

Antioxidant Capacity and Antitumoral Activity of Citrus Paradisi Essential Oil

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Abbreviations: CPEo: Citrus Paradisi Essential Oil; DPPH: Diphenyl Picryl Hydrazine; ABTS: Azino-Bis-3-Ethylbenzothiazoline-6-Sulfonic Acid; FRAP: Ferric-Reducing Antioxidant Power; TCA: Trichloroacetic Acid; FCS: Foetal Calf Serum; SD: Standard Deviation

ABSTRACT

Background: The present study was focused on the study of essential oil extraction from the zest of grapefruit (*Citrus paradisi*) and the determination of antioxidant and antitumor activities.

Methods: Grapefruit essential oil was extracted by hydro distillation, grapefruit essential oil was analyzed for antioxidant potential in the chemical system by the DPPH, ABTS, FRAP test, and scavenging activity of nitric oxide. The cytotoxicity study of grapefruit essential oil was assessed using MTT assay against HeLa and MCF-7 tumor cells.

Results: The extraction yield obtained after optimization was 1.32 %. Antioxidant activities were studied using chemical systems. The results have been very interesting since we got 54 %, 83 %, 66 % respectively with DPPH, ABTS, FRAP, and a notable effect to scavenging NO• radical. In addition, a significant antitumor effect of grapefruit essential oil was observed in HeLa and MCF-7 tumor cells.

Conclusion: The data serve as the basis for the consumption of Citrus paradisi essential oil, with significant antioxidant activity, with no harmful effect for the normal cells, and with antitumor potential.

Keywords: Citrus Paradisi; Essential Oil; Antioxidant Activity; Antitumor Activity; MTT; HeLa; MCF-7

Introduction

Phytomedicine has been of increasing interest in recent years. Over 65 % of the world's population relies on traditional and medical approaches to treating diseases [1]. The return to phytotherapy is more than using an herb to treat an illness or a crude herbal preparation rather than a specific isolated component. Plants are considered essential resources for researchers to prove and develop

new drugs. Generally, using plants in the treatment of disease has a long history [2,3]. Thus plants have been primary resources for producing traditional drugs effective in treating cancer and inflammatory diseases [4]. In addition, plants containing essential oils have bioactivity against a host of bacteria, fungi, viruses [5,6]. Currently, essential oils of medicinal plants are increasingly

used thanks to their antibacterial, anti-inflammatory, antiseptic, antiviral, antifungal, antioxidant, and antitumor potential [7]. Essential oils are also used in other areas of economic interest, such as cosmetics, perfumes, and aromatherapy [7,8]. Fragrant and aromatic plants such as members of the *Asteraceae*, *Lamiaceae*, *Rutaceae*, and *Verbenaceae* produce essential oils, which historically have been important in traditional medicines [5]. Citrus species have beneficial pharmacological activities, including antibacterial, vascular protectant, antispasmodic, analgesic, antipyretic, anti-inflammatory, and antitumoral effects [9-11].

Citrus peel essential oils have also been searched for their natural antioxidant and antimicrobial properties [12,13]. Furthermore, studies have shown the synergistic effect of two or more ingredients of essential oils against various human pathogens. It is necessary to know more about Citrus paradisi essential oil (CPEo) antioxidant, cytotoxic, and antitumor activity. Therefore, the DPPH, ABTS, FRAP, and NO• techniques studied the antioxidant potential in a chemical system. Furthermore, to check the antitumor activity, MTT was realized using HeLa and MCF-7 tumor cells.

Material and Methods

Samples

In this study, the grapefruit (citrus paradisi) was collected in March 2018 from Sfax, Tunisia. Sfax is one of the biggest coastal cities in Tunisia. It is located in the eastern part of the country. The climate in this area is arid to semiarid with irregular and torrential precipitations [14]. After the collection, it was washed, peeled off and cut into small pieces.

Hydrodistillation

The zest of citrus paradisi was hydro distilled using a Clevenger-type apparatus to recuperate the essential oils for 2h to produce the volatile constituents. The distilled essential oils were dried over anhydrous sodium sulfate; then separated from the distillate by liquid-liquid extraction using cyclohexane solvent. The recuperated oils were stored at + 4 °C.

