21(7): 2243-2251.
- Sun XL, Hao YX, Wang QY, Dong FL, Wang JN (2016) Cell growth and proliferation on the interface of a silk fabric tubular scaffold. *Textile Res J* 86: 2193-2201.
- Tu FF, Liu YF, Li HL, Shi PG, Hao YX, et al. (2018) Vascular cell co-culture on silk fibroin matrix. *Polymers* 10(1): 39.
- Wang JN, Wei YL, Yi HG, Liu ZW, Sun D, et al. (2014) Cytocompatibility of a silk fibroin tubular scaffold. *Mater Sci Eng C Mater Biol Appl* 34: 429-436.
- Wei YL, Sun D, Yi HG, Wang JN (2014) Characterization of a PEG-DE cross-linked tubular silk scaffold. *Textile Res J* 84(9): 959-967.
- Liu YF, Tu FF, Li HL, Shi PG, Yin Y, et al. (2018) Preparation, characterization and in vivo graft patency of a silk fibroin tubular scaffold. *Mater Technol* 33: 227-234.
- Farokhi M, Mottaghtalab F, Samani S, Shokrgozar MA, Kundu SC, et al. (2018) Preparation of a codelivery system based on vancomycin/silk scaffold containing silk nanoparticle loaded VEGF. *ACS Biomater Sci & Eng* 4(8): 2836-2846.
- Chouhan D, Lohe T, Samudrala PK, Mandal BB (2018) In situ forming injectable silk fibroin hydrogel promotes skin regeneration in full thickness burn wounds. *Adv Healthcare Mater*, pp. e1801092.
- Sheikh FA, Ju HW, Lee JM, Moon BM, Park HJ, et al. (2015) 3D electrospun silk fibroin nanofibers for fabrication of artificial skin. *Nanomedicine* 11(3): 681-691.
- Wang JN, Yan SQ, Lu CD, Bai L (2009) Biosynthesis and characterization of typical fibroin crystalline polypeptides of silkworm *Bombyx mori*. *Mater Sci Eng* 29(4): 1321-1325.
- Yang GQ, Wu MY, Yi HG, Wang JN (2016) Biosynthesis and characterization of a non-repetitive polypeptide derived from silk fibroin heavy chain. *Mater Sci Eng C Mater Biol Appl* 59: 278-285.
- Wu Y, Kang Z, Tian ZF, Wu MY, Wang JN (2017) Biosynthesis and characterization of recombinant silk-like polypeptides derived from the heavy chain of silk fibroin. *Polymers* 9(12): 669.
- Sun D, Hao YX, Yang GQ, Wang JN (2015) Hemocompatibility and cytocompatibility of the hirudin-modified silk fibroin. *J Biomed Mater Res B Appl Biomater* 103(3): 556-562.
- Ko E, Lee JS, Kim H, Yang SY, Yang D, et al. (2018) Electrospun silk fibroin nanofibrous scaffolds with two-stage hydroxyapatite functionalization for enhancing the osteogenic differentiation of human adipose-derived mesenchymal stem cells. *ACS Appl Mater Interfaces* 10(9): 7614-7625.
- Mathur AB, Tonelli A, Rathke T, Hudson S (2010) The dissolution and characterization of *Bombyx mori* silk fibroin in calcium nitrate-methanol solution and the regeneration of films. *Biopolymers* 42(1): 61-74.
- (2003) ASTM Medical Devices, Emergency Medical Services: ASTM International 13: 309.

ISSN: 2574-1241

DOI: [10.26717/BJSTR.2018.12.002233](https://doi.org/10.26717/BJSTR.2018.12.002233)

Jiannan Wang. Biomed J Sci & Tech Res



This work is licensed under Creative Commons Attribution 4.0 License

Submission Link: <https://biomedres.us/submit-manuscript.php>



Assets of Publishing with us

- Global archiving of articles
- Immediate, unrestricted online access
- Rigorous Peer Review Process
- Authors Retain Copyrights
- Unique DOI for all articles

<https://biomedres.us/>