

# Impact of Intrauterine Exposure on Fetal Brain Development and Brain Injury

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#### ABSTRACT

Intrauterine exposures to environmental factors influence fetal brain development. During early development of fetal brain, billions of cells differentiate into neurons and form connections. Synaptic activity affects the strength and number of synapses that form between neurons. The intrauterine environment plays an essential role in the mechanisms of fetal brain development and injury. Various intrauterine insults that impact the process of brain development, including genetic, traumatic, infectious, maternal stress, and environmental etiologies, can result in abnormal development or neurological injuries. Perinatal brain injury can cause lifetime neurologic disability. Understanding of the relationship between the intrauterine environment and fetal brain development remains limited and is needed to shed light on effective strategies to predict and prevent the risk of brain injury during intrauterine fetal development. This review focuses on intrauterine exposure to various environmental factors, their impact on brain development, and resultant brain injury among premature and term infants.

## Introduction

Human prenatal brain development after fertilization is usually classified into four periods: 0-7 gestational weeks (GW) and neuronal proliferation during 8–15 GW, 16–25 GW, and > 26 GW [1]. In humans, neurons are mostly produced in the first trimester of gestation. The rapid development of the fetus's cerebral cortex from the day of fertilization occurs for a period lasting from 8 to 15 weeks, and by 16 weeks, the number of neurons in the cerebral cortex reaches the adult level [2,3]. Abnormal brain growth may result from an unsuitable intrauterine environment. Adverse intrauterine environments that may have a negative effect on the fetal brain include maternal diabetes, undernutrition, infection, hypoxia, stress, alcohol, smoking, toxins, and anemia; hypertensive disorders in pregnancy; high-altitude pregnancies; and placental insufficiency. These adverse environmental factors may trigger epigenetic alteration and have a significant impact on fetal brain development through genome-wide changes of epigenetic regulation. The common epigenetic modifications include acetylation of histone and methylation of DNA, in addition to non-coding RNA epigenetic regulations [4,5] and chromatin modification [6,7], which are vulnerable to the maternal environment [8]. The purpose of this review is to summarize articles on the deleterious effects of some types of intrauterine exposure on fetal brain development and brain injury. The hope is to provide the impetus for further studies to delineate the function of the intrauterine environment on fetal brain development, through evidence from premature and term infants, as well as the role of the intrauterine environment in lifelong brain injuries and the pathologic mechanisms by which these injuries occur.

## Fetal Brain Development

**Synapse Development:** Synapses connect billions of neurons during intrauterine fetal brain development, which is important in all functional neuronal circuits [9]. Synaptic plasticity is characterized by the removal and insertion of amino-3-hydroxy-5-methy1-4-isoxazolepropinic receptors (AMPARs) into the

postsynaptic membrane, and by the shrinkage or enlargement of dendritic spines, where the majority of excitatory synapses are positioned [9,10]. Synapse formation exceeds elimination, leading to a surplus of immature excitatory synapses during early brain development. Subsequently, synapse elimination and destabilization diminish the number of synapses, thus refining neural circuits that generate cognition and behavior [11]. Cell surface receptors such as metabotropic glutamate receptors (mGluRs), NMDA-type glutamate receptors (NMDARs), and tyrosine kinase (TRK) receptors activate mTOR signaling through the AKT pathway and the phosphoinositide-3 (PI3K) pathway, and MAPK via the ERK pathway. The ERK/MAPK pathway plays a key role in synaptic plasticity, consolidation of memory, and the transition from pluripotent stem cells to neuronal progenitors [12]. The myocyte enhancer factor (MEF2) family of transcription factors can regulate synapse elimination during brain development [13,14]. Synaptic strength is mainly influenced by changes in synaptic structure that depend on instruction of local protein synthesis, structural remodeling of the cytoskeleton, and receptor signaling [15,16]. Glutamate serves as both as a key neuromodulator to control synape and cirruit function and the mammalian brain's primary excitatory neurotransmitter over a wide range of temporal scales and spatial. The group metabotropic GluRs (mGluRs) are abundant at excitatory synapses throughout the brain, where they are speculated sited to adjust to glutamatergic signaling [17]. They are vital to synaptogenesis and the shape of neural circuitry during the period of brain development [17]. Some evidence has demonstrated the important function of signaling lipids in mediating signal transduction and membrane traffic at pre-and post-synapses. For example, phosphoinositides can conduct ion channels, regulate exocytosis and endocytosis of synaptic vesicles and postsynaptic receptors, and signal from activated neuroreceptors such as NMDARs and mGluRs to allow plastic adjusted function of synapse [18,19].

Oligodendroglial Cells: Oligodendroglial cells in the central nervous system (CNS) synthesize myelin, transform from progenitor to the mature oligodendrocyte, and play a key role in salutatory conduction of action potentials [20-22]. After 20 GW, oligodendrocyte progenitor cells (OPCs) are shaped in the ventricular zone [23]. OPCs are generated in the brain and spinal cord from multipotent stem cells, and then they proliferate and differentiate. Neurogenesis and oligodendrogliogenesis progress at different rates in the human brain. OPCs first emerge in the ganglionic eminence at approximately 9 GW in pregnant women [24,25]. In humans, cortical oligodendrogenesis begins at around 10 GW, but it progresses well into adulthood [26]. Olig2-positive stem cells from early fetal development exist in the germinal matrix of the brain and transfer from the original regions in the brain to the axon-dense zones of the neocortex, spinal cord, diencephalon, and brainstem.

Gliogenesis: Gliogenesis is often generated during the last trimester of gestation in humans [27]. As mentioned above, the timing of an insult in pregnancy is critical to compare and estimate the neurodevelopmental response of offspring. While early insult in pregnancy is related to structural brain abnormalities such as neural tube defects, late-gestation insults may disturb the migration progression of postmitotic neurons and cause deviant cortical development [28]. Later insults have been demonstrated to be associated with more with behavioral, cognitive, and psychiatric disorders, such as autism, obsessive compulsive disorders, and schizophrenia [29,30]. The regulation of oligodendrocyte differentiation and myelination in the fetal brain involves negative and positive regulators [23]. There are three negative regulatory pathways for oligodendrocyte differentiation, including the BMP signaling, Notch signaling, and Wnt/ $\beta$ -catenin pathways. These and Wnt pathways are involved in oligodendrocyte maturation. Some studies showed that white matter disorders are associated with dysregulation of the BMP and  $Wnt/\beta$ -catenin signaling pathways [31,32]. The maturation of oligodendrocytes relies on ATP through oxidative phosphorylation in mitochondria [33]. Mitochondria support oligodendrocyte differentiation and survival [34]. It is commonly found that after hypoxia-ischemia, mitochondrial dysfunction occurs in the developing brain [35]. Studies have demonstrated that microglia have an effect on the maturation of oligodendrocytes during normal brain development. Activated microglia discharge high levels of pro-inflammatory cytokines, including interleukin (IL)-1β, IL-2, and IL-17; tumor necrosis factor-alpha (TNFα); and excitotoxic factors such as glutamate, nitric oxide, and hyaluronan, or endothelial growth factor, which impair immature oligodendrocyte differentiation and proliferation and assist in decreasing the number of oligodendrocytes [36-39]. Mitochondria are very important in the developing brain and throughout life in energy phosphate tasks such as regulating the cellular redox state, maintaining organelle function, cellular proliferation, mediating the DNA or protein responsible for transcription, excitotoxic injury, translation and assembly of the enzyme complexes of the respiratory chain, and apoptosis. These mitochondrial functions in cellular proliferation rely on mitochondrial dynamics. Mitochondria are extremely plastic and mobile, altering their shape through fission and fusion to reach sites of high energy demand in cells [40]. Mitochondrial impairment results in deregulation of calcium homeostasis, bioenergetic failure, mitochondrial permeabilization with release of proapoptotic proteins, and production of reactive oxygen species (ROS), leading to cell death [41].

**Myelination:** The myelination of mature oligodendrocytes continues especially in late gestation and is susceptible to excitotoxic and damage associated with premature exposure to the extrauterine environment without neuroprotection. Neonatal or fetal brain injury may occur as a result of thrombosis, infection,

hemorrhage, trauma, or hypoxia and can lead to lifelong cognitive, sensory, or motor dysfunction. Defining the type and range of brain damage is not as simple as it would seem. Magnetic resonance imaging (which requires dangerous ionizing radiation) and transcranial ultrasound (which has limited sensitivity) can only be used to examine the damaged fetal brain, not to predict function.