Antioxidant Activity: Chemical System

DPPH Radical Scavenging Assay: Different aliquots from the stock solution (200µl in 1 ml Me OH) of essential oils were mixed with 500µl of 0.2mM diphenyl picryl hydrazine (DPPH) final volume brought to 1mL. The mixtures were vigorously shaken and allowed to stand in the dark for 30min at room temperature. The absorbance was measured by spectrophotometry (LKB BIOCHROM® ULTROSPECUS 4054 UV/VIS) at 517 nm against a control sample without DPPH. The percentage of radical scavenging activity was calculated using the following equation:

$$\text{DPPH scavenging effect (\%)} = (A_0 - A_1)/A_0 \times 100$$

A_0 : The absorbance of the control at 30min

A_1 : The absorbance of the sample at 30min.

ABTS+ Radical Scavenging Effect: The antiradical activity was performed by the ABTS+ free radical decolorization assay as developed by Re, et al. [15]. The 2,2-azino-bis-3-ethylbenzothiazoline-6-sulfonic acid (ABTS) was prepared as an aqueous stock solution (7mM). The ABTS radical cations (ABTS+) were produced by the reaction of the ABTS stock solution with 2.5mM of ammonium persulfate methanolic solution. First, the reaction mixture is incubated in the dark for 16h at room temperature. Then, the solution is diluted to an absorbance of 0.7 ± 0.02 at 734nm to form the working reagent. Next, the reaction mixtures containing 100µl of the sample at different concentrations and 900µL of reagent were incubated at 30 °C for 6min. Finally, the antioxidant power of each sample was expressed as the inhibition percentage calculated according to the following formula:

$$\text{ABTS+ scavenging effect (\%)} = (A_0 - A_1)/A_0 \times 100$$

A_0 : the absorbance of the control at 6min

A_1 : the absorbance of the sample at 6min.

Ferric-Reducing Antioxidant Power (FRAP): This method is based on the plant's ability to reduce ferric iron (Fe^{3+}) to ferrous iron (Fe^{2+}). The mechanism is known to be a marker of electron donor activity [16]. For 1mL of the sample at different concentrations, 2.5mL of a solution phosphate buffer (0.2M, pH 6.6) and 2.5mL of 1 % $\text{K}_3\text{Fe}(\text{CN})_6$ potassium ferricyanide solution were added. The mixture is incubated at 50 °C for 20min and then cooled to room temperature. Then, 2.5mL of 10 % trichloroacetic acid (TCA) is added to stop the reaction, and then the tubes are centrifuged at 3000rpm for 10min. 2.5ml of the supernatant are then added to 2.5mL of distilled water and 500µL of a 0.1 % solution of iron trichloride ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$) [16]. The absorbance reading is performed against a blank at 700 nm using a spectrophotometer. Ascorbic acid is used as a positive control. The increase in the absorption capacity of the components indicates the increase in the reduction of iron.

Scavenging Activity of Nitric Oxide (NO•): NO scavenging activity of the essential oils was determined as previously described [17]. Briefly, 0.1mL of the essential oils (0–0.3mg/ml in DMSO) was incubated with 0.5mL of sodium nitrite (0.01mg/mL in 100mM sodium citrate pH 5) at 37 °C for 2h. After incubation, 0.5mL of Griess reagent was added, and the absorbance was read at 540nm using a spectrophotometer (Pharmacia, Uppsala, Sweden). The equation obtained the percentage of RNS scavenging:

$$\text{NO• Scavenging effect (\%)} = (A_0 - A_1)/A_0 \times 100$$

A_0 : The absorbance of the control

A_1 : The absorbance of the sample.

