

# Agriculturally Important Microbiomes: Biodiversity and Multifarious PGP Attributes for Amelioration of Diverse Abiotic Stresses in Crops for Sustainable Agriculture

Ajar Nath Yadav\*

Department of Biotechnology, Akal College of Agriculture, Eternal University, India

Received: August 24, 2017; Published: September 01, 2017

\*Corresponding author: Ajar Nath Yadav, Department of Biotechnology, Akal College of Agriculture, Eternal University, Sirmour-173101, India,

Tel: +91-9882545085; Email: ajar@eternaluniversity.edu.in

**Abbreviations:** ACC: 1-aminocyclopropane-1-carboxylate; BNF: Biological Nitrogen Fixation; PGP: plant growth promoting

## Introduction

The Microbiomes from plant and different extreme environments have been reported and characterized for genotypic and multifarious functional attributes. The plant Microbiomes (phyllospheric, endophytic and rhizospheric) and microbiomes of extreme habitat (acidophilic, alkaliphilic, halophilic, psychrophilic, thermophilic and xerophilic) are natural bioresources, which may play critical roles in the maintenance of global nutrient balance and ecosystem functions. The crops microbiomes and extreme habitat microbiomes with plant growth promoting (PGP) attributes have emerged as an important and promising tool to enhance plant growth, crop yield and soil fertility. The microbes possess PGP attributes such as solubilization of phosphorus, potassium and zinc; ACC deaminase; biological nitrogen fixation and production of auxin, gibberellic acids, cytokinin, Fe-chelating compounds, ammonia, HCN, hydrolytic enzymes and secondary metabolites. These PGP microbes could be applied as biofertilizers [An eco-friendly agriculturally important bioinoculants] to replace the chemical fertilizers and for amelioration of different abiotic stresses in crops including, salinity, temperature, drought and pH. The agriculturally important microbes may play an important role in plant growth, development, and soil health for sustainable agriculture.

Plant microbiomes and its interaction with plant is a key for plant growth and development. In general, there are three kinds of plant-microbes interactions considered i.e. epiphytic, endophytic and rhizospheric. The rhizosphere is the zone of soil influenced by roots through the release of substrates that affect microbial activity [1,2]. The phyllosphere is a common niche for synergism between microbes and aerial parts of plant [3,4]. The phyllospheric microbes may perform an effective function

in controlling the air borne pathogens inciting plant disease. The endophytic microbes are referred to those microorganisms, which colonize in the interior of the plant parts viz: root, stem or seeds without causing any harmful effect on host plant. The biodiversity of plant microbiomes revealed that the representative microbes from archaea (Euryarchaeota); bacteria (Acidobacteria, Actinobacteria, Bacteroidetes, Deinococcus-Thermus, Firmicutes and Proteobacteria) and fungi (Ascomycota and Basidiomycota) have been characterized genotypically and phenotypically for their beneficial attributes for human welfare.

On the basis of different research, it may be suggested that the distribution of microbes although varied in all bacterial phyla, but proteobacteria were most dominant and ubiquitous followed by actinobacteria. Least number of microbes have been reported from phylum Deinococcus-Thermus and Acidobacteria followed by Bacteroidetes [5-11]. There are very few reports of halophilic archaea as plant growth promoting including rhizospheric as well as endophytic [12,13]. A number of microbial species belonging to different genera such as *Alcaligenes*, *Arthrobacter*, *Azospirillum*, *Bacillus*, *Burkholderia*, *Enterobacter*, *Flavobacterium*, *Haloarcula*, *Halobacterium*, *Halococcus*, *Methylobacterium*, *Paenibacillus*, *Penicillium*, *Piriformospora*, *Pseudomonas*, *Rhizobium* and *Serratia* have been sorted out from rhizosphere of different crops and characterized for different PGP attributes [14-21]. Many microbes such as *Achromobacter*, *Beijerinckia*, *Brevibacterium*, *Diplococcus*, *Flexibacterium*, *Methylobacterium*, *Micrococcus*, *Micromonospora*, *Nocardioidea*, *Pantoea*, *Penicillium*, *Pseudomonas* and *Streptomyces* have been reported as phyllospheric microbes with beneficial attributes for crops under the diverse abiotic stress conditions [22-28]. There are large numbers of endophytic microbiomes including

*Azoarcus*, *Burkholderia*, *Curtobacterium*, *Gluconoacetobacter*, *Herbaspirillum*, *Klebsiella*, *Micromonospora*, *Nocardioides*, *Pantoea*, *Pseudomonas*, *Streptomyces* and *Thermomonospora* have been identified from different host plants [23,28-31].