Placenta and Fetal Brain Development: The placenta is the maternal-fetal interface that has an essential role in the transfer of nutrients and oxygen to the fetus and provides and secretes growth-regulating factors to ensure the neurodevelopment of fetal brain. In addition, the placenta functions as an immuno-defender to protect the fetus from maternal infection and inflammation. The placenta, which controls the intrauterine environment, is of fetomaternal organ. It is well-known to secrete neurotransmitters, which are associated with abnormal neurodevelopment and normal fetal brain development. The maternal component of the placenta is the decidua. The fetal placental tissues include the umbilical cord, chorionic villi, amniotic membrane, and chorionic membrane [42,43]. Placental metabolism, placental hormone production, and substrate transport are all essential for fetal development. Normal development of the placenta includes two concurrent and complex processes: the cytotrophoblast (CT) cells invade the endothelium of the maternal spiral artery and then the fetal vascular tree develops. Endothelial cell invasion initially leads to the formation of a trophoblast "plug," resulting in a hypoxic milieu environment within the intervillous space (oxygen partial pressure [PaO,] < 20 mm Hg) [44]. After 10 GW, the CT plug dissipates, which results in increased placental blood flow and PaO<sub>2</sub> [45,46]. Several mechanisms affect placental function. The sustained high-pressure flow through the intervillous space (2-3 m/s, while normal dilated vessels are 10 cm /s) leads to increased shear stress and damage to the trophoblast cells of the chorionic villus, thereby damaging the capacity of the villi for nutrient and gas exchange [46,47]. Unsuccessful spiral arterial conversion makes these vessels prone to adrenergic stimulation and vasoconstriction, which leads to intervillous PaO, fluctuations and placental hypoxic perfusion injury [48]. Dysregulation of angiogenesis and anti-angiogenic factors in the placental interface results in abnormal development of the fetal vascular tree in the placenta [49], subsequently impairing the function of the placenta, and has been relevant in the development of preeclampsia, gestational diabetes-related pregnancy, fetal growth restriction (FGR), placenta early exfoliation, intrapartum fetal compromise (IFC), and preterm birth [50-52].

Risk factors of arterial disruption involve trauma, preeclamptic arteriopathy, uterine rupture, abruption placenta, and vasoactive drugs, such as nicotine or cocaine. Marginal retroplacental hemorrhages mostly occur at the margin of venous drainage of the placenta [53,54]. Many other events, such as malnutrition, genetic abnormalities, and infection can also disturb placental function and alter the fetal brain's environment. The failure function of placenta can directly injure the developing brain or raise its vulnerability to injury, result in lasting neurological disabilities [55,56].

#### **Fetal Brain Injury**

Intrauterine Fetal Brain Injury: Chorioamnionitis, hypoxia, fetal inflammatory response, and preterm birth can contribute to brain injury and progression of the subsequent neurological deficits [57]. Hypoxia and inflammation mediate neuropathology, acting to induce neuroinflammation and breakdown of the blood brain barrier (BBB), resulting in oligodendrocyte cell damage [58]. The activated immune cells such as mast cells, microglia, and neutrophils release the key cytokines IL-1 $\alpha$ , IL-1 $\beta$ , IL-6, and IL-18, which sequentially stimulate the discharge of TNF, ROS, and excitatory amino acid agonists, including glutamate, which work together to initiate neural cell apoptosis [59,60]. The substances above can also directly influence the differentiation of neurons and OPCs by inducing apoptosis and can cause mitochondrial failure [61,62]. Neighboring reactive astrocytes, by releasing TNF- $\alpha$ and IL-1β, cause proliferative inhibition of oligodendrocytes and downstream activation of apoptotic pathways [63].

Impact of Intrauterine Infection: Many microorganisms, which include certain viruses, bacteria, and protozoa, have been linked to intrauterine infection. These infections can result in clinical syndromes, including TORCH infections, referring to infections caused by toxoplasma, other microorganisms, rubella virus, cytomegalovirus (CMV), and herpes simplex viruses (HSV) [64]. Other common infections in women are caused by aerobes, such as group B streptococcus (GBS) (15%); and gram-negative rods, including Escherichia coli (8%), anaerobes, including Bacteroides sp. (30%) and Gardnerella vaginalis (25%) [65]. These microorganisms are associated with preterm birth [66]. Further studies demonstrated that a persistent intrauterine inflammatory exposure may also result in fetal brain injury [67]. The characterization of chorioamnionitis is an intra-amniotic infection in which bacteria invaded the amniotic cavity, resulting in acute inflammation of the fetal membranes and/or the placenta [65]. Chorioamnionitis, which results in spontaneous preterm birth and premature rupture of membranes (PROM), is defined as a feto-placental environment of acute inflammation [68]. Chorioamnionitis often leads to fetal inflammation and damage to the immature brain, raising the possibility of diffuse white matter injury and intraventricular hemorrhage [69]. Fetal inflammatory response syndrome (FIRS), resulting from systemic immune activation, is characterized as inflammation of multiple fetal organs in utero [70]. Infections can trigger inflammatory pathways, causing the discharge of diverse proinflammatory biomarkers, such as interleukins, cytokines, and other molecules. Proinflammatory cytokines such as IL-1β, IL-6, and tumor necrosis factor-alpha (TNF- $\alpha$ ) from microglia and astrocytes may directly insult neurons and oligodendrocytes. The injection of IL-1 $\beta$  in neonatal rats leads to delayed myelination and neuronal death [71]. TNF- $\alpha$  induces apoptosis in developing oligodendrocytes and cell death in mature oligodendrocytes [72].

Toxoplasma: Primary infection of congenital toxoplasmosis in pregnant women is uncommon, but it remains as a latent, chronic, cryptic brain infection throughout the life of the host, often with severe consequences [73]. Some studies have demonstrated that infection of the toxoplasma may alter cognitive functions and human behavior and may cause headaches, the onset of schizophrenia, and cryptogenic epilepsy [74]. Chronic, adultacquired Toxoplasma gondii infection in outbred mice can cause behavioral and neurologic abnormalities secondary to the loss of brain parenchyma and inflammation [74]. There is also improved expression of message for synaptic remodeling and markers of neuronal cell death and mediators of inflammation in brains of chronically infected mice, comparing to uninfected control mice [74]. Synaptic transmission underlies vastly complicated instruction by protein networks assembled at the presynaptic location of neurotransmitter discharge and the postsynaptic device for neurotransmitter reception. Haroon and colleagues have indentified that T. gondii--activated cytokines disturb synaptic signaling [75]. Chronic T. gondii infection brings dissimilar changes in synaptic protein composition, which downregulate a huge number of proteins occupied in synaptic plasticity and pose a danger for neuropsychiatric disorders [75].

**Rubella:** During the Rubella epidemic in the U.S, in 1964-1965, thousands of infants were infected and subsequently suffered lifelong problems. During this epidemic, 8–13% of cases were congenital rubella syndrome (CRS) occurring in early pregnancy. Rubella virus has a particular affinity for the central nervous system, resulting in mental retardation, encephalitis, cataracts, central auditory imperceptions, glaucoma, and cochlear atrophy [76]. Fetal damage associated with rubella is inclined to occur only when an infection occurs in the first 16 weeks of pregnancy. In general, the earlier the infection begins, the more severe the malformation that is observed [77]. The later sequelae of rubella in early pregnancy include diabetes and autism [78-81].

**Cytomegalovirus:** Human cytomegalovirus (HCMV) infection is the main cause of congenital viral infection and brain disease in developed countries. HCMV is also a major pathogen in congenital illness and can lead to permanent disabilities, including hearing loss, mental retardation, and vision loss. It was reported that 50% of children with congenital HCMV infection in Japan developed hearing loss 6 months after their diagnoses [82]. HCMV in fibroblasts acquires its covering by budding into exosome-like vesicles, which subsequently combine with the plasma membrane to discharge mature virions from the cell. Compared to the infected cells, the glycerophospholipid component of HCMV virions is strikingly different. The liposome of virions has been found to be similar to that of synaptic vesicles via comparing Monica the published results [83]. These similarities showed that HCMV in fibroblasts obtains its envelope by budding into vesicles, which can fuse at the plasma membrane to discharge mature versions of this cell. Synaptosomal-associated protein of 25 kDa (SNAP-25), a constituent of the SNARE complex, which mediates exocytosis of synaptic vesicles in exocrine cells and neurons, has been found to be involved in the exit of HSV-1 from neurons [83,84].