Cytotoxicity Activity

Cell Lines and Culture Conditions: In this study, cancerous cells were used: HeLa and MCF-7. HeLa is a transformed cell line expressing the HPV18 virus (human papillomavirus) [18]. MCF-7 (Michigan Cancer Foundation-7) was isolated from a 69 years old woman with metastatic disease [19]. HeLa and MCF-7 cell lines were supplied by ATCC (Manassas, VA, USA). All cells were grown in RPMI 1640 medium (Gibco) supplemented with 10 % (v/v) foetal calf serum (FCS) and 2mM L-glutamine in tissue culture flasks (Nunc). They are incubated at 37 °C, 95 % air and 5 % CO₂.

MTT: For the test of cytotoxicity activity, the MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) (Sigma-Aldrich, St. Louis, MO, USA) test was done [20]. First, HeLa or MCF-7 cells in 96-well plates were exposed to different concentrations of CPEo (3 wells for each concentration) and incubated for 48h at 37 °C. Then, 20µl of MTT was added to each well. After 4h of incubation at 37 °C, the supernatant was removed, 180µL of DMSO/Methanol (1V/1V) were added to each well to solubilize the formazan crystals. Finally, the plates were shaken 15min at room temperature, and the absorbance was detected at 570nm with a spectrophotometric plate reader.

Statistical Analysis

The statistical studies are carried out using the program SPSS

(19.0). The t-Student test carried out the comparison between the averages. The results are represented as mean ± standard deviation (SD).

Results

The Yield of Grapefruit Essential Oils

The yield of the extraction is defined as the ratio between the mass of essential oil obtained and the mass of the plant material.

$$Y (\%) = A/A_1 \times 100$$

A: Quantity of extracts recovered in g.

A₁: Quantity of dry vegetable matter used for extraction expressed in g.

CPEo yield was 1.32g Eo / 100g dry matter. This result proves that the zest of the fruit was rich in Eo.

Antioxidant Effects

DPPH Radical Scavenging Assay: The curve relating to DPPH radical scavenging shows that the inhibition percentage of the free radical increases with the increase of concentration, either for BHT or CPEo (Figure 1). In effect, Citrus paradisi essential oil has a significant antiradical effect attending 54 % and can be qualified such having an important antioxidant activity.

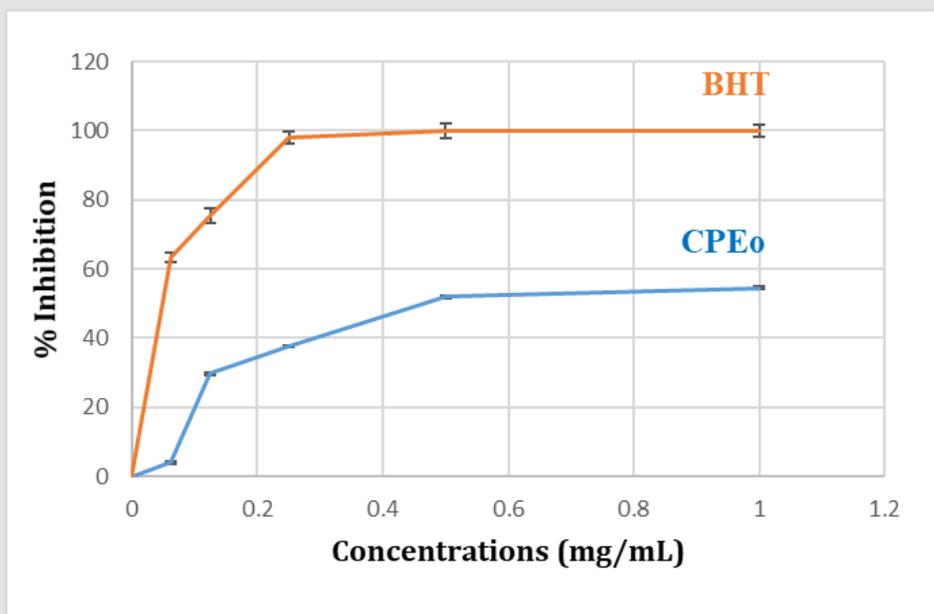


Figure 1: Antiradical activity against the radical DPPH in the percentage of inhibition CPEo. Aliquots of various concentrations of CPEo (0, 0.2, 0.4, 0.6, 0.8, 1mg/mL) and standard BHT were mixed with DPPH and incubated in the dark. The antioxidant activity of CPEo was measured using a spectrophotometer at 517nm. Results were expressed as mean inhibition percentage (%) ± standard deviations (n = 3). BHT was used as the reference standard.