The plant microbiomes have been shown to be beneficial for different crops by promoting plant growth either directly, e.g. by biological N<sub>2</sub>-fixation, solubilization of phosphorus (P), potassium (K) and zinc (Zn); production of Fe-chelating compounds, cytokinins, auxins and gibberellins or indirectly, via production of antagonistic substances by inducing resistance against plant pathogens. Biological nitrogen fixation (BNF) is one of the possible biological alternatives to N-fertilizers and could lead to more productive and sustainable agriculture without harming the environment. Plant-associated microbes typically produce plant growth hormones such as auxins and gibberellins. The gibberellins production is most typical for the root-associated microbes and auxin production is common to all plant-associated microbes. Auxins can promote the growth of roots and stems quickly or slowly [32-34].

Phosphorus (P) is major essential macronutrient for biological growth and development. Microbes offer a biological rescue system capable of solubilizing the insoluble inorganic P of soil and make it available to the plants. P-solubilization is a common trait among microbes associated with different crops. For instance, the majority of microbial populations from wheat, rice, maize, and legumes were able to solubilize mineral phosphates, and a vast number of PGP microbes with P-solubilizing property have been reported which include members belonging to *Burkholderia*, *Enterobacter*, *Halolamina*, *Pantoea*, *Pseudomonas*, *Citrobacter* and *Azotobacter* [35-41]. Ethylene is a stress-induced plant hormone that can inhibit plant growth. Some microbes can lower the level of ethylene in the plant by cleaving the plant-produced ethylene precursor 1-aminocyclopropane-1-carboxylate (ACC). Ethylene is a key regulator of the colonization of plant tissue by bacteria which in turn suggests that the ethylene inhibiting effects of ACC-deaminase may be a microbial colonization strategy. Generally, ethylene is an essential metabolite for the normal growth and development of plants [42-45]. Microbial strains exhibiting ACC deaminase activity have been identified in a wide range of genera such as *Achromobacter*, *Azospirillum*, *Bacillus*, *Burkholderia*, *Enterobacter*, *Pseudomonas*, *Serratia* and *Rhizobium* [46-49]. The indirect mechanism of plant growth occurs when microbes lessen or prevent the detrimental effects of pathogens on plants by production of inhibitory substances or by increasing the natural resistance of the host. Phytopathogenic microbes can control by releasing siderophores, chitinases, antibiotics, fluorescent pigment or by cyanide production [50,51]. Biocontrol systems are eco-friendly, cost-efficient and involved in improving the soil consistency and maintenance of natural soil flora [52-54]. The microbes with zinc solubilizing and Fe-chelating compounds production attributes may be used for bio-fortification of Fe and Zn for different cereal crops.

## Conclusion and Future Vision

The microbes are capable of colonizing the rhizosphere, phyllosphere as well as living inside the plant tissues as endophytes.

The agriculturally important microbes plays important role in plant growth, development, and soil health for sustainable agriculture. The plant microbiomes (phyllospheric, endophytic and rhizospheric) and microbiomes of extreme habitat (acidophilic, alkaliphilic, halophilic, psychrophilic, thermophilic and xerophilic) are natural bioresources. Due to the diverse range of activities as well as the number of microbes in varying habitats around the world, these are important bioresources towards rationalized use of chemicals fertilizers in agriculture. The microbes having multifarious PGP attributes could be applied as biofertilizers to replace the chemicals fertilizers and for amelioration of abiotic stress under normal and stressed conditions.