Herpes Simplex Viruses: The rate of neonatal herpes simplex virus (HSV) infection ranges from 1 in 2,500 to 1 in 20,000 live births. Manifestations of congenital HSV include hydranencephaly, chorioretinitis, skin lesions, scars, and microcephaly. The condition of neonates who have HSV infection can deteriorate rapidly due to encephalitis, disseminated intravascular coagulopathy, shock, or respiratory distress. Infants who survive neonatal HSV encephalitis have high rates of neurological sequelae, including mental retardation, visual or motor deficits, Alzheimer's disease (AD), and seizure disorders [85]. HSV-1-infected neurons also have shown considerably reduced expression of the presynaptic proteins synaptophysin and syanpsin-1 and depressed synaptic transmission. In mice, these effects rely on intraneuronal accumulation of A $\beta$  and GSK-3 activation [86].

Bacterial Infection: Group B Streptococcus and E. Coli: GBS is the leading reason for congenital bacterial infection in developed nations. The incidence of transmission to newborns in GBS-positive women is about 21% [87]. No direct evidence has shown that GBS infection plays a function in cerebral palsy (CP), but nearly 50% of infants who survive GBS meningitis experience longterm neurodevelopmental sequelae [88]. In addition, mediation of extensive cortical neuronal injury through reactive oxygen intermediates was observed in GBS-infected neonatal rats [89]. The association of cellular response of the fetal brain with perinatal inflammatory or infectious damage reflects activation of astrocytes and microglia with oligodendrocyte dysfunction and neuronal loss. In developing countries, E. coli is one of the major pathogens leading to early-onset infections in preterm neonates. In human newborn infants, cerebral white matter injury has been observed by MRI following an episode of E. coli meningitis [90].

#### Hypoxic-Ischemic Injury

Unpredictable and severe events that involve placental abruption, cord prolapse, uterine rupture, or eclampsia are strongly associated with a high risk of catastrophic fetal hypoxia [91]. Hypoxic-ischemic injury that leads to mental retardation, motor impairment (CP), hypoxic ischemic encephalopathy (HIE), and seizures is a considerable contributor to morbidity and

mortality in infants [92]. The fetal brain of prematurity before 32 GW is immature, and the white matter is especially susceptible [93]. The susceptibility of the immature CNS to hypoxia-ischemia is mainly dependent on regional status and the timing of decisive developmental processes, for example, proliferation, differentiation, migration, programmed cell death, and myelination, as well as on the instruction of metabolism and cerebral blood flow. The fetal brain is hypersensitive to hypoxic damage and oxidative stress because of its high oxygen consumption, lack of glucose stores, high lipid content [94,95], and considerably low concentrations and activity of antioxidant enzymes [96,97]. The upregulation of IL-1 and TNF-R1 can result from periods of hypoxia in the brain. Pro-inflammatory cytokines mediate the immune response to inflammation and infection and influence a wide range of physiological action that involves cell survival, fever, acute-phase response gene expression, glial activation, hypotension, T- and B-lymphocyte stimulation, and leukopenia [98-101]. Hypoxia, which damages OPCs by activating the enzymes caspase-3 and caspase-9, leading to cell death, is also related to calcium influx after inflammation-induced glutamate discharge from immune cells, which causes excitotoxicity and results in bax translocation to the mitochondria on OPCs and release of cytochrome-c [102]. The exact mechanisms underlying hypoxic cerebral damage are multifarious and are not totally mediated by the initial hypoxic injury, but instead are compounded by insults happening during the reperfusion stage [103] because of toxicity from ROS and activation of N-methyl-D-aspartate-type glutamate receptors [104]. In fact, the severity of the secondary injury happening during the reperfusion phase associates best with the severity of neurodevelopmental disability at 1 and 4 years of age [105]. There is a strong connection between hypoxia and hypotension with fetal injury, mainly fetal death and neuronal damage. During hypoxia, the blood flow of cerebral hemispheres is reduced, whereas perfusion to the thalamus, brainstem, and basal ganglia is increased [106]. Cerebral ischemia has a dramatic and rapid effect on synaptic function and structure [107].

#### Seizures

Seizures are one of the most common neurological emergencies in newborns. A reduction in the normal environment of fetal neurosteroids is associated with undesirable outcomes, such as episodes of potentially destructive seizures, which can cause destructive and permanent conversion in neurodevelopment [108,109]. Premature birth is related to an increased rate of seizure disorders [110]. Neonatal seizures create a long-term increase in seizure susceptibility and change in inhibition/excitation balance of synaptic transmission in layer II/III neurons of the somatosensory cortex [111]. In summary, neonatal seizures have enduring effects on synaptic plasticity in the somatosensory cortex [112].

#### **Cerebral Palsy**

CP is defined as a disorder of posture and movement that includes abnormalities in reflexes, tone, movement, and coordination, and delays or aberration in primitive reflexes and in motor milestone achievement that is enduring, and is caused by a lesion, nonprogressive interference, immature brain, or abnormality of the developing fetal and infant brain [113,114]. CP is also defined by type (dyskinetic, dystonic, or spastic), topography (limb involvement), and descriptors of the extent and pattern of involvement (quadriplegia, hemiplegia, diplegia, and monoplegia) [114]. Autopsy of the brain of a preterm child with CP showed white matter atrophy, dysmyelination, ventriculomegaly, and reactive gliosis [115].

#### **Autism Spectrum Disorders**

Autism spectrum disorders (ASDs) are characterized by a complex and strong genetic component with broad familial inheritance patterns and have been found to be related to mutations in as many as 1,000 genes [116]. Environmental factors, including maternal diabetes, prenatal infections, prenatal and perinatal stress, zinc deficiency, and toxins, can also contribute to the risk of autism during early life [117,118]. Some evidence shows that the placenta plays a key role in ASD pathogenesis [119]. That the architecture of placenta from ASD patients consists of smaller branch angles than in population-based counterparts, fewer branch points, better extension to the surface boundary, and thicker and less tortuous arteries may indicate that both environmental and genetic factors have an impact on vascular branching morphogenesis in pregnant women [120,121]. A recent study of the placenta from patients with ASD demonstrated considerably higher incidences of fetal inflammation, maternal vascular mal-perfusion, and acute generalized inflammation, suggesting that these conditions are deleterious to fetal brain development [119]. Some forms of intellectual disabilities and syndromic autism are linked to mutations in genes that regulate protein synthesis and influence transmission, plasticity, and structure of synapses [12]. Failures to sustain RNA-binding protein levels and the accurate number of mRNA molecules are critical access points of synaptopathies [122].

#### Schizophrenia

Schizophrenia is a greatly polygenic disorder, involving hundreds of genes. Genes implicated in synaptic plasticity and glutamatergic function figure prominently among genes associated with schizophrenia [123]. A deficit in glutamatergic synapses can provoke schizophrenia [124]. Both neurochemical alterations and structural changes may lead to defective neuronal transmission in schizophrenia [123]. The uterine environment may have a significant influence on later development of schizophrenia [125-127]. Influenza infection during early gestation that has a strong correlation with schizophrenia in offspring can result in overexpression of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and IL-6, probably through changing either epigenetic modification or gene expression [128]. Schizophrenia has been associated with exposure that may happen in early life and might be linked to pregnancy (hyperglycemic conditions, hemorrhage, or preeclampsia), labor (uterine rupture, birth asphyxia), or fetal conditions (genetic anomalies or intrauterine growth restriction) [56,129].

#### **Stress Or Mood Disorders**

Many maternal stressors such as trauma, depression, and malnutrition can have an effect on the placenta and change maternal glucocorticoid levels, which play a major role in programmed cell death and neuronal maturation and to which the developing brain is extremely susceptible [130]. 11 $\beta$ HSD2 is expressed at very high levels in the placenta, which protects the fetus from the normal increase in maternal cortisol occurring during gestation. Maternal mood disorders have a relationship with disruption of the placental barrier, in part through suppressing 11 $\beta$ HSD2 expression, resulting in abnormal neurodevelopment in the offspring [131]. Human placental lactogen expression was considerably decreased in placentas from women diagnosed with depression and who had high depression scores [131].