ABTS+ Radical Scavenging Effect: The results illustrated in the curve show that the percentage of inhibition gradually increases

with the concentration of grapefruit essential oil. At a concentration of 1mg/mL, the inhibition rate reaches 83 % (Figure 2).

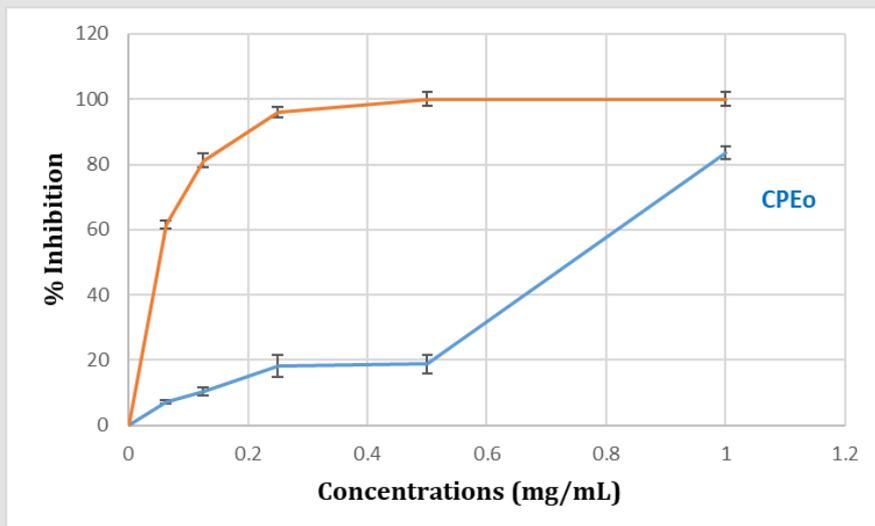


Figure 2: Anti-radical activity against the radical ABTS in percentage of inhibition of CPEo. Various concentrations of CPEo (0 to 1mg/mL) and acid ascorbic were mixed with FRAP reagents. The reduction of ferric ion (Fe^{3+}) to ferrous form (Fe^{2+}) by CPEo produces an intense blue light revealed as a change in absorption at 700nm. Results were expressed as mean inhibition percentage (%) \pm standard deviations (n = 3). Ascorbic acid at various concentrations (0 to 1mg/mL) was used as standard.

Ferric-Reducing Antioxidant Power (FRAP): The FRAP assay is usually used to measure the capacity of the sample to reduce the

ferric complex to the ferrous form. The FRAP assay confirmed the antioxidant activity of CPEo, with values equal to 66 % (Figure 3).