## References

1. Yadav AN, Verma P, Kaushik R, Dhaliwal HS, Saxena AK (2017) Archaea endowed with plant growth promoting attributes. *EC Microbiol* 8(6): 294-298.
2. Yadav AN, Verma P, Kour D, Rana KL, Kumar V, et al. (2017) Plant microbiomes and its beneficial multifunctional plant growth promoting attributes. *Int J Environ Sci Nat Resour* 3(1): 1-8.
3. Verma P, Yadav AN, Khannam KS, Kumar S, Saxena AK et al. (2016) Molecular diversity and multifarious plant growth promoting attributes of Bacilli associated with wheat (*Triticum aestivum* L.) rhizosphere from six diverse agro-ecological zones of India. *J Basic Microbiol* 56(1): 44-58.
4. Verma P, Yadav AN, Khannam KS, Mishra S, Kumar S, et al. (2016) Appraisal of diversity and functional attributes of thermotolerant wheat associated bacteria from the peninsular zone of India. *Saudi J Biol Sci*. DOI: 10.1016/j.sjbs.2016.01.042.
5. Sahay H, Yadav AN, Singh AK, Singh S, Kaushik R, et al. (2017) Hot springs of Indian Himalayas: Potential sources of microbial diversity and thermostable hydrolytic enzymes. *Biotech* 7(2): 1-11.
6. Yadav AN (2015) Bacterial diversity of cold deserts and mining of genes for low temperature tolerance. IARI, New Delhi, Ranchi, India, pp. 234
7. Saxena AK, Yadav AN, Rajawat M, Kaushik R, Kumar R, et al. (2016) Microbial diversity of extreme regions: An unseen heritage and wealth. *Indian J Plant Genet Resour* 29(3): 246-248.
8. Shukla L, Suman A, Yadav AN, Verma P, Saxena AK (2016) Syntrophic microbial system for ex-situ degradation of paddy straw at low temperature under controlled and natural environment. *J App Biol Biotech* 4(2): 30-37.
9. Yadav AN, Verma P, Kumar M, Pal KK, Dey R, et al. (2015) Diversity and phylogenetic profiling of niche-specific Bacilli from extreme environments of India. *Ann Microbiol* 65(2): 611-629.
10. Suman A, Verma P, Yadav AN, Saxena AK (2015) Bioprospecting for extracellular hydrolytic enzymes from culturable thermotolerant bacteria isolated from Manikaran thermal springs. *Res J Biotechnol* 10(4): 33-42.
11. Suman A, Yadav AN, Verma P (2016) Endophytic Microbes in Crops: Diversity and Beneficial impact for Sustainable Agriculture. In: Singh DP, Abhilash PC, Prabha R (Eds.), *Microbial Inoculants in Sustainable Agricultural Productivity, Research Perspectives*. Springer-Verlag, India, pp. 117-143.
12. Yadav AN, Sharma D, Gulati S, Singh S, Kaushik R, et al. (2015) Haloarchaea endowed with phosphorus solubilization attribute implicated in phosphorus cycle. *Sci Rep* 5: 12293.
13. Gaba S, Singh RN, Abrol S, Yadav AN, Saxena AK, et al. (2017) Draft genome sequence of *Halolamina pelagica* CDK2 isolated from natural salterns from Rann of Kutch, Gujarat, India. *Genome Announc* 5(6): 1-2.
14. Barea JM, Pozo MJ, Azcon R, Azcon Aguilar C (2005) Microbial cooperation in the rhizosphere. *Journal of experimental botany* 56(417): 1761-1778.