#### **Premature Birth and Fetal Brain Development**

Preterm And Fetal Brain Development: Steroid precursors generate from the placenta that maintain the neuroprotective and trophic functions of neuroactive steroids in the fetal brain [132]. In preterm birth, the loss of neuroactive steroid precursors leads to disruption of the normal track of fetal brain development and delays the progress of myelination [133]. The GABAA receptors that play a key role in late gestation are vital to interaction with the placenta-derived neuroactive steroids [134,135]. Research studies showed that tobacco smoke during pregnancy may result in chronic hypoxia and be associated with increased placenta resistance and carboxyhemoglobin and decreased uterine blood flow [136]. Some scientists have also found the connection between elevated levels of serotonin and altered oligodendrocyte development and myelination [137]. Recent studies showed that extracellular vehicles (EVs) including proteins, nucleic acids, and lipids are a mechanism for communication between fetus and mother [138]. How EVs influence the maternal response to pregnancy and fetal development is currently an area of vigorous exploration [139,140].

**Premature Birth and Neurodevelopmental Disorders:** Preterm birth can result from maternal/fetal inflammatory responses and intrauterine infection and result in fetal brain damage with a negative effect on the function and structure of the entire brain [141]. Serial MRI examinations have shown that the gray and white matter volume of premature infants is reduced compared to full-term control groups [142,143]. Loss of neurons as a result of apoptosis may partly explain the reduction in gray matter volume of the basal ganglia and cortex in both humans and mice. This loss of neurons is the most common form of cerebral abnormalities in premature infants, which include hippocampus and gray matter abnormalities and diffuse white matter injury [144,145]. However, focal necrotic lesions of cystic ventricular leukocyte softening are seldom seen in preterm infants [146]. Prematurity can also result in CP and visual and hearing impairments [147]. The common forms of white matter injuries in preterm birth occur as diffuse white matter injury, periventriclular leukomalacia (PVL), and germinal matrix hemorrhage [148]. The less frequent forms of injury are cerebral sinus vein thrombosis, primary intraparenchymal hemorrhage, hyperbilirubinemiainduced kernicterus, and infectious meningitis/encephalitis [149]. PVL lesions have been demonstrated that have a relationship with the loss of pre-oligodendrocytes and OPCs [150].

Prospective: Extracellular vesicles provide a promising strategy for early prediction of intrauterine brain development EVs, including microvesicles and exosomes, participate in signal transmission between neurons, play a fundamental role in activity of the nervous system, and facilitate communication of the CNS with all body systems [151]. EVs may be produced in almost all cells of the body, function to transport biologically active molecules to target cells, and provide intercellular communications [152,153]. EVs are secreted by numerous cell types in the brain, including microglia, astrocytes, oligodendrocytes, and neurons [154-158]. Neuronal communications with glial cells are mediated via EVs by the transport of mRNAs, miRNAs, and proteins, where vesicles' discharge into the extracellular space is taken up through recipient cells [154,159-161]. Synaptic pruning was performed through neuronal EVs via neuron-specific signal transduction between microglia and neurons; it was not improved via non-neuronal EVs [162]. Some evidence indicates that synaptic dysfunction is an essential role in the pathophysiology of neurodegenerative disorders. Exosomal miRNAs have also been demonstrated to play a latently neuroprotective function in subsequent ischemic brain injury. Exosomes from multipotent mesenchymal stem cells (MSCs) mediate miR-133b transfer to neurons and astrocytes, which modify gene expression in charge of functional recovery and neurite remodeling after stroke [163]. EVs provide an apparatus of communication not only between glial cells and nerves, but also permitting the interconnection of the CNS with all body systems [151,164]. The pathology of neurodegenerative disorders is a result of intercellular spreading and aggregation of proteins in the brain [165]. In recent years, the decrease of EV production through an nSMase-ceramide pathway resulted in the alleviation of AD in a mouse model of this disease [166].  $\alpha$ -synuclein of the Parkinson's disease gene encapsulated in neuron-derived EVs is present not

only in the membranes, but also in the extracellular space of EVs and are secreted from neurons [167,168].

## Conclusion

This review has addressed the dysregulated synaptic function and plasticity, receptors, molecular signaling cascades, and spine architecture that underlie cognitive deficits and the behaviors associated with other forms of syndromic ASDs. Synapse dysfunction is linked to the pathophysiology of diverse neurodevelopmental disorders such as intellectual disability, schizophrenia, and autism [12,169]. EVs are membrane-bound nanoparticles discharged into the extracellular space through most types of cells. Many CNS cells can release EVs, including exosomes, which may play a key role in the spread of pathogenic agents in various diseases. EVs have been studied extensively in pathologies including neurodegenerative disorders, such as prion protein in prion diseases,  $\alpha$ -synuclein protein in Parkinson's disease, tau and amyloid-ß peptide in AD, mutant huntingtin in Huntington's disease, and superoxide dismutase-1 and transactive response DNA-binding protein 43 kDa (TDP-43) in amyotrophic lateral sclerosis [166,170-175]. MSCs show homing abilities, which make it possible for them to travel to sites of inflammation or brain injury and to be used in treatments of various neurological disorders [176]. It may be difficult for clinicians to detect subtle injuries in the fetal brain. With the recognition of novel vesicle biomarkers, we hopefully will develop the ability to use EVs as a tool in clinical practice for treatment of nervous system diseases in the future.

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#### Declaration

The authors declare that there is no conflict of interest regarding the publication of this paper.

#### References

- 1. Dobbing J, Sands J (1973) Quantitative growth and development of human brain. Arch. Dis. Child., 1973, 48(10): 757-767.
- 2. Donnelly EH, Smith JM, Farfán EB, Ozcan I (2011) Prenatal radiation exposure: background material for counseling pregnant patients following exposure to radiation. Disaster Med Public Health Prep 5(1): 62-68.
- 3. Yang B, Ren BX, Tang FR (2017) Prenatal irradiation-induced brain neuropathology and cognitive impairment. Brain Dev 39(1): 10-22.
- 4. Nelissen E C, van Montfoort A P, Dumoulin JC, Evers JL (2011) Epigenetics and the placenta. Hum. Reprod. Update 17(3): 397-417.
- Safi-Stibler S, Anne Gabory A (2020) Epigenetics and the developmental origins of health and disease: Parental environment signalling to the epigenome, critical time windows and sculpting the adult phenotype. Semin Cell Dev Biol 97: 172-180.

- 6. Nishikawa K, Kinjo AR (2017) Essential role of long non-coding RNAs in de novo chromatin modifications: the genomic address code hypothesis. Biophys Rev 9(2): 73-77.
- Amin V, Harris RA, Onuchic V, Jackson AR, Charnecki T, et al. (2015) Epigenomic footprints across 111 reference epigenomes reveal tissuespecific epigenetic regulation of lincRNAs. Nat Commun 6: 6370.
- Goyal D, Limesand SW, Goyal R (2019) Epigenetic responses and the developmental origins of health and disease. J Endocrinol 242(1): T105-T119.
- Derkach VA, Oh MC, Guire ES, Soderling TR (2007) Regulatory mechanisms of AMPA receptors in synaptic plasticity. Nat Rev Neurosci 8(2): 101-103.
- Forsyth JK, Lewis DA (2017) Mapping the consequences of impaired synaptic plasticity in schizophrenia through development: an integrative model for diverse clinical features. Trends Cogn Sci 21(10): 760-778.
- 11. Rakic P, Bourgeois JP, Eckenhoff MF, Zecevic N, et al. (1986) Goldman-Rakic, P.S., Concurrent overproduction of synapses in diverse regions of the primate cerebral cortex. Science 232(4747): 232-235.
- 12. Zoghbi HY, Bear MF (2012) Synaptic dysfunction in neurodevelopmental disorders associated with autism and intellectual disabilities. Cold Spring Harb. Perspect. Biol 4(3): a009886.
- 13. Pfeiffer BE, Zang T, Wilkerson JR, Taniguchi M, Maksimova MA, et al. (2010) X mental retardation protein is required for synapse elimination by the activity-dependent transcription factor MEF2. Neuron 66(2): 191-197.
- 14. Tsai NP, Wilkerson JR, Guo W, Maksimova MA, DeMartino GN, et al. (2012) Multiple autism-linked genes mediate synapse elimination via proteasomal degradation of a synaptic scaffold PSD-95. Cell 151(7): 1581-1594.
- Yasuda R, Nakahata Y (2018) Plasticity of spine structure: local signaling, translation, and cytoskeletal reorganization. Front Synaptic Neurosci 10: 29.
- 16. Jędrzejewska-Szmek J, Blackwell KT (2019) From membrane receptors to protein synthesis and actin cytoskeleton: mechanisms underlying long lasting forms of synaptic plasticity. Semin. Cell Dev Biol 95: 120-129.
- 17. Reiner A, Levitz J (2018) Glutamatergic signaling in the central nervous system: ionotropic and metabotropic receptors in concert. Neuron 98(6): 1080-1098.
- Di Paolo G, De Camilli P (2006) Phosphoinositides in cell regulation and membrane dynamics. Nature 443(7112): 651-657.
- 19. Phillips R, Ursell T, Wiggins P, Sens P (2009) Emerging roles for lipids in shaping membrane-protein function. Nature 459(7245): 379-385.
- Miller, RH (1996) Oligodendrocyte origins. Trends Neurosci 19(3): 92-96.
- 21. Reynolds R, Hardy R (1997) Oligodendroglial progenitors labeled with the O4 antibody persist in the adult rat cerebral cortex *in vivo*. J Neurosci Res 47(5): 455-470.
- 22. Raff M C (1989) Glial cell diversification in the rat optic nerve. Science, 1989, 243 (4897): 1450-1455.
- 23. Lee YA (2017) White matter injury of prematurity: its mechanisms and clinical features. J Pathol. Transl Med 51(5): 449-455.
- 24. Jakovcevski I, Filipovic R, Mo Z, Rakic S, Zecevic N, et al. (2009) Oligodendrocyte development and the onset of myelination in the human fetal brain. Front Neuroanat 3: 5.
- 25. Tolcos M, Petratos S, Hirst JJ, Wong F, Spencer SJ, et al. (2017) Blocked, delayed, or obstructed: What causes poor white matter development in intrauterine growth restricted infants? Prog Neurobiol 154: 62-77.