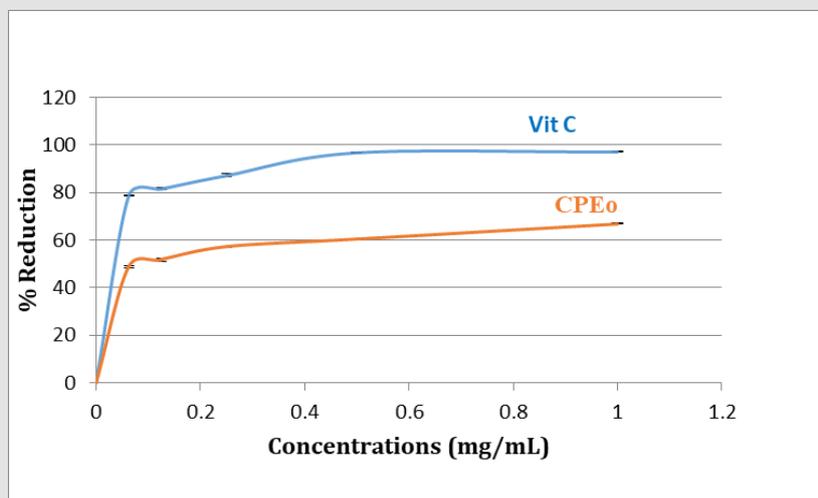


Figure 3: Inhibition (%) of standard Vit C and CPEo by Ferric Reducing Antioxidant Power (FRAP) assay. Various concentrations of CPEo (0 to 1mg/mL) and acid ascorbic were mixed with FRAP reagents. The reduction of ferric ion (Fe^{3+}) to ferrous form (Fe^{2+}) by CPEo produces an intense blue light revealed as a change in absorption at 700nm. Results were expressed as mean inhibition percentage (%) \pm standard deviations (n = 3). Ascorbic acid at various concentrations (0 to 1mg/mL) was used as standard.

NO• Scavenging Assay

The capacity of citrus paradisi essential oils to scavenge NO was also measured. CPEo was checked for its inhibitory effect on

nitric oxide production. The inhibition on free radical percentage increases with elevated concentrations of essential oil and Vitamin C (Figure 4).

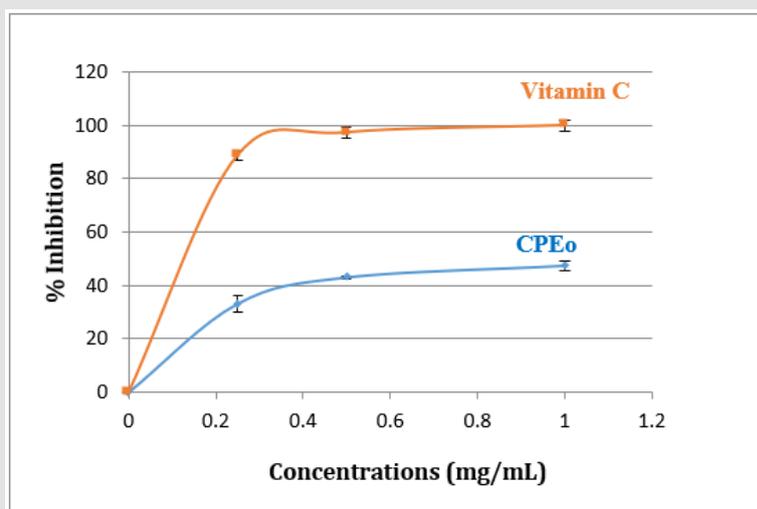


Figure 4: Inhibition (%) of standard Vit C and CPEo by Ferric Reducing Antioxidant Power (FRAP) assay. Various concentrations of CPEo (0 to 1mg/mL) and acid ascorbic were mixed with FRAP reagents. The reduction of ferric ion (Fe³⁺) to ferrous form (Fe²⁺) by CPEo produces an intense blue light revealed as a change in absorption at 700nm. Results were expressed as mean inhibition percentage (%) ± standard deviations (n = 3). Ascorbic acid at various concentrations (0 to 1mg/mL) was used as standard.

Cytotoxicity Activity

The MTT test studied the cytotoxic effect on HeLa and MCF-7 cells. Cells were cultured in 96-well plates for 48 h, in the presence and absence of the different concentrations of CPEo. The percentage

of cytotoxicity was calculated. Our results showed an antitumor potential of the grapefruit essential oils on both cancer cells: Hela and MCF-7 according to the concentrations used: from 0.78 to 25µg / mL (Figure 5).