15. Lavania M, Chauhan PS, Chauhan S, Singh HB, Nautiyal CS (2006) Induction of plant defense enzymes and phenolics by treatment with plant growth-promoting rhizobacteria *Serratia marcescens* NBRI1213. *Curr Microbiol* 52(5): 363-368.
16. Tilak K, Ranganayaki N, Pal K, De R, Saxena A, et al. (2005) Diversity of plant growth and soil health supporting bacteria. *Curr Sci* 89(1): 136-150.
17. Yang J, Kloepper JW, Ryu C-M (2009) Rhizosphere bacteria help plants tolerate abiotic stress. *Trends plant sci* 14(1): 1-4.
18. Kumar V, Yadav AN, Saxena A, Sangwan P, Dhaliwal HS (2016) Unravelling rhizospheric diversity and potential of phytase producing microbes. *SM J Biol* 2(1): 1009.
19. Yadav AN, Verma P, Kumar R, Kumar V, Kumar K (2017) Current applications and future prospects of eco-friendly microbes. *EU Voice* 3(1): 1-3.
20. Verma P, Yadav AN, Khannam KS, Panjiar N, Kumar S, et al. (2015) Assessment of genetic diversity and plant growth promoting attributes of psychrotolerant bacteria allied with wheat (*Triticum aestivum*) from the northern hills zone of India. *Ann Microbiol* 65(4): 1885-1899.
21. Yadav AN, Verma P, Kumar V, Sangwan P, Mishra S, et al. (2017) Biodiversity of the Genus *Penicillium* in Different Habitats. In: *New and Future Developments in Microbial Biotechnology and Bioengineering*, vol -1. Elsevier.
22. Hornschuh M, Grotha R, Kutschera U (2002) Epiphytic bacteria associated with the bryophyte *Funaria hygrometrica*: effects of *Methylobacterium* strains on protonema development. *Plant Biol* 4(6): 682-687.
23. Verma P, Yadav AN, Kazy SK, Saxena AK, Suman A (2014) Evaluating the diversity and phylogeny of plant growth promoting bacteria associated with wheat (*Triticum aestivum*) growing in central zone of India. *Int J Curr Microbiol Appl Sci* 3(5): 432-447.
24. Holland MA, Davis R, Moffitt S, O'Laughlin K, Peach D, et al. (2000) Using "leaf prints" to investigate a common bacterium. *Am Biol Teach* 62(2): 128-131.
25. Verma P, Yadav AN, Kazy SK, Saxena AK, Suman A (2013) Elucidating the diversity and plant growth promoting attributes of wheat (*Triticum aestivum*) associated acidotolerant bacteria from southern hills zone of India. *Natl J Life Sci* 10(2): 219-227.
26. Tancos K, Cox K (2017) Effects of Consecutive Streptomycin and Kasugamycin Applications on Epiphytic Bacteria in the Apple Phyllosphere. *Plant Dis* 101(1): 158-164.
27. Dobrovol'skaya T, Khusnetdinova K, Manucharova N, Golovchenko A (2017) Structure of epiphytic bacterial communities of weeds. *Microbiology* 86(2): 257-263.
28. Verma P, Yadav AN, Shukla L, Saxena AK, Suman A (2015) Alleviation of cold stress in wheat seedlings by *Bacillus amyloliquefaciens* IARI-HHS2-30, an endophytic psychrotolerant K-solubilizing bacterium from NW Indian Himalayas. *Natl J Life Sci* 12(2): 105-110.
29. Hallmann J, Quadt-Hallmann A, Mahaffee W, Kloepper J (1997) Bacterial endophytes in agricultural crops. *Can J Microbiol* 43(10): 895-914.
30. Quadt-Hallmann A, Kloepper J, Benhamou N (1997) Bacterial endophytes in cotton: mechanisms of entering the plant. *Can J Microbiol* 43(6): 577-582.
31. Ryan RP, Germaine K, Franks A, Ryan DJ, Dowling DN (2008) Bacterial endophytes: recent developments and applications. *FEMS Microbiol Lett* 278(1): 1-9.
32. Yadav AN, Sachan SG, Verma P, Tyagi SP, Kaushik R, et al. (2015) Culturable diversity and functional annotation of psychrotrophic bacteria from cold desert of Leh Ladakh (India). *World J Microbiol Biotechnol* 31(1): 95-108.
33. Chaiharn M, Lumyong S (2011) Screening and optimization of indole-3-acetic acid production and phosphate solubilization from rhizobacteria aimed at improving plant growth. *Curr Microbiol* 62(1): 173-181.
34. Xie H, Pasternak JJ, Glick BR (1996) Isolation and characterization of mutants of the plant growth-promoting rhizobacterium *Pseudomonas putida* GR12-2 that overproduce indoleacetic acid. *Curr Microbiol* 32(2): 67-71.
35. Kumar V, Yadav AN, Verema P, Sangwan P, Abhishake S, et al. (2017)  $\beta$ -Propeller phytases: Diversity, catalytic attributes, current developments and potential biotechnological applications. *Int J Biol Macromolec* 98: 595-609.