- 26. Sampaio RC, Truwit CL (2001) Myelination in the developing human brain. In: Handbook of Developmental Cognitive Neuroscience; Monica Luciana Collins and Charles A. Nelson, Eds.; MIT Press: Cambridge, MA, 35-44.
- 27. Xu G, Broadbelt KG, Haynes RL, Folkerth RD, Borenstein NS, et al. (2011) Late development of the GABAergic system in the human cerebral cortex and white matter. J Neuropathol Exp Neurol 70(10): 841-858.
- Burd I, Balakrishnan B, Kannan S (2012) Models of fetal brain injury, intrauterine inflammation, and preterm birth. Am J Reprod Immunol 67(4): 287-294.
- 29. Cordeiro CN, Tsimis M, Burd I (2015) Infections and brain development. Obstet Gynecol. Surv., 2015, 70(10): 644.
- 30. Meyer U, Feldon J, Schedlowski M, Yee BK (2006) Immunological stress at the maternal–foetal interface: a link between neurodevelopment and adult psychopathology. Brain Behav Immun 20(4): 378-388.
- 31. Fancy SP, Harrington EP, Baranzini SE Silbereis JC, Shiow LR, Yuen TJ, et al. (2014) Parallel states of pathological Wnt signaling in neonatal brain injury and colon cancer. Nat Neurosci 17(4): 506-512.
- 32. Fancy SP, Baranzini SE, Zhao C, Yuk DI, Irvine KA, et al. (2009) Dysregulation of the Wnt pathway inhibits timely myelination and remyelination in the mammalian CNS. Genes Dev 23(13): 1571-1585.
- Brand MD, Nicholls DG (2011) Assessing mitochondrial dysfunction in cells. Biochem J 435(2): 297-312.
- 34. Schoenfeld R, Wong A, Silva J, Li M, Itoh A, et al. (2010) Oligodendroglial differentiation induces mitochondrial genes and inhibition of mitochondrial function represses oligodendroglial differentiation. Mitochondrion 10(2): 143-150.
- 35. Kirkinezos IG, Moraes CT (2001) Reactive oxygen species and mitochondrial diseases. Semin Cell Dev Biol 12(6): 449-457.
- 36. Pang Y, Campbell L, Zheng B, Fan L, Cai Z, et al. (2010) Lipopolysaccharideactivated microglia induce death of oligodendrocyte progenitor cells and impede their development. Neuroscience 166(2): 464-475.
- 37. Kaur C, Sivakumar V, Ang LS, Sundaresan A (2006) Hypoxic damage to the periventricular white matter in neonatal brain: role of vascular endothelial growth factor, nitric oxide and excitotoxicity. J Neurochem. 98(4): 1200-1216.
- Matute C, Alberdi E, Domercq M, Sánchez-Gómez MV, Pérez-Samartín A, et al. (2007) Excitotoxic damage to white matter. J Anat 210(6): 693-702.
- 39. Back SA, Tuohy TM, Chen H, Wallingford N, Craig A, et al. (2005) Hyaluronan accumulates in demyelinated lesions and inhibits oligodendrocyte progenitor maturation. Nat Med 11(9): 966-972.
- 40. MacAskill AF, Atkin TA, Kittler JT (2010) Mitochondrial trafficking and the provision of energy and calcium buffering at excitatory synapses. Eur J Neurosci 32(2): 231-240.
- 41. Rousset CI, Baburamani AA, Thornton C, Hagberg H (2012) Mitochondria and perinatal brain injury. J Matern Fetal Neonatal Med 25 Suppl 1: 35-38.
- 42. Parolini O, Alviano F, Bagnara GP, Bilic G, Bühring HJ, et al. (2008) Concise review: isolation and characterization of cells from human term placenta: outcome of the first international Workshop on Placenta Derived Stem Cells. Stem Cells 26(2): 300-311.
- 43. Pischiutta F, Sammali E, Parolini O, Carswell HV O, Zanier ER, et al. (2018) Placenta-derived cells for acute brain injury. Cell Transplant 27(1): 151-167.
- 44. Jauniaux E, Watson AL, Hempstock J, Bao YP, Skepper JN, et al. (2000) Onset of maternal arterial blood flow and placental oxidative stress. Am J Pathol 157(6): 2111-2122.

- 45. Burton GJ, Jauniaux E, Watson AL (1999) Maternal arterial connections to the placental intervillous space during the first trimester of human pregnancy: the Boyd Collection revisited. Am J Obstet Gynecol 181(3): 718-724.
- 46. Turner JM, Mitchell MD, Kumar SS (2020) The physiology of intrapartum fetal compromise at term. Am J Obstet Gynecol 222(1): 17-26.
- 47. Savasan ZA, Goncalves LF, Bahado-Singh RO (2014) Second- and thirdtrimester biochemical and ultrasound markers predictive of ischemic placental disease. Semin Perinatol 38(3): 167-176.
- 48. Adamsons K, Mueller-Heubach E, Myers RE (1971) Production of fetal asphyxia in the rhesus monkey by administration of catecholamines to the mother. Am J Obstet Gynecol 109(2): 248-262.
- 49. Fisher SJ (2004) The placental problem: linking abnormal cytotrophoblast differentiation to the maternal symptoms of preeclampsia. Reprod Biol Endocrinol 2(1): 53.
- 50. Fisher SJ (2015) Why is placentation abnormal in preeclampsia? Am J Obstet Gynecol 213(4): S115-S122.
- Ananth CV (2014) Ischemic placental disease: a unifying concept for preeclampsia, intrauterine growth restriction, and placental abruption. Semin Perinatol 8(3): 131-132.
- 52. Maltepe E, Fisher SJ (2015) Placenta: the forgotten organ. Annu Rev Cell Dev Biol 31: 523-552.
- 53. Williams MA, Lieberman E, Mittendorf R, Monson RR, Schoenbaum SC, et al. (1991) Risk factors for abruptio placentae. Am J Epidemiol 134(9): 965-972.
- 54. Ananth CV, Smulian JC, Vintzileos AM (1999) Incidence of placental abruption in relation to cigarette smoking and hypertensive disorders during pregnancy: a meta-analysis of observational studies. Obstet Gynecol 93(4): 622-628.
- Redline RW (2005) Severe fetal placental vascular lesions in term infants with neurologic impairment. Am. J. Obstet. Gynecol., 2005, 192(2): 452-457.
- 56. Kratimenos P, Penn AA (2019) Placental programming of neuropsychiatric disease. Pediatr. Res., 2019, 86(2): 157-164.
- 57. Khwaja O, Volpe JJ (2008) Pathogenesis of cerebral white matter injury of prematurity. Arch. Dis. Child. Fetal Neonatal Ed., 2008, 93(2): F153-F161.
- Malaeb S, Dammann O (2009) Fetal inflammatory response and brain injury in the preterm newborn. J. Child Neurol., 2009, 24(9): 1119-1126.
- 59. Hagberg H, Mallard C, Ferriero DM, Vannucci SJ, Levison SW, et al. (2015) The role of inflammation in perinatal brain injury. Nat Rev Neurol 11(4): 192-208.
- 60. Bona E, Andersson AL, Blomgren K, Gilland E, Puka-Sundvall M, et al. (1999) Chemokine and inflammatory cell response to hypoxia-ischemia in immature rats. Ped Res 45: 500-509.
- 61. Ferriero DM, Holtzman DM, Black SM, Sheldon RA (1996) Neonatal mice lacking neuronal nitric oxide synthase are less vulnerable to hypoxicischemic injury. Neurobiol Dis 3(1): 64-71.
- 62. Ye P, D'ercole A (1999) Insulin-like growth factor I protects oligodendrocytes from tumor necrosis factor-α-induced injury. Endocrinology 140(7): 3063-3072.
- 63. Deng Y, Xie D, Fang M, Zhu G, Chen C, et al. (2014) Astrocytederived proinflammatory cytokines induce hypomyelination in the periventricular white matter in the hypoxic neonatal brain. PLos One 9(1): e87420.
- 64. Hagberg H, Mallard C (2000) Antenatal brain injury: aetiology and possibilities of prevention. Semin. Neonatol 5(1): 41-51.