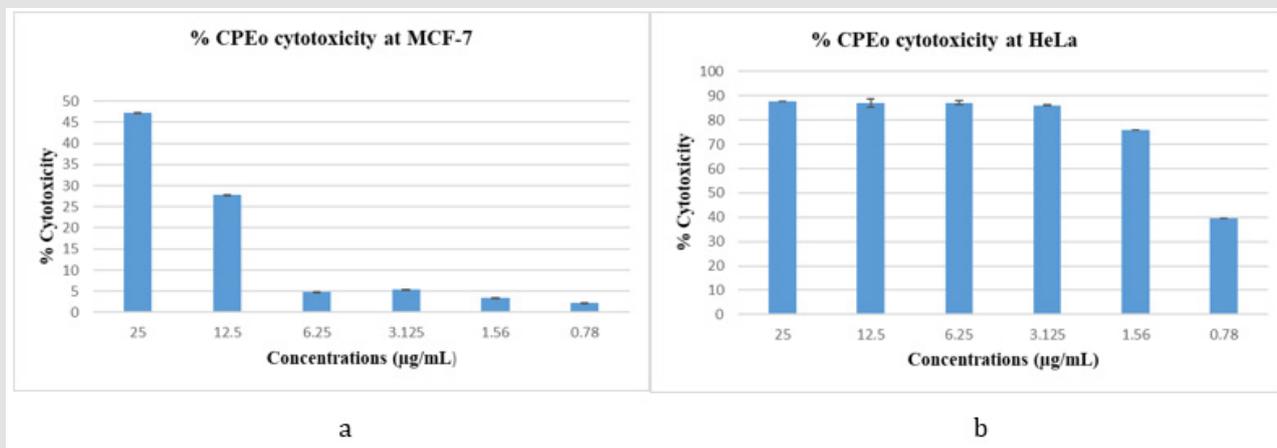


Figure 5: (a) Study the cytotoxicity of the essential oil of the grapefruit outer wall in the HeLa (b) and MCF-7 cell lines.

Cytotoxic activity of citrus paradisi essential oil on cancer cell lines. MTT assessed cell viability. The percent growth reduction was calculated from the extinction difference between treated cell culture and the control. Results are the means of three repetitions.

Discussion

The current study was designed to extract the essential oil from grapefruit. It was obtained by hydro distillation, which is the

oldest method and the most used because it is very easy to achieve. Our results agree with studies, which confirmed that Citrus is rich in essential oil, and their yields varied from one plant species to

another, ranging from 0.2 to 2.0 % [21]. Other work has shown that yield of Citrus paradisi essential oil collected in September in southern Taiwan was attending 0.37 %, and others were collected in October from Iran were 0.85 % [22,23]. So the difference in EO yields may be related to the plant's origin, the environmental conditions, time of collection, and extraction method [24]. The CPEo analyzed in this work showed potent radical scavenging activity. We have demonstrated that CPEo has an interesting antioxidant activity highlighted by the DPPH, ABTS+, FRAP, and NO. Our results agree with the literature, which showed that citrus EO possesses an anti-free radical activity DPPH of 17.7 to 64 % [7]. Other studies on grapefruit seed oil showed a high antioxidant potential (61 %) [25]. Furthermore, the effect of citrus paradisi essential oils on ABTS free radical was determined and exhibited a higher radical scavenging activity. Our results corroborate with other studies showing a significant antioxidant effect of C. paradisi EO against ABTS radicals [22]. In addition, the CPEo had a significant FRAP value, which was in accord with the results obtained by Jang et al. that confirmed the higher FRAP values in the peel of grapefruit [26]. The superoxide anion assay is commonly used to evaluate the superoxide anion radical-scavenging ability of plant extract. Therefore, CPEo was checked for their inhibitory effect on nitric oxide production in the present study, attending 47 %. Our results agree with studies that confirmed that the superoxide values of grapefruits varied from 36.83 % to 50.31 % in peels [27]. The antitumor potential of CPEo was tested on HeLa and MCF-7 tumor cells. Our results showed a considerable antitumor activity dependent on essential oil concentration. This result agrees with the work of Zu et al., which shows that essential oil from grapefruit exhibited an anti-proliferative effect on MCF-7 cells [28]. Furthermore, Monajemi et al. studied the impact of different concentrations of essential oils of other species of the citrus family as Citrus limon, Citrus medica, and Citrus sinensis on MCF-7 and Hela [29]. They proved a significant decrease in viability in a dose-dependent manner in both tumor cells [29].