36. Yadav AN, Sachan SG, Verma P, Saxena AK (2015) Prospecting cold deserts of north western Himalayas for microbial diversity and plant growth promoting attributes. *J Biosci Bioeng* 119(6): 683-693.
37. Illmer P, Schinner F (1992) Solubilization of inorganic phosphates by microorganisms isolated from forest soils. *Soil Biol Biochem* 24 (4): 389-395.
38. Kaur R, Saxena A, Sangwan P, Yadav AN, Kumar V, et al. (2017) Production and characterization of a neutral phytase of *Penicillium oxalicum* EUFR-3 isolated from Himalayan region. *Nus Biosci* 9(1): 68-76.
39. Wakelin SA, Warren RA, Harvey PR, Ryder MH (2004) Phosphate solubilization by *Penicillium* spp. closely associated with wheat roots. *Biol Fertil Soils* 40(1): 36-43.
40. Pradhan N, Sukla L (2006) Solubilization of inorganic phosphates by fungi isolated from agriculture soil. *African J Biotechnol* 5(10): 850-854.
41. Yadav AN, Rana KL, Kumar V, Dhaliwal HS (2016) Phosphorus solubilizing endophytic microbes: Potential application for sustainable agriculture. *EU Voice* 2: 21-22.
42. Suman A, Verma P, Yadav AN, Srinivasamurthy R, Singh A, et al. (2016) Development of hydrogel based bio-inoculant formulations and their impact on plant biometric parameters of wheat (*Triticum aestivum* L.). *Int J Curr Microbiol Appl Sci* 5(3): 890-901.
43. Penrose DM, Glick BR (2003) Methods for isolating and characterizing ACC deaminase-containing plant growth-promoting rhizobacteria. *Physiol Plant* 118(1): 10-15.
44. Glick BR, Cheng Z, Czarny J, Duan J (2007) Promotion of plant growth by ACC deaminase-producing soil bacteria. *Eur J Plant Pathol* 119(3): 329-339.
45. Siddikee MA, Chauhan P, Anandham R, Han G-H, Sa T (2010) Isolation, characterization, and use for plant growth promotion under salt stress, of ACC deaminase-producing halotolerant bacteria derived from coastal soil. *J Microbiol Biotechnol* 20(11): 1577-1584.
46. Khalid A, Akhtar M, Mahmood M, Arshad M (2006) Effect of substrate-dependent microbial ethylene production on plant growth. *Microbiology* 75(2): 231-236.
47. Srivastava AK, Kumar S, Kaushik R, Saxena AK, Padaria JC, et al. (2014) Diversity analysis of *Bacillus* and other predominant genera in extreme environments and its utilization in Agriculture.
48. Sun Y, Cheng Z, Glick BR (2009) The presence of a 1-aminocyclopropane-1-carboxylate (ACC) deaminase deletion mutation alters the physiology of the endophytic plant growth-promoting bacterium *Burkholderia phytofirmans* PsjN. *FEMS Microbiol Lett* 296(1): 131-136.
49. Xu M, Sheng J, Chen L, Men Y, Gan L, et al. (2014) Bacterial community compositions of tomato (*Lycopersicon esculentum* Mill.) seeds and plant growth promoting activity of ACC deaminase producing *Bacillus subtilis* (HYT-12-1) on tomato seedlings. *World J Microbiol Biotechnol* 30(3): 835-845.
50. Lottmann J, Heuer H, de Vries J, Mahn A, Düring K, et al. (2000) Establishment of introduced antagonistic bacteria in the rhizosphere of transgenic potatoes and their effect on the bacterial community. *FEMS Microbiol Ecol* 33(1): 41-49.
51. Yadav AN, Sachan SG, Verma P, Kaushik R, Saxena AK (2016) Cold active hydrolytic enzymes production by psychrotrophic Bacilli isolated from three sub-glacial lakes of NW Indian Himalayas. *J Basic Microbiol* 56(3): 294-307.

52. Verma P, Yadav AN, Kumar V, Khan MA, Saxena AK (2017) Microbes in Termite Management: Potential Role and Strategies. In: Sustainable Termite Management Springer.
53. Jha PN, Gupta G, Jha P, Mehrotra R (2013) Association of rhizospheric/endophytic bacteria with plants: a potential gateway to sustainable agriculture. Greener J Agric Sci 3(2): 73-84.
54. Uppal A, El Hadrami A, Adam L, Tenuta M, Daayf F (2008) Biological control of potato *Verticillium* wilt under controlled and field conditions using selected bacterial antagonists and plant extracts. Biol Con 44(1): 90-100.



#### Assets of Publishing with us

- Global archiving of articles
- Immediate, unrestricted online access
- Rigorous Peer Review Process
- Authors Retain Copyrights
- Unique DOI for all articles

<http://biomedres.us/>