- 65. Tita ATN, Andrews WW (2010) Diagnosis and management of clinical chorioamnionitis. Clin Perinatol 37(2): 339-354.
- 66. Glass HC, Pham TN, Danielsen B, Towner D, Glidden D, et al. (2009) Antenatal and intrapartum risk factors for seizures in term newborns: a population-based study, California 1998-2002. J Pediatr 154(1): 24-28. e1.
- 67. Chau V, Poskitt KJ, McFadden DE, Bowen-Roberts T, Synnes A, et al. (2009) Effect of chorioamnionitis on brain development and injury in premature newborns. Ann Neurol 66(2): 155-164.
- 68. Romero R, Mazor M (1988) Infection and preterm labor. Obstet Gynecol 31: 553-584.
- 69. Wu YW, Colford Jr JM (2000) Chorioamnionitis as a risk factor for cerebral palsy: a meta-analysis. JAMA 2000, 284(11): 1417-1424.
- 70. Bashiri A, Burstein E, Mazor M (2006) Cerebral palsy and fetal inflammatory response syndrome: a review. J Perinat Med 34(1): 5-12.
- 71. Cai Z, Lin S, Pang Y, Rhodes PG (2004) Brain injury induced by intracerebral injection of interleukin-1beta and tumor necrosis factoralpha in the neonatal rat. Pediatr Res 56(3): 377-384.
- 72. Jurewicz A, Matysiak M, Tybor K, Kilianek L, Raine CS, et al. (2005) Tumour necrosis factor-induced death of adult human oligodendrocytes is mediated by apoptosis inducing factor. Brain 128(11): 2675-2688.
- 73. Boyer K, Marcinak J, McLeod R (2007) Toxoplasma gondii (toxoplasmosis). In: Principles and Practice of Pediatric Infectious Diseases. (3<sup>rd</sup> edn.). In: Long S; Pickering LK; Prober CG (Eds.), Churchill Livingstone: New York, section 274.
- 74. Hermes G, Ajioka JW, Kelly KA, Mui E, Roberts F, et al. (2008) Neurological and behavioral abnormalities, ventricular dilatation, altered cellular functions, inflammation, and neuronal injury in brains of mice due to common, persistent, parasitic infection. J Neuroinflammation 5: 48.
- 75. Haroon F, Händel U, Angenstein F, Goldschmidt J, Kreutzmann P, et al. (2012) Toxoplasma gondii actively inhibits neuronal function in chronically infected mice. PLoS One 7(4): e35516.
- Plotkin SA (2006) The history of rubella and rubella vaccination leading to elimination. Clin. Infect. Dis., 2006, 43 (Supplement\_3): S164-S168.
- 77. Webster WS (1998) Teratogen update: Congenital rubella. Teratology 1998, 58(1): 13-23.
- 78. Gershon AA, John Bennett. Raphael Dolin. Martin J, Blaser, et al. (2014) Rubella virus (German Measles). In: Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases. (8<sup>th</sup> edn.). Elsevier Health Sciences: Amsterdam, The Netherlands.
- 79. Lambert N, Strebel P, Orenstein W, Icenogle J, Poland GA, et al. (2015) Rubella. Lancet 385(9984): 2297-2307.
- Desmond MM, Wilson S, Melnick JL, Singer DB, Zion TE, et al. (1967) Congenital rubella encephalitis. Course and Early Sequelae. J Pediatr 71(3): 311-331.
- 81. Desmond MM, Montgomery JR, Melnick JL, Cochran GG, Verniaud W, et al. (1969) Congenital rubella encephalitis. Effects on growth and early development. Am J Dis Child 118(1): 30-31.
- 82. Ogawa H, Suzutani T, Baba Y, Koyano S, Nozawa N, et al. (2007) Etiology of severe sensorineural hearing loss in children: independent impact of congenital cytomegalovirus infection and GJB2 mutations. J Infect Dis 195(6): 782-788.
- 83. Miranda-Saksena M, Boadle RA, Aggarwal A, Tijono B, Rixon F J, et al. (2009) Herpes simplex virus utilizes the large secretory vesicle pathway for anterograde transport of tegument and envelope proteins and for viral exocytosis from growth cones of human fetal axons. J Virol 83(7): 3187-3199.

- 84. Liu ST, Sharon-Friling R, Ivanova P, Milne SB, Myers DS, et al. (2011) Synaptic vesicle-like lipidome of human cytomegalovirus virions reveals a role for SNARE machinery in virion egress. Proc Natl Acad Sci U S A 108(31): 12869-12874.
- 85. Stamos JK, Rowley AH (1994) Timely diagnosis of congenital infections. Pediatr Clin North Am 41(5): 1017-1033.
- 86. Piacentini R, Li Puma DD, Ripoli C, Marcocci ME, De Chiara G, et al. (2015) Herpes simplex virus type-1 infection induces synaptic dysfunction in cultured cortical neurons via GSK-3 activation and intraneuronal amyloid-beta protein accumulation. Sci Rep 5: 15444.
- 87. Kraśnianin E, Skret-Magierło J, Witalis J, Barnaś E, Kluz T, et al. (2009) The incidence of streptococcus group B in 100 parturient women and the transmission of pathogens to the newborn. Ginekol Pol 80 (4).
- 88. Edwards MS, Rench MA, Haffar AA, Murphy MA, Desmond MM, et al. (1985) Long-term sequelae of group B streptococcal meningitis in infants. J Pediatr 106(5): 717-722.
- 89. Kim YS, Sheldon RA, Elliott BR, Liu Q, Ferriero DM, et al. (1995) Brain injury in experimental neonatal meningitis due to group B streptococci. J Neuropathol. Exp Neurol 54(4): 531-539.
- 90. Stevenson DK (2006) Cerebral white matter injury in the newborn following Escherichia coli meningitis. Yearbook of Neonatal and Perinatal Medicine pp. 149-152.
- Low JA, Pickersgill H, Killen H, Derrick EJ (2001) The prediction and prevention of intrapartum fetal asphyxia in term pregnancies. Am J Obstet Gynecol 184(4): 724-730.
- 92. Vannucci RC (1990) Experimental biology of cerebral hypoxia-ischemia relation to perinatal brain damage Pediatr Res 27(4): 317.
- 93. Baud 0 (2007) Neurological adverse effects of postnatal steroids in preterm infant. Arch Pediatr 14(6): 596-598.
- 94. Bloom SL, Swindle RG, McIntire DD, Leveno KJ (1999) Fetal pulse oximetry: duration of desaturation and intrapartum outcome. Obstet Gynecol 93(6): 1036-1040.
- 95. Stockwell BR, Angeli JPF, Bayir H, Bush AI, Conrad M, et al. (2017) Ferroptosis: a regulated cell death nexus linking metabolism, redox biology, and disease. Cell 171(2): 273-285.
- 96. Cuypers A, Plusquin M, Remans T, Jozefczak M, Keunen E, et al. (2010) Cadmium stress: an oxidative challenge. Biometals 23(5): 927-940.
- 97. Miller SL, Wallace EM, Walker DW (2012) Antioxidant therapies: a potential role in perinatal medicine. Neuroendocrinology 96(1): 13-23.
- 98. Dinarello CA (1991) Interleukin-1 and interleukin-1 antagonism. Blood 77(8): 1627-1652.
- 99. Sims JE, Smith DE (2010) The IL-1 family: regulators of immunity. Nat Rev Immunol 10(2): 89-102.
- Dinarello CA, Simon A, Van Der Meer JW (2012) Treating inflammation by blocking interleukin-1 in a broad spectrum of diseases. Nat Rev Drug Discov 11(8): 633-652.
- 101. Garlanda C, Dinarello CA, Mantovani A (2013) The interleukin-1 family: back to the future. Immunity, 2013, 39(6): 1003-1018.
- 102. Paton MCB, McDonald CA, Allison BJ, Fahey MC, Jenkin G, et al. (2017) Perinatal brain injury as a consequence of preterm birth and intrauterine inflammation: designing targeted stem cell therapies. Front Neurosci 11: 200.
- 103. Gunn AJ, Bennet L (2009) Fetal hypoxia insults and patterns of brain injury: insights from animal models. Clin Perinatol 36(3): 579-593.
- 104. Choi DW, Rothman SM (1990) The role of glutamate neurotoxicity in hypoxic-ischemic neuronal death. Annu. Rev. Neurosci 13(1): 171-182.