Conclusion

Citrus paradisi essential oils are well known for their flavor and fragrance properties and numerous aromatherapeutic and medicinal applications. Accordingly, the essential oils of Citrus species exhibited intense antioxidant activity. In addition, the essential oil of Citrus paradisi showed an antitumor effect against both cancers cells, HeLa and MCF-7.

Acknowledgment

This work is dedicated to the memory of Pr. Saloua Lassoued Elamri. We will never forget her, and she is always in our hearts.

Conflict of Interest

The authors declare that they have no competing interests.

References

1. Fabricant DS, Farnsworth NR (2001) The value of plants used in traditional medicine for drug discovery. *Environ Health Perspect* 109(1): 69-75.
2. Ben Amor I, Musarra Pizzo M, Smeriglio A, D'Arrigo M, Pennisi R, et al. (2021) Phytochemical Characterization of Olea europaea Leaf Extracts and Assessment of Their Antimicrobial and Anti-HSV-1 Activity. *Viruses* 13(6): 1085.
3. Gargouri B, Mansour RB, Abdallah FB, Abdelfetteh Elfekih, Saloua Lassoued, et al. (2011) Protective effect of quercetin against oxidative stress caused by dimethoate in human peripheral blood lymphocytes. *Lipids Health Dis* 10: 149.
4. Sewell RDE, Rafieian Kopaei M (2014) The history and ups and downs of herbal medicine usage. *J HerbMed Pharmacol* 3(1): 1-3.
5. Edris AE (2007) Pharmaceutical and therapeutic potentials of essential oils and their individual volatile constituents: A review *Phytother Res* 21(4): 308-323.
6. Martinez Velazquez M, Rosario Cruz R, Castillo Herrera G, J M Flores-Fernandez, A H Alvarez, et al. (2011) Acaricidal effect of essential oils from *Lippia graveolens* (Lamiales: Verbenaceae), *Rosmarinus officinalis* (Lamiales: Lamiaceae), and *Allium sativum* (Liliales: Liliaceae) against *Rhipicephalus* (Boophilus) microplus (Acari: Ixodidae). *J Med Entomol* 48(4): 822-827.
7. Choi H S, Song H S, Ukeda H, M Sawamura (2000) Radical-scavenging activities of citrus essential oils and their components: detection using 1, 1-Diphenyl-2-picrylhydrazyl. *J Agric Food Chem* 48(9): 4156-4161.
8. Zenasni L (2014) Etude de polymorphisme chimique des huiles essentielles de Thymus satureioides Coss et d'Origanum compactum Benth et du genre Nepeta et évaluation de leur propriété antibactérienne. Thèse de doctorat. Université Mohammed VAgdal Rabat, pp.155.
9. Graziano AC, Cardile V, Crasci L, Sivia Caggia, Paola Dugo, et al. (2012) Protective effects of an extract from Citrus bergamia against inflammatory injury in interferon- γ and histamine exposed human keratinocytes. *Life Sci* 90(25-26): 968-974.
10. Singh J, Sood S, Muthuraman A (2014) *In-vitro* evaluation of bioactive compounds, antioxidant, lipid peroxidation and lipooxygenase inhibitory potential of Citrus karna L. peel extract. *J Food Sci Technol* 51(1): 67-74.
11. Zhou Z, Xi W, Hu Y, Chao Nie, Zhiqin Zhou (2016) Antioxidant activity of Citrus fruits. *Food Chem* 196: 885-896.
12. Tepe B, Daferera D, Sokmen A, Münevver Sökmen, Moschos Polissiou. (2015) Antimicrobial and antioxidant activities of the essential oil and various extracts of Salvia tomentosa Miller (*Lamiaceae*). *Food Chemistry* 90(3): 333-340.
13. Viuda MM, Ruiz NY, Fernández LJ (2008) Antibacterial activity of different essential oils obtained from spices widely used in Mediterranean diet. *International Journal of Food Science & Technology* 43: 526-531.
14. Boughariou E, Allouche N, Jmal I, Mokadem N, Ayed B, et al. (2018) Modeling aquifer behaviour under climate change and high consumption: Case study of the Sfax region, southeast Tunisia. *J Afr Earth Sci* 141: 118-129.
15. Re R, Pellegrini N, Proteggente A, A Pannala, M Yang, et al. (1999) Antioxidant activity applying an improved ABTS radical cation decolorization assay. *Free Radic Biol Med* 26(9-10): 1231-1237.
16. Karagözler A, Erdağ B, Emek Y, Deniz Aktaş Uygun (2008) Antioxidant activity and proline content of leaf extracts from Dorystoechas hastate. *Food Chemistry* 111(2): 400-407.
17. Kavooosi G, Purfard AM, Aram F (2012) Radical scavenging properties of essential oils from Zataria multiflora and Ferula asafetida. *Asian Pacific Journal of Tropical Biomedicine* 2(3): S1351-S1356.