- 105. Roth S, Baudin J, Cady EA, Johal K, Townsend J, et al. (1997) Relation of deranged neonatal cerebral oxidative metabolism with neurodevelopmental outcome and head circumference at 4 years. Dev Med Child Neurol 39(11): 718-725.
- 106. Jensen A, Hohmann M, Kunzel W (1987) Dynamic changes in organ blood flow and oxygen consumption during acute asphyxia in fetal sheep. J Dev Physiol 9(6): 543-559.
- 107. Hofmeijer J, van Putten MJ (2012) Ischemic cerebral damage: an appraisal of synaptic failure. Stroke, 2012, 43(2): 607-615.
- 108. Yawno T, Yan EB, Hirst JJ, Walker DW (2011) Neuroactive steroids induce changes in fetal sheep behavior during normoxic and asphyxic states. Stress 14(1): 13-22.
- 109. Shaw JC, Berry MJ, Dyson RM, Crombie GK, Hirst JJ, et al. (2019) Reduced neurosteroid exposure following preterm birth and its' contribution to neurological impairment: a novel avenue for preventative therapies. Front Physiol 10: 599.
- 110. Jensen FE (2009) Neonatal seizures: an update on mechanisms and management. Clin. Perinatol 36(4): 881-900.
- 111. Khan OI, Zhao Q, Miller F, Holmes G (2010) Interictal spikes in developing rats cause long-standing cognitive deficits. Neurobiol. Dis 39(3): 362-371.
- 112. Isaeva E, Isaev D, Holmes GL (2013) Alteration of synaptic plasticity by neonatal seizures in rat somatosensory cortex. Epilepsy Res 106(1-2): 280-283.
- 113. Allen MC (2008) Neurodevelopmental outcomes of preterm infants. Curr. Opinion Neurol., 2008, 21(2): 123-128.
- 114. Rogers EE, Hintz SR (2016) Early neurodevelopmental outcomes of extremely preterm infants. Semin. Perinatol 40(8): 497-509.
- 115. Leviton A, Gressens P (2007) Neuronal damage accompanies perinatal white-matter damage. Trends Neurosci 30(9): 473-478.
- 116. Ramaswami G, Geschwind DH (2018) Genetics of autism spectrum disorder. In: Handbook of Clinical Neurology; 147: 321-329.
- 117. Waye MM, Cheng HY (2018) Genetics and epigenetics of autism: a Review. Psychiatry Clin. Neurosci., 72(4): 228-244.
- 118. Park HR, Lee JM, Moon HE, Lee DS, Kim BN, et al. (2016) A short review on the current understanding of autism spectrum disorders. Exp. Neurobiol 25(1): 1-13.
- 119. Straughen JK, Misra DP, Divine G, Shah R, Perez G, et al. (2017) The association between placental histopathology and autism spectrum disorder. Placenta 57: 183-188.
- 120. Chang JM, Zeng H, Han R, Chang YM, Shah R, et al. (2017) Autism risk classification using placental chorionic surface vascular network features. BMC Med Inform Decis Mak 17(1): 162.
- 121. Park BY, Misra DP, Moye J, Miller RK, Croen L, et al. (2018) Placental gross shape differences in a high autism risk cohort and the general population. PLoS One 13(8): e0191276.
- 122. Grant SG (2012) Synaptopathies: diseases of the synaptome. Curr Opinion Neurobiol 22(3): 522-529.
- 123. Sarkar A, Marchetto MC, Gage FH (2017) Synaptic activity: an emerging player in schizophrenia. Brain Res 1656: 68-75.
- 124. Lewis DA, Moghaddam B (2006) Cognitive dysfunction in schizophrenia: convergence of γ-aminobutyric acid and glutamate alterations. Arch. Neurol 63(10): 1372-1376.
- 125. Brown AS (2012) Epidemiologic studies of exposure to prenatal infection and risk of schizophrenia and autism. Dev. Neurobiol 72(10): 1272-1276.

- 126. Brown AS, Derkits EJ (2010) Prenatal infection and schizophrenia: a review of epidemiologic and translational studies. Am. J. Psychiatry 167(3): 261-280.
- 127. Fatemi SH, Folsom TD (2009) The neurodevelopmental hypothesis of schizophrenia, revisited. Schizophrenia Bull 35(3): 528-548.
- 128. Fatemi SH, Folsom TD, Rooney RJ, Mori S, Kornfield TE, et al. (2012) The viral theory of schizophrenia revisited: abnormal placental gene expression and structural changes with lack of evidence for H1N1 viral presence in placentae of infected mice or brains of exposed offspring. Neuropharmacology, 2012, 62(3): 1290-1298.
- Cannon M, Jones PB, Murray RM (2002) Obstetric complications and schizophrenia: historical and meta-analytic review. Am. J. Psychiatry 159(7): 1080-1092.
- 130. Stroud LR, Papandonatos GD, Parade SH, Salisbury AL, Phipps MG, et al. (2016) Prenatal major depressive disorder, placenta glucocorticoid and serotonergic signaling, and infant cortisol response. Psychosom. Med 78(9): 979.
- 131. Capron LE, Ramchandani PG, Glover V (2018) Maternal prenatal stress and placental gene expression of NR3C1 and HSD11B2: the effects of maternal ethnicity. Psychoneuroendocrinology 87: 166-172.
- 132. Hirst JJ, Kelleher MA, Walker DW, Palliser HK (2014) Neuroactive steroids in pregnancy: key regulatory and protective roles in the foetal brain. J Steroid Biochem Mol Biol 139: 144-153.
- 133. Bennett GA, Palliser HK, Saxby B, Walker DW, Hirst JJ, et al. (2013) Effects of prenatal stress on fetal neurodevelopment and responses to maternal neurosteroid treatment in guinea pigs. Dev Neurosci 35(5): 416-426.
- 134. Lin SC, Bergles DE (2004) Synaptic signaling between GABAergic interneurons and oligodendrocyte precursor cells in the hippocampus. Nat Neurosci 7(1): 24-32.
- 135. Wang H, Yan Y, Kintner DB, Lytle C, Sun D, et al. (2003) GABA-mediated trophic effect on oligodendrocytes requires Na-K-2Cl cotransport activity. J Neurophysiol 90(2): 1257-1265.
- 136. Olsen J (1992) Cigarette smoking in pregnancy and fetal growth. Does the type of tobacco play a role? Int. J. Epidemiol 21(2): 279-284.
- 137. Fan LW, Bhatt A, Tien LT, Zheng B, Simpson KL, et al. (2015) Exposure to serotonin adversely affects oligodendrocyte development and myelination *in vitro*. J Neurochem 133(4): 532-543.
- Nair S, Salomon C (2018) Extracellular vesicles and their immunomodulatory functions in pregnancy. Sem. Immunopathol 40(5): 425-437.
- 139. Fallen S, Baxter D, Wu X, Kim TK, Shynlova O, et al. (2018) Extracellular vesicle RNAs reflect placenta dysfunction and are a biomarker source for preterm labour. J Cell Mol Med 22(5): 2760-2773.
- 140. Liu H, Kang M, Wang J, Blenkiron C, Lee A, et al. (2018) Estimation of the burden of human placental micro-and nano-vesicles extruded into the maternal blood from 8 to 12 weeks of gestation. Placenta 72: 41-47.
- 141. Blencowe H, Lee AC, Cousens S, Bahalim A, Narwal R, et al. (2013) Preterm birth-associated neurodevelopmental impairment estimates at regional and global levels for 2010. Pediatr Res 74 Suppl 1: 17-34.
- 142. Ball G, Boardman JP, Rueckert D, Aljabar P, Arichi T, et al. (2011) The effect of preterm birth on thalamic and cortical development. Cereb. Cortex 22(5): 1016-1024.
- 143. Inder TE, Huppi PS, Warfield S, Kikinis R, Zientara GP, et al. (1999) Periventricular white matter injury in the premature infant is followed by reduced cerebral cortical gray matter volume at term. Ann. Neurology 46(5): 755-760.