18. Hatfull G, Bankier BG, Barrell P, PJ Farrell (1988) Sequence analysis of Raji Epstein-Barr virus DNA. *Virology* 164(2): 334-340.
19. Gargouri B, Nasr R, Mseddi M, Riadh benmansour, Saloua Lassoued (2011) Induction of epstein-barr virus (EBV) lytic cycle *in vitro* causes lipid peroxidation, protein oxidation and DNA damage in lymphoblastoid B cell lines. *Lipids in Health and Disease* 10: 1-8.
20. Mosmann T (1983) Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays. *J Immunol Methods* 65(1-2): 55-63.
21. Tu NT, Onishi Y, Choi HS, Yusuke Kondo, Solomon Mitiku Bassore, et al. (2002) Characteristic odor components of Citrus sphaerocarpa Tanaka (Kabosu) cold-pressed peel oil. *Journal of Agricultural Food Chemistry* 50(10): 2908-2913.
22. Ming Chiu OU, Yi Hsin Liu, Yung Wei Sun, Chin Feng Chan (2015) The Composition, antioxidant and Antibacterial Activities of Cold-Pressed and Distilled Essential Oils of Citrus paradisi and Citrus grandis (L) Osbeck. *Evidence-Based Complementary and Alternative Medicine* 80: 4091-4099.
23. Alireza SD, Mohammad MS, Hassan Vatandoost, Mohammad Reza Abai (2016) Chemical Compositions of the Peel Essential Oil of Citrus aurantium and Its Natural Larvicidal Activity against the Malaria Vector Anopheles stephensi (Diptera: Culicidae) in Comparison with Citrus paradisi. *Journal of Arthropod-Borne Diseases* 10(4): 577-585.
24. Hassiotis C, Lazari D, Vlachonassios K (2010) The Effects of Habitat Type and Diurnal Harvest on Essential Oil Yield and Composition of Lavandula Angustifolia Mill. In *Fresenius Environmental Bulletin* 19(8): 1491-1498.
25. Atolani O, Omere J, Otuechere CA, Adewale Adewuyi (2012) Antioxidant and cytotoxicity effects of seed oils from edible fruits. *Journal of Acute Disease* 1(2): 130-134.
26. Jang HD, Chang KS, Chang TC, Hsu Chuan Liang (2010) Antioxidant potentials of buntan pumelo (Citrus grandis Osbeck) and its ethanolic and acetified fermentation products. *Food Chemistry* 111(3): 554-558.
27. Xi W, Fang B, Zhao Q, Bining Jiao, Zhiqin Zhou (2014) Flavonoid composition and antioxidant activities of Chinese local pummelo (Citrus grandis Osbeck.) varieties. *Food Chemistry* 161: 230-238.
28. Zu Y, Yu H, Liang L, Yujie Fu, Thomas Efferth, et al. (2010) Activities of ten essential oils towards Propionibacterium acnes and PC-3, A-549 and MCF-7 cancer cells. *Molecules* 15(5): 3200-3210.
29. Monajemi R, Oryan S, Haeri Roohani A, Alireza Ghannadid, Abbas Jafarian (2005) Cytotoxic Effects of Essential Oils of Some Iranian Citrus Peels. In *Iranian Journal of Pharmaceutical Research* 3: 183-187.

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