- 144. Beauchamp MH, Thompson DK, Howard K, Doyle LW, Egan GF, et al. (2008) Preterm infant hippocampal volumes correlate with later working memory deficits. Brain 131(11): 2986-2994.
- 145. Brown NC, Inder TE, Bear MJ, Hunt RW, Anderson PJ, et al. (2009) Neurobehavior at term and white and gray matter abnormalities in very preterm infants. J Pediatr 155(1): 32-38. e1.
- 146. Volpe JJ (2003) Cerebral white matter injury of the premature infantmore common than you think. Pediatrics 112(1): 176-180.
- 147. Doyle LW (2006) Evaluation of neonatal intensive care for extremelylow-birth-weight infants. Semin. Fetal Neonatal Med 11(2): 139-145.
- 148. Folkerth RD (2005) Neuropathologic substrate of cerebral palsy. J Child Neurol 20(12): 940-949.
- 149. Phillips AW, Johnston MV, Fatemi A (2013) The potential for cell-based therapy in perinatal brain injuries. Transl. Stroke Res 4(2): 137-148.
- 150. Volpe JJ (2001) Neurobiology of periventricular leukomalacia in the premature infant. Pediatr Res 50(5): 553-562.
- 151. Shaimardanova AA, Solovyeva VV, Chulpanova DS, James V, Kitaeva KV, et al. (2020) Extracellular vesicles in the diagnosis and treatment of central nervous system diseases. Neural Regen Res 15(4): 586-596.
- 152. Hartmann A, Burg G (1989) Fulminant acne in Klinefelter syndrome treated with testosterone. A side effect of anti-tallness therapy. Monatsschr. Kinderheilkd 137(8): 466-467.
- 153. Merchant ML, Rood IM, Deegens JK, Klein JB (2017) Isolation and characterization of urinary extracellular vesicles: implications for biomarker discovery. Nat Rev Nephrol 13(12): 731-749.
- 154. Fruhbeis C, Frohlich D, Kuo WP, Amphornrat J, Thilemann S, et al. (2013) Neurotransmitter-triggered transfer of exosomes mediates oligodendrocyte-neuron communication. PLoS Biol 11(7): e1001604.
- 155. Guescini M, Genedani S, Stocchi V, Agnati LF (2010) Astrocytes and glioblastoma cells release exosomes carrying mtDNA. J Neural Transm (Vienna) 117 (1): 1-4.
- 156. Lachenal G, Pernet-Gallay K, Chivet M, Hemming FJ, Belly A, et al. (2011) Release of exosomes from differentiated neurons and its regulation by synaptic glutamatergic activity. Mol Cell Neurosci 46(2): 409-418.
- 157. Faure J, Lachenal G, Court M, Hirrlinger J, Chatellard-Causse C, et al. (2006) Exosomes are released by cultured cortical neurones. Mol. Cell. Neurosci 31(4): 642-648.
- 158. Potolicchio I, Carven GJ, Xu X, Stipp C, Riese RJ, et al. (2005) Proteomic analysis of microglia-derived exosomes: metabolic role of the aminopeptidase CD13 in neuropeptide catabolism. J Immunol 175(4): 2237-2243.
- 159. Wang S, Cesca F, Loers G, Schweizer M, Buck F, et al. (2011) Synapsin I is an oligomannose-carrying glycoprotein, acts as an oligomannosebinding lectin, and promotes neurite outgrowth and neuronal survival when released via glia-derived exosomes. J. Neurosci 31(20): 7275-7290.
- 160. Morel L, Regan M, Higashimori H, Ng SK, Esau C, et al. (2013) Neuronal exosomal miRNA-dependent translational regulation of astroglial glutamate transporter GLT1. J Biol Chem 288(10): 7105-7116.

- 161. Frühbeis C, Fröhlich D, Kuo WP, Krämer-Albers (2013) EM Extracellular vesicles as mediators of neuron-glia communication. Front Cell Neurosci 7: 182.
- 162. Schafer DP, Lehrman EK, Kautzman AG, Koyama R, Mardinly AR, et al. (2012) Microglia sculpt postnatal neural circuits in an activity and complement-dependent manner. Neuron 74(4): 691-705.
- 163. Xin H, Li Y, Liu Z, Wang X, Shang X, Cui Y, et al. (2013) MiR-133b promotes neural plasticity and functional recovery after treatment of stroke with multipotent mesenchymal stromal cells in rats via transfer of exosome-enriched extracellular particles. Stem Cells 31(12): 2737-2746.
- 164. Chivet M, Hemming F, Fraboulet S, Sadoul R (2012) Emerging role of neuronal exosomes in the central nervous system. Front Physiol 3: 145.
- 165. Carbonell F, Iturria-Medina Y, Evans AC (2018) Mathematical modeling of protein misfolding mechanisms in neurological diseases: a historical overview. Front Neurol 9: 37.
- 166. Fevrier B, Villette D, Archer F, Loews D, Daigle W, et al. (2004) Cells release prions in association with exosomes. Proc Natl Acad Sci U S A 101(26): 9683-9688.
- 167. Grey M, Dunning CJ, Gaspar R, Grey C, Brundin P, et al. (2015) Acceleration of alpha-synuclein aggregation by exosomes. J Biol Chem 290(5): 2969-2982.
- 168. Danzer KM, Kranich LR, Ruf WP, Cagsal-Getkin O, Winslow AR, et al. (2012) Exosomal cell-to-cell transmission of alpha synuclein oligomers. Mol Neurodegener 7: 42.
- 169. Volk L, Chiu SL, Sharma K, Huganir RL (2015) Glutamate synapses in human cognitive disorders. Annu Rev Neurosci 38: 127-149.
- 170. Quek C, Hill AF (2017) The role of extracellular vesicles in neurodegenerative diseases. Biochem Biophys Res Commun 483(4): 1178-1186.
- 171. Rajendran L, Bali J, Barr MM, Court FA, Kramer-Albers EM, et al. (2014) Emerging roles of extracellular vesicles in the nervous system. J Neurosci 34(46): 15482-15489.
- 172. Emmanouilidou E, Melachroinou K, Roumeliotis T, Garbis SD, Ntzoun M, et al. (2010) Cell-produced alpha-synuclein is secreted in a calciumdependent manner by exosomes and impacts neuronal survival. J Neurosci 30(20): 6838-6851.
- 173. Nonaka T, Masuda-Suzukake, M, Arai T, Hasegawa Y, Akatsu H, et al. (2013) Prion-like properties of pathological TDP-43 aggregates from diseased brains. Cell Rep 4(1): 124-134.
- 174. Rajendran L, Honsho M, Zahn TR, Keller P, Geiger KD, et al. (2006) Alzheimer's disease  $\beta$ -amyloid peptides are released in association with exosomes. Proc Natl Acad Sci U S A 103(30): 11172-11177.
- 175. Saman S, Kim W, Raya M, Visnick Y, Miro S, et al. (2012) Exosomeassociated tau is secreted in tauopathy models and is selectively phosphorylated in cerebrospinal fluid in early Alzheimer disease. J Biol Chem 287(6): 3842-8439.
- 176. Lee HK, Finniss S, Cazacu S, Xiang C, Brodie C, et al. (2014) Mesenchymal stem cells deliver exogenous miRNAs to neural cells and induce their differentiation and glutamate transporter expression. Stem Cells Dev 23(23): 2851-2861.